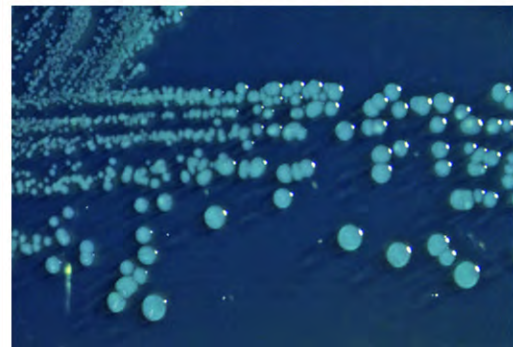
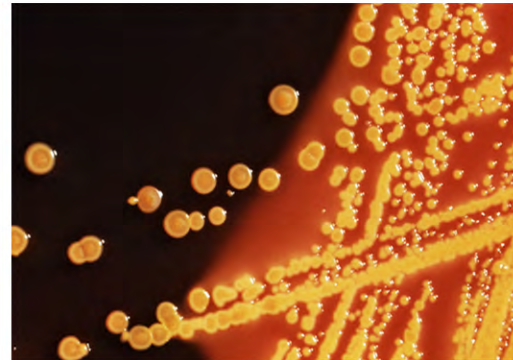


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# **National Antimicrobial Resistance Monitoring System: Enteric Bacteria**

## **2009**

### **Human Isolates Final Report**



National Center for Emerging and Zoonotic Infectious Diseases  
Division of Foodborne, Waterborne, and Environmental Diseases



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## List of Abbreviations and Acronyms

ACSSuT	Resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, and tetracycline
ACSSuTAuCx	Resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline, amoxicillin-clavulanic acid, and ceftriaxone
ACT/S	Resistance to at least ampicillin, chloramphenicol, and trimethoprim-sulfamethoxazole
ANT/S	Resistance to at least ampicillin, nalidixic acid and trimethoprim-sulfamethoxazole
AT/S	Resistance to at least ampicillin and trimethoprim-sulfamethoxazole
CDC	Centers for Disease Control and Prevention
CI	Confidence interval
CLSI	Clinical and Laboratory Standards Institute
EIP	Emerging Infections Program
ELC	Epidemiology and Laboratory Capacity
FDA-CVM	Food and Drug Administration-Center for Veterinary Medicine
FoodNet	Foodborne Diseases Active Surveillance Network
MIC	Minimum inhibitory concentration
NARMS	National Antimicrobial Resistance Monitoring System for Enteric Bacteria
OR	Odds ratio
PHLIS	Public Health Laboratory Information System
USDA	United States Department of Agriculture
WHO	World Health Organization



## NARMS Working Group

### **Centers for Disease Control and Prevention**

#### *Enteric Diseases Epidemiology Branch*

#### *Enteric Diseases Laboratory Branch*

#### *Division of Foodborne, Waterborne and Environmental Diseases*

#### *National Center for Emerging and Zoonotic Infectious Diseases*

Olympia Anderson  
Frederick Angulo  
Ezra Barzilay  
Jason Folster  
Peter Gerner-Smith  
Audrey Green  
Patricia Griffin  
Robert Michael Hoekstra  
Rebecca Howie  
Kevin Joyce  
Maria Karlsson  
Beth Karp  
Amy Krueger  
Kathryn Lupoli  
Andre McCullough  
Felicita Medalla  
Gary Pecic  
Jared Reynolds  
Regan Rickert  
Jacinta Smith  
Andrew Stuart  
Robert Tauxe  
Jean Whichard

### **U.S. Food and Drug Administration**

#### *Center for Veterinary Medicine*

Heather Green  
Claudine Kabera  
Patrick McDermott  
Emily Tong  
Niketia Womack

### **Participating State and Local Health Departments**

#### *Alabama Department of Public Health*

LaDonna Cranidiotis  
Sherri Davidson  
Sharon Massingale  
Patricia Morrow  
Joanna Roberson

#### *Alaska Department of Health and Social Services*

Shellie Smith  
Catherine Xavier

#### *Arizona Department of Health Services*

Shoana Anderson  
Aarikha D'Souza  
Daniel Flood  
Melissa Hoffman  
Ken Komatsu  
William Slanta  
Victor Waddell

#### *Arkansas Department of Health*

Rossina Stefanova

#### *California Department of Health Services*

Wendy Cheung  
Claudia Crandall  
Samar Fontanoz  
Paul Kimsey  
Will Probert  
Sam Shin  
Duc Vugia

#### *Colorado Department of Public Health and Environment*

Alicia Cronquist  
Laura Gillim-Ross  
Joyce Knutsen  
Hugh Maguire

#### *Connecticut Department of Public Health*

Diane Barden  
Sharon Hurd  
Aristea Kinney  
Mona Mandour

#### *Delaware Health and Social Services*

Gaile McLaughlin  
Bela Patel  
Debra Rutledge

#### *Florida Department of Health*

Ronald Baker  
Maria Calcaterra  
Sonia Etheridge  
Dian Sharma

#### *Georgia Division of Public Health*

Jim Benson  
Elizabeth Franko  
Tameka Hayes  
Mary Hodel  
Susan Lance  
Bob Manning  
Mahin Park  
Lynett Poventud  
Suzanne Segler  
Stepy Thomas  
Melissa Tobin-D'Angelo

#### *Hawaii Department of Health*

Rebecca Kanenaka  
Norman O'Connor

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Raouf Arafat  
Adebowale Awosika-Olumo  
Gregory Dufour  
Vern Juchau  
Sudha Pottumarthy  
Joan Rogers

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Colleen Greenwalt  
Vivian Lockary  
Raemi Nolevanko

#### *Illinois Department of Public Health*

Nancy Barstead  
Bob Cox  
Mark Dworkin  
Juan Garcia  
Rebecca Hambelton  
Stephen Hendren  
Steve Hopkins  
Patrick Miller  
Mohammad Nasir  
Kiran Patel  
Tricia Patterson  
Guinevere Reserva  
Bindu Shah  
Andrea Stadsholt

*Indiana State Department of Health*

Brent Barrett  
Amie May  
John Radosevic

*Iowa Department of Public Health, University Hygienic Laboratory*

Mary DeMartino  
Randy Groepper

*Kansas Department of Health and Environment*

Cheryl Banez-Ocfemia  
Robert Flahart  
Gail Hansen  
Carissa Pursell  
June Sexton  
Kathleen Waters

*Kentucky Department of Public Health*

Robin Cotton  
Karim George  
William Grooms  
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Sheena Chu  
Sue Sabet  
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Roshan Reporter  
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Erin Delaune  
Wayne Dupree  
Catrin Jones-Nazar  
Lori Kravet  
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Raoult Ratard  
Theresa Sokol  
Susanne Straif-Bourgeois

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Jeff Randolph  
Vicki Rea  
Lori Webber  
Donna Wrigley  
Anthony Yartel

*Maryland Department of Health and Mental Hygiene*

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Kirsten Larson  
Celere Leonard  
Amanda Palmer  
Jafar Razeq  
Pat Ryan

*Massachusetts Department of Public Health*

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Robert Goldbaum  
Emily Harvey  
Patricia Kludt  
Joseph Peppe  
Tracy Stiles

*Michigan Department of Community Health*

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Frances Downes  
Teri Lee Dyke  
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Fe Leano  
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Kirk Smith  
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Jane Campbell  
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Sheryl Hand  
Cathie Hoover  
Daphne Ware

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Steve Gladbach  
Jason Herstein  
Harvey Marx  
JoAnn Rudroff

*Montana Department of Public Health and Human Services*

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Anne Weber  
Susanne Zanto

*Nebraska Health and Human Services and the Nebraska Public Health Laboratory*

Amy Armbrust  
Jude Dean  
Paul Fey  
Peter Iwen  
Tom Safranek

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Patricia Armour  
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Jaime Frank  
Paul Hug  
Bradford Lee  
Susanne Quianzon  
Lisa Southern  
Stephanie Van Hooser

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Christine Bean  
Elizabeth Daly  
Wendy Lamothe  
Nancy Taylor  
Daniel Tullo

*New Jersey Department of Health*

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Michelle Malavet  
Sylvia Matiuck  
Paul Seitz

*New Mexico Department of Health*

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Lisa Onischuk  
Erica Pierce  
Paul Torres

*New York City Department of Health*

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Lillian Lee  
Jennifer Rakeman  
Vasudha Reddy

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Dale Morse  
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Lisa Well

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Paula Bailey  
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*Texas Department of State Health Services*

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Elizabeth Delamater  
Linda Gaul  
Eldridge Hutcheson  
Miriam Johnson  
Susan Neill  
Pushker Raj  
Ana Valle

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Cindy Fisher  
David Jackson  
Barbara Jepson  
Susan Mottice

*Vermont Department of Health*

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Valerie Cook  
Eunice H. Froeliger  
Christine LaBarre

*Virginia Division of Consolidated Laboratory Services and Virginia Department of Health*

Ellen Basinger  
Sherry Giese  
Jody Lowman  
Mary Mismas  
Denise Toney

*Washington Department of Health*

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Romesh Gautom  
Donna Green  
Brian Hiatt  
Yolanda Houze  
Kathryn MacDonald

*West Virginia Department of Health and Human Resources*

Danae Bixler  
Christi Clark  
Maria del Rosario  
Loretta Haddy  
Andrea Labik  
Megan Young

*Wisconsin Department of Health and Family Services*

John Archer  
Susann Ahrabi-Fard  
Charles Brokopp  
Jeffrey Davis  
Rick Heffernan  
Rachel Klos  
Tim Monson  
Dave Warshauer

*Wyoming Department of Health*

Richard Harris

John Harrison

Clay Van Houten

Tracy Murphy

Jim Walford

### **Mutually exclusive criteria heading**

In Box 2 (page 47), we describe the 4 most common multidrug-resistant (MDR) patterns among non-typhoidal *Salmonella* isolates based on resistance to 7 of the 15 agents currently tested in NARMS: ampicillin (A), chloramphenicol (C), streptomycin (S), sulfonamide (Su), tetracycline (T), amoxicillin-clavulanic acid (Au), and ceftriaxone (Cx). Resistance to the 7 agents has been used in NARMS to categorize specific MDR patterns. Unlike MDR criteria used for tables in previous reports and other sections of this report, we used mutually exclusive criteria in the new section. Use of mutually exclusive criteria is important in monitoring major and emerging patterns, which may be driven by different resistance mechanisms.

### **Update of trimethoprim-sulfamethoxazole data for *Shigella***

Automated fluorescence-based methods have been used since 2001 to determine minimum inhibitory concentrations (MIC) for the drugs tested for *Enterobacteriaceae*. These automated fluorescence-based methods are designed to emulate MICs that would be obtained if the results were read visually. Recent laboratory comparison studies showed that the automated fluorescence-based method was not reproducibly emulating visually-determined results for trimethoprim-sulfamethoxazole with *Shigella* species. The test manufacturer has updated the automated fluorescence-based method to improve concordance with visual results, and these updates have been applied retroactively to the affected data in the database. This has resulted in lower MIC results and lower prevalence of trimethoprim-sulfamethoxazole resistance for *Shigella*.

## Introduction

The National Antimicrobial Resistance Monitoring System (NARMS) for Enteric Bacteria is a collaboration among the Centers for Disease Control and Prevention (CDC), [U.S. Food and Drug Administration's Center for Veterinary Medicine](#) (FDA-CVM), and [U.S. Department of Agriculture](#) (USDA). The primary purpose of NARMS at CDC is to monitor antimicrobial resistance among enteric bacteria isolated from humans. Other components of the interagency NARMS program include surveillance for resistance in enteric bacteria isolated from foods, conducted by the FDA-CVM (<http://www.fda.gov/AnimalVeterinary/SafetyHealth/AntimicrobialResistance/NationalAntimicrobialResistanceMonitoringSystem/default.htm>), and resistance in enteric bacteria isolated from animals, conducted by the USDA Agricultural Research Service ([http://www.ars.usda.gov/main/site\\_main.htm?modecode=66-12-05-08](http://www.ars.usda.gov/main/site_main.htm?modecode=66-12-05-08)).

Many NARMS activities are conducted within the framework of CDC's Emerging Infections Program (EIP), Epidemiology and Laboratory Capacity (ELC) Program, and the Foodborne Diseases Active Surveillance Network (FoodNet). In addition to surveillance of resistance in enteric pathogens, the NARMS program at CDC also includes public health research into the mechanisms of resistance, education efforts to promote prudent use of antimicrobial agents, and studies of resistance in commensal organisms.

Before NARMS was established, CDC monitored antimicrobial resistance in *Salmonella*, *Shigella*, and *Campylobacter* through periodic surveys of isolates from a panel of sentinel counties. NARMS at CDC began in 1996 with prospective monitoring of antimicrobial resistance among clinical non-typhoidal *Salmonella* and *Escherichia coli* O157 isolates in 14 sites. In 1997, testing of clinical *Campylobacter* isolates was initiated in the five sites participating in FoodNet. Testing of clinical *Salmonella enterica* serotype Typhi and *Shigella* isolates was added in 1999. Since 2003, all 50 states have been forwarding a representative sample of non-typhoidal *Salmonella*, *Salmonella* ser. Typhi, *Shigella*, and *E. coli* O157 isolates to NARMS for antimicrobial susceptibility testing, and 10 FoodNet states have been participating in *Campylobacter* surveillance. Since 2008, all 50 states have been forwarding every *Salmonella* Paratyphi A and C to NARMS for antimicrobial susceptibility testing.

This annual report includes CDC's surveillance data for 2009 for non-typhoidal *Salmonella*, typhoidal *Salmonella*, *Shigella*, *Campylobacter* and *E. coli* O157 isolates. Data for earlier years are presented in tables and graphs when appropriate. Antimicrobial classes defined by Clinical and Laboratory Standards Institute (CLSI) are used in data presentation and analysis. CLSI classes constitute major classifications of antimicrobial agents, e.g., aminoglycosides and cepheems.

This report also includes the World Health Organization's categorization of antimicrobials of critical importance to human medicine ([Table 1](#)). The table includes only antimicrobials that are tested in NARMS.

Additional NARMS data and more information about NARMS activities are available at <http://www.cdc.gov/narms>

## WHO Categorization of Antimicrobial Agents

In 2007, the World Health Organization (WHO) convened for the second time a panel of experts to develop a list of essential antimicrobial agents according to their importance to human medicine (WHO, 2007). The participants categorized antimicrobial agents as either Critically Important, Highly Important, or Important based upon two criteria: (1) sole therapies or one of the few alternatives to treat serious human diseases and (2) used to treat disease caused by organisms that may be transmitted via non-human sources or diseases caused by organisms that may acquire resistance genes from non-human sources. Antimicrobial agents tested in NARMS have been included in the WHO categorization table.

- Antimicrobial agents are critically important if both criteria (1) and (2) are true.
- Antimicrobial agents are highly important if either criterion (1) or (2) is true.
- Antimicrobial agents are important if neither criterion is true.

**Table 1. WHO categorization of antimicrobials of critical importance to human medicine**

WHO Category Level	Importance	CLSI Class	Antimicrobial Agent tested in NARMS
<b>I</b>	<b>Critically important</b>	Aminoglycosides	Amikacin
			Gentamicin
			Streptomycin
		β-lactam / β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid
			Cephems
		Ketolides	Ceftriaxone
			Telithromycin
		Macrolides	Azithromycin
			Erythromycin
Penicillins	Ampicillin		
	Quinolones	Ciprofloxacin	
			Nalidixic acid
<b>II</b>	<b>Highly important</b>	Aminoglycosides	Kanamycin
		Cephems	Cefoxitin
			Cephalothin
		Folate pathway inhibitors	Sulfamethoxazole / Sulfisoxazole
			Trimethoprim-sulfamethoxazole
		Phenicols	Chloramphenicol
Tetracyclines*	Tetracycline		
<b>III</b>	<b>Important</b>	Lincosamides	Clindamycin

\*In 2010, WHO recategorized tetracycline from highly important to critically important. The NARMS 2010 annual report will reflect this change.

### Population

In 2009, all 50 states participated in NARMS, representing the entire U.S. population of approximately 307 million persons ([Table 2](#)). Surveillance was conducted in all states for non-typhoidal *Salmonella*, typhoidal *Salmonella*, *Shigella*, and *Escherichia coli* O157. For *Campylobacter*, surveillance was conducted in 10 states that comprise the Foodborne Diseases Active Surveillance Network (FoodNet), representing approximately 47 million persons (15% of the U.S. population).

### Clinically Important Antimicrobial Resistance Patterns

In the United States, fluoroquinolones (e.g., ciprofloxacin) and third-generation cephalosporins (e.g., ceftriaxone) are commonly used to treat severe *Salmonella* infections, including *Salmonella* ser. Typhi, the organism that causes typhoid fever. In *Enterobacteriaceae*, resistance to nalidixic acid, an elementary quinolone, correlates with decreased susceptibility to ciprofloxacin (MIC  $\geq 0.12$   $\mu\text{g/mL}$ ) and possible fluoroquinolone treatment failure. A substantial proportion of *Enterobacteriaceae* isolates tested in 2009 demonstrated resistance to these clinically important antimicrobial agents.

Among non-typhoidal *Salmonella* isolates:

- 1.8% (39/2192) were resistant to nalidixic acid. The most common serotypes among the nalidixic acid-resistant isolates were Enteritidis (15/39, 38%) and Typhimurium (8/39, 21%)
  - 3.7% (15/410) of *Salmonella* ser. Enteritidis isolates were nalidixic acid resistant
  - 2.2% (8/371) of *Salmonella* ser. Typhimurium isolates were nalidixic acid resistant
- 3.4% (75/2192) were resistant to ceftriaxone. The most common serotypes among the ceftriaxone resistant isolates were Typhimurium (24/75, 32%) and Heidelberg (18/75, 24%)
  - 21% (18/86) of *Salmonella* ser. Heidelberg isolates were ceftriaxone resistant
  - 6.5% (24/371) of *Salmonella* ser. Typhimurium isolates were ceftriaxone resistant

Among *Salmonella* ser. Typhi isolates:

- 60% (217/361) were resistant to nalidixic acid and 3.3% (12/361) to ciprofloxacin

Among *Shigella* isolates:

- 2.1% (10/475) were resistant to nalidixic acid and 0.6% (3/475) to ciprofloxacin

In *Campylobacter*, fluoroquinolones and macrolides (e.g., erythromycin) are important agents in the treatment of severe infections. Among *Campylobacter* isolates:

- 23% (344/1502) were ciprofloxacin resistant, including
  - 23% (312/1355) of *Campylobacter jejuni* isolates
  - 22% (31/143) of *Campylobacter coli* isolates
- 1.7% (25/1502) were erythromycin resistant, including
  - 1.5% (21/1355) *Campylobacter jejuni* isolates
  - 2.8% (4/143) of *Campylobacter coli* isolates

### Multidrug Resistance

Multidrug resistance is described in NARMS as resistance to three or more CLSI antimicrobial classes. Antimicrobial classes of agents defined by the Clinical and Laboratory Standards Institute (CLSI) are used in this report ([Table 3](#), [Table 4](#)). For non-typhoidal *Salmonella*, an important multidrug-resistant phenotype includes resistance to at least ampicillin, chloramphenicol, streptomycin, sulfonamide (sulfamethoxazole or sulfisoxazole), and tetracycline (ACSSuT). The ACSSuT phenotype includes resistance to at least five CLSI classes. Another important phenotype includes resistance to at least ampicillin, chloramphenicol, streptomycin, sulfonamide, tetracycline, amoxicillin-clavulanic acid, and ceftriaxone (ACSSuTAuCx). The ACSSuTAuCx phenotype includes resistance to at least 7 CLSI classes.



Among non-typhoidal *Salmonella* isolates:

- 13% (284/2192) were resistant to two or more CLSI classes of agents, 9.5% (209/2192) to three or more CLSI classes. Of the 209 isolates resistant to three or more CLSI classes, 50% were ser. Typhimurium. The serotypes with the highest proportion of isolates resistant to three or more CLSI classes were
  - Typhimurium (28%, 104/371), Heidelberg (26%, 22/86), and Newport (7.6%, 18/236)
- 5.1% (112/2192) were at least ACSSuT resistant. The serotypes with the highest proportion of isolates resistant to this phenotype were
  - Typhimurium (19%, 72/371) and Newport (6.4%, 15/236)
- 1.4% (30/2192) were at least ACSSuTAuCx resistant. The serotypes with the highest proportion of isolates resistant to this phenotype were
  - Newport (6.4%, 15/236) and Typhimurium (1.6%, 6/371)

Additional isolates resistant to three or more CLSI classes include

- 13% (46/361) of *Salmonella* ser. Typhi isolates
- 36% (173/475) of *Shigella* isolates
- 5.9% (11/188) of *E. coli* O157 isolates

## Box 1. Changes in antimicrobial resistance: 2009 vs. 2003–07

To understand changes in prevalence of antimicrobial resistance over time, we used logistic regression to compare the prevalence of specific antimicrobial resistance patterns among *Salmonella* and *Campylobacter* isolates tested in 2009 with the reference, which was the average prevalence of resistance in 2003–07. Since 2003, all 50 states have participated in *Salmonella* surveillance and all 10 FoodNet sites in *Campylobacter* surveillance. A description of the methods is included in this report (refer to Surveillance and Laboratory Testing Methods).

The differences between the prevalence of resistance in 2009 and the average prevalence of resistance in 2003–07 (Figure 1) were statistically significant for the following:

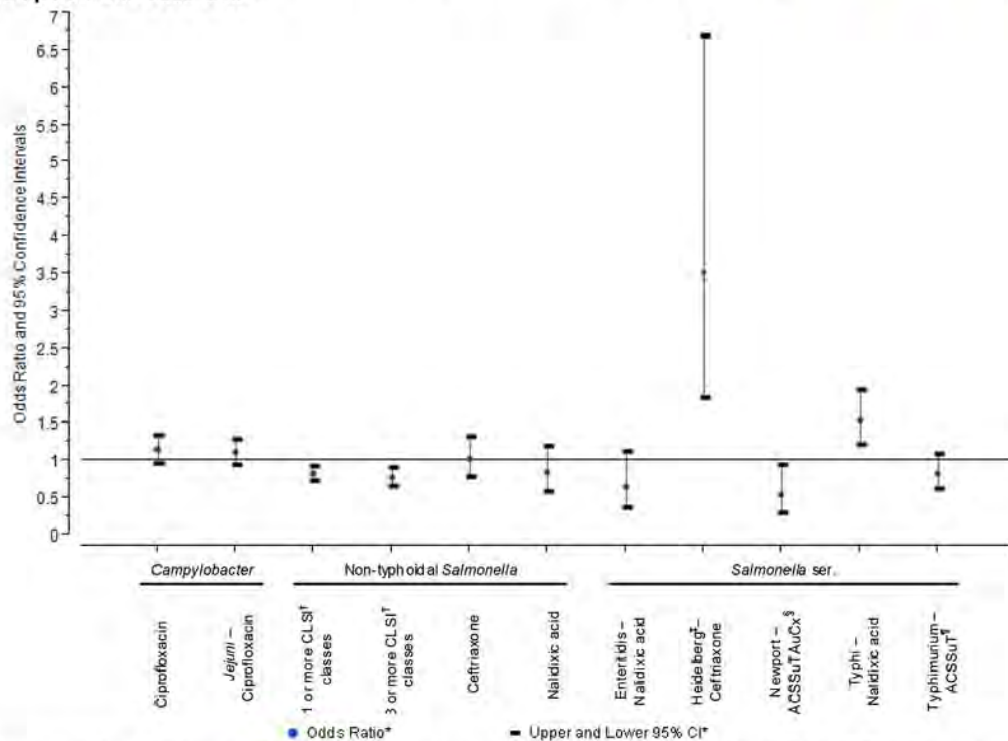
- Resistance to one or more CLSI classes in non-typhoidal *Salmonella* (NTS), lower in 2009 (17%) than in 2003–07 (20%) (OR=0.8, 95% CI [0.7–0.9])
- Resistance to three or more CLSI classes in NTS, lower in 2009 (9.5%) than in 2003–07 (12%) (OR=0.8, 95% CI [0.7–0.9])
- ACSSuTAuCx resistance in *Salmonella enterica* ser. Newport, lower in 2009 (6.4%) than in 2003–07 (13%) (OR=0.5, 95% CI [0.3–1.0])
- Nalidixic acid resistance in *Salmonella enterica* ser. Typhi, higher in 2009 (60%) than in 2003–07 (49%) (OR=1.6, 95% CI [1.2–2.0])

Ceftriaxone resistance in *Salmonella* ser. Heidelberg was higher in 2009 (21%) than the average prevalence of resistance in 2003–07 (7.9%) (OR=3.5, 95% CI [1.8, 6.7]) (Figure 1). The data indicate that increased resistance was mainly driven by California and Washington. Trend analysis excluding California and Washington shows no significant change (OR=1.4, 95% CI [0.6, 3.6]). Thus, the reported OR represents a summary of unequal trends among sites.

The differences between the prevalence of resistance in 2009 and the average prevalence of resistance in 2003–07 (Figure 1) were not statistically significant for the following:

- Nalidixic acid resistance in NTS (OR=0.9, 95% CI [0.6–1.2])
- Ceftriaxone resistance in NTS (OR=1.0, 95% CI [0.8–1.3])
- Nalidixic acid resistance in *Salmonella enterica* ser. Enteritidis (OR=0.7, 95% CI [0.4–1.1])
- ACSSuT resistance in *Salmonella enterica* ser. Typhimurium (OR=0.8, 95% CI [0.6–1.1])
- Ciprofloxacin resistance in *Campylobacter* (OR=1.1, 95% CI [1.0–1.3])
- Ciprofloxacin resistance in *Campylobacter jejuni* (OR=1.1, 95% CI [1.0–1.4])

Figure 1. Summary of trend analysis of the prevalence of specific resistance patterns among *Salmonella* and *Campylobacter* isolates, 2009 compared with 2003–2007\*



\*The reference is the average prevalence of resistance in 2003–07. Logistic regression models adjusted for site. The odds ratios (ORs) and 95% confidence intervals (CIs) for 2009 compared with the reference were calculated using unconditional maximum likelihood estimation. ORs that do not include 1.0 in the 95% CIs are reported as statistically significant.

<sup>†</sup>Antimicrobial classes of agents defined by the Clinical and Laboratory Standards Institute (CLSI) are used.

<sup>‡</sup>Descriptive analysis suggests that increased resistance in 2009 was mainly driven by California and Washington. Thus, the reported OR represents a summary of unequal trends across sites.

<sup>§</sup>ACSSuTAuCx: resistance to at least ampicillin, chloramphenicol, streptomycin, sulfonamide, tetracycline, amoxicillin-clavulanic acid, and ceftriaxone.

<sup>¶</sup>ACSSuT: resistance to at least ampicillin, chloramphenicol, streptomycin, sulfonamide, and tetracycline.

## Antimicrobial Resistance: 1996–2009

The following figures display resistance from 1996–2009 for non-typhoidal *Salmonella*, 2000–2009 for *Salmonella* ser. Typhi, and 1997–2009 for *Campylobacter*.

Figure 1. Percentage of non-typhoidal *Salmonella* isolates resistant to nalidixic acid, by year, 1996–2009

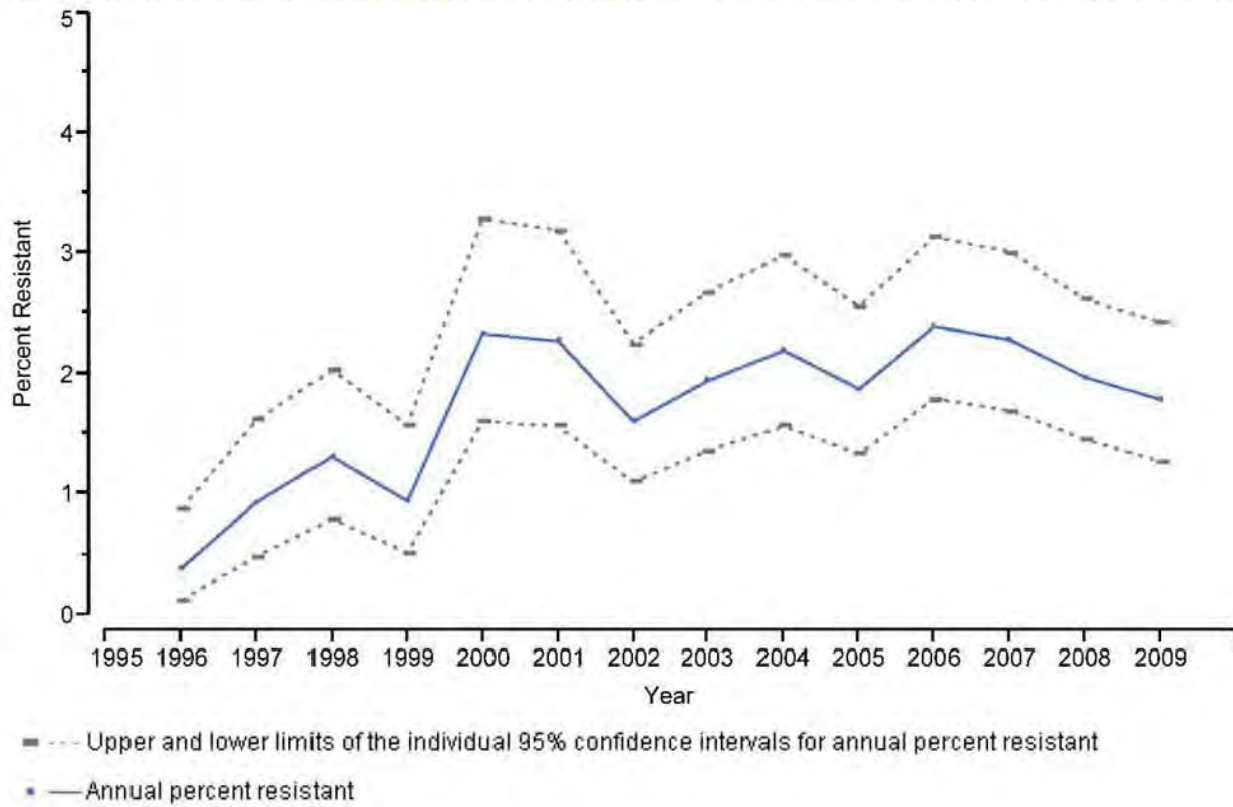


Figure 2. Percentage of *non-typhoidal Salmonella* isolates resistant to ceftriaxone, by year, 1996–2009

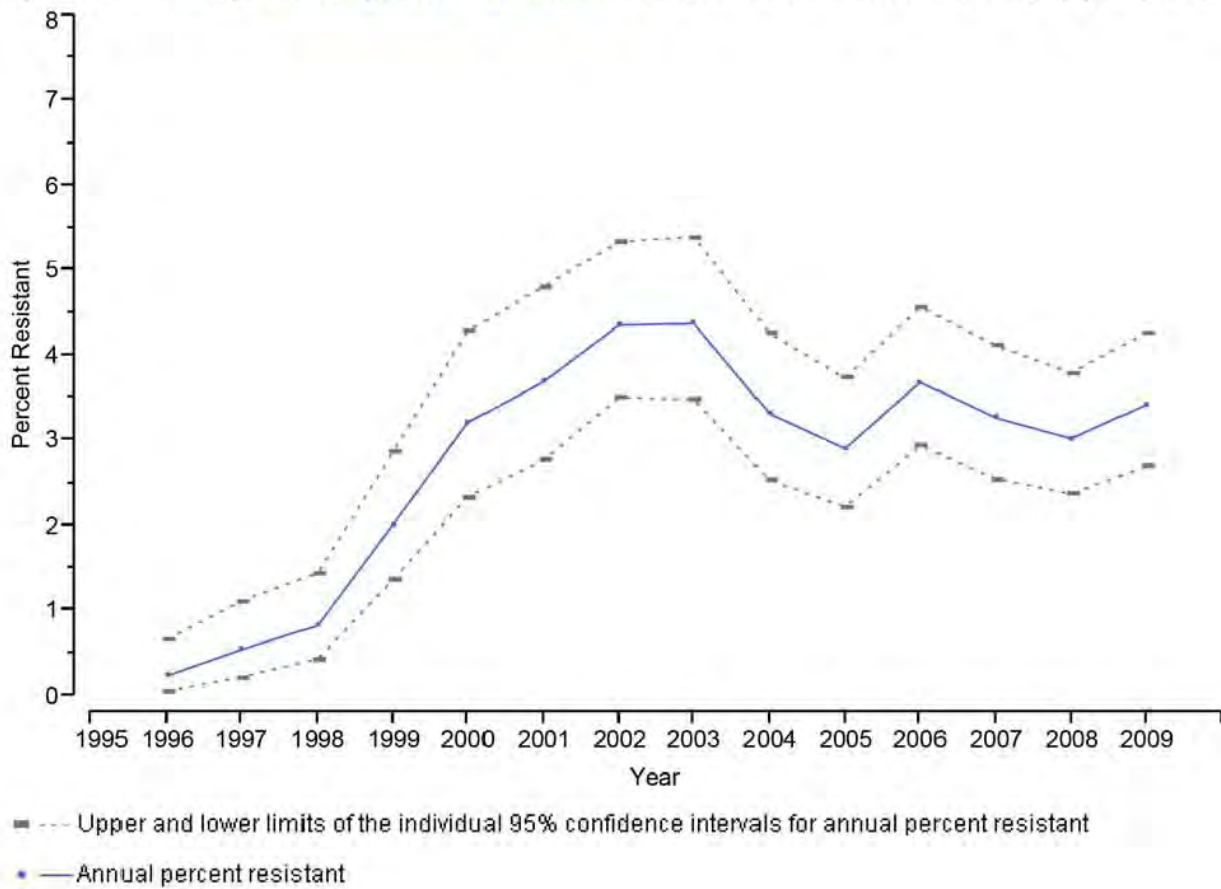


Figure 3. Percentage of *Salmonella ser. Enteritidis* isolates resistant to nalidixic acid, by year, 1996–2009

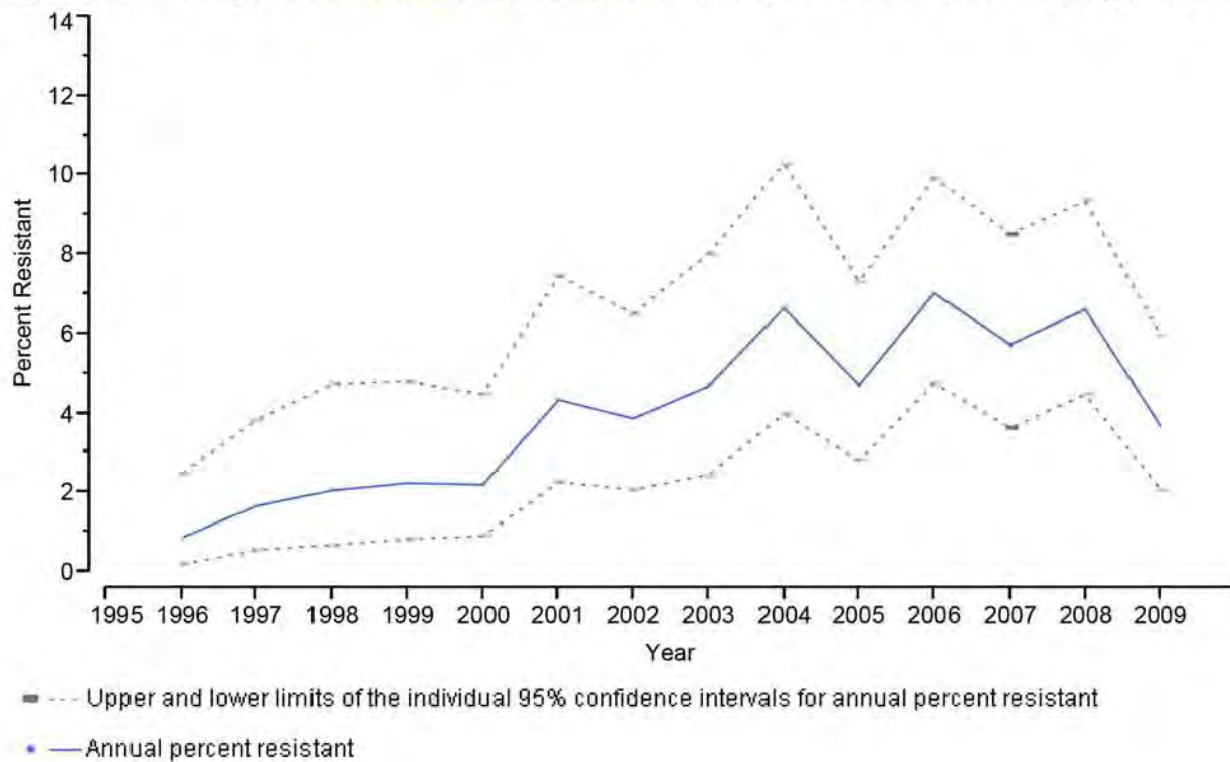


Figure 4. Percentage of *Salmonella ser. Heidelberg* isolates resistant to ceftriaxone, by year, 1996–2009

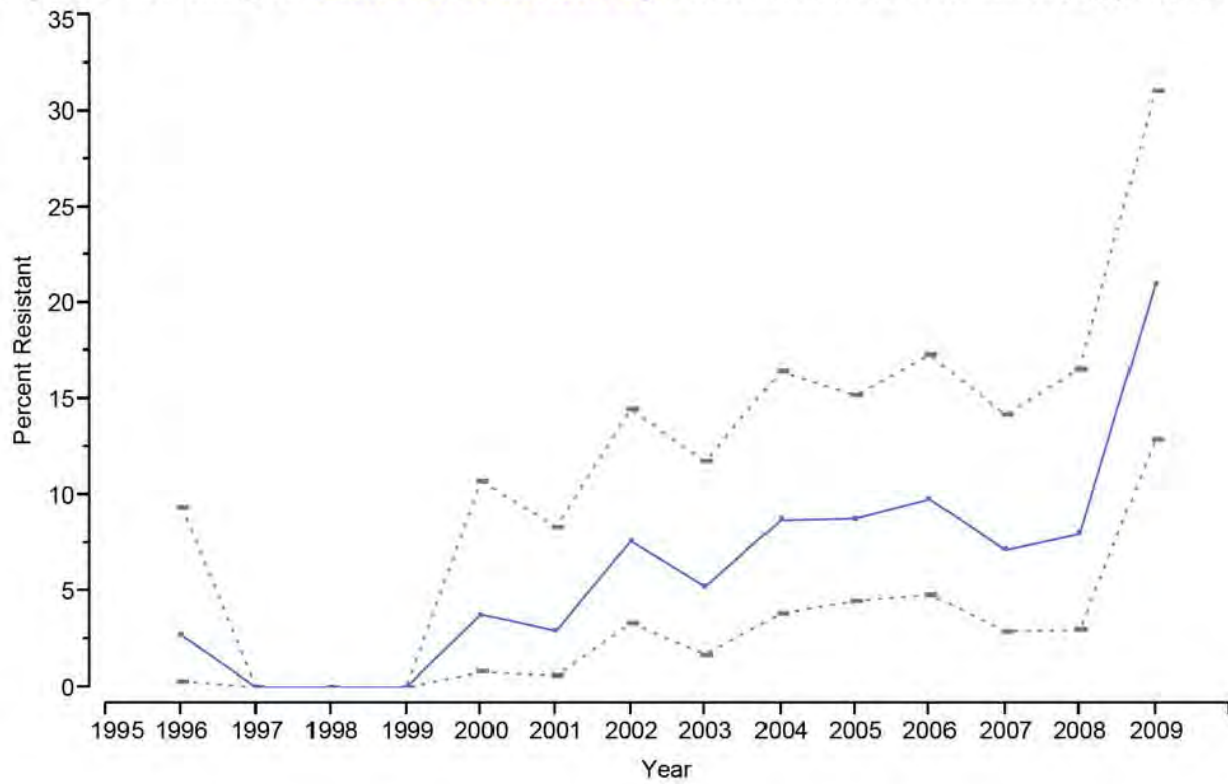
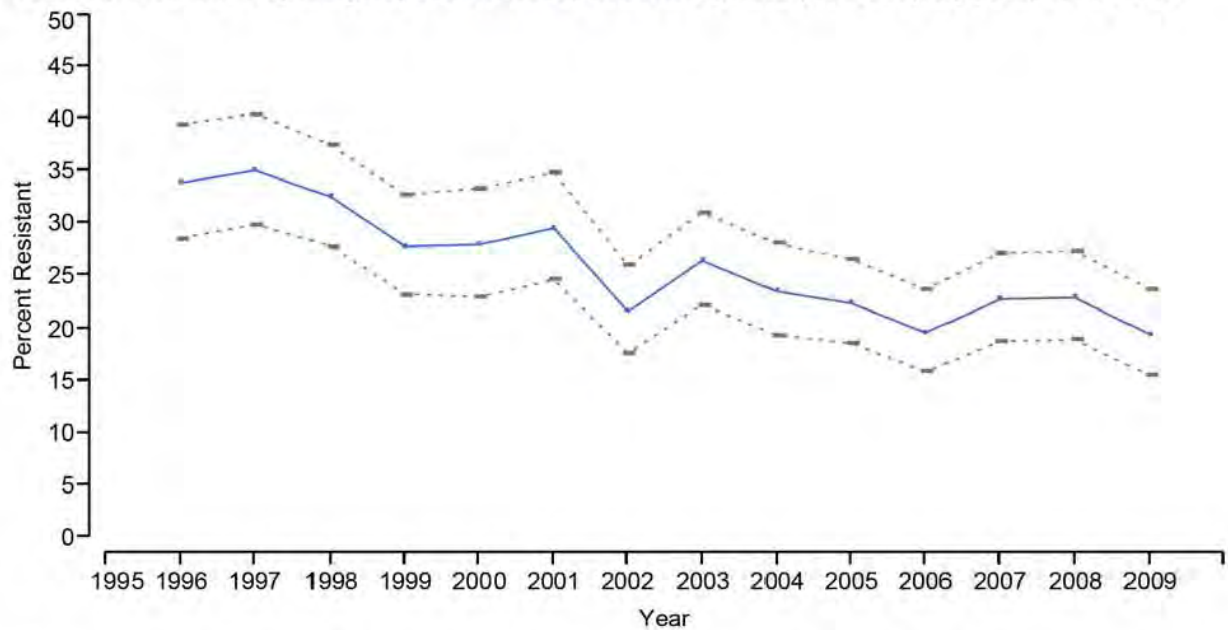
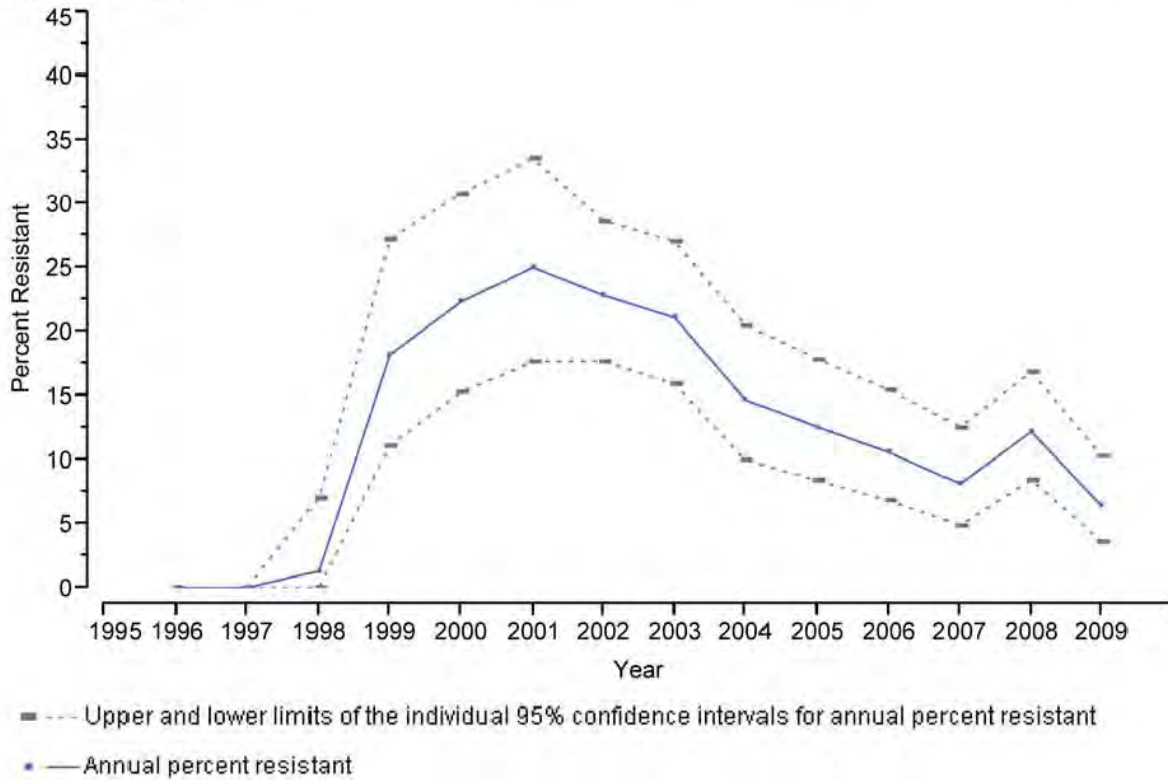


Figure 5. Percentage of *Salmonella ser. Typhimurium* isolates resistant to at least ampicillin, chloramphenicol, streptomycin, sulfonamide, and tetracycline (ACSSuT), by year, 1996–2009

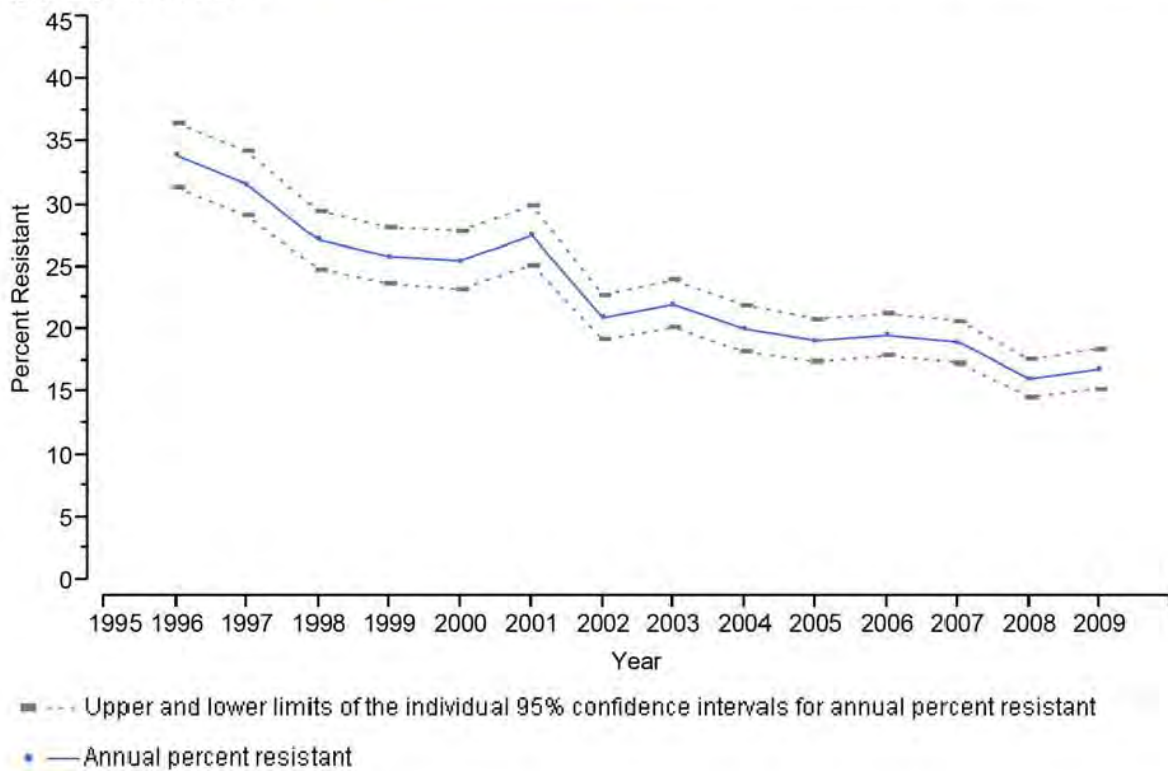


- --- Upper and lower limits of the individual 95% confidence intervals for annual percent resistant
- — Annual percent resistant

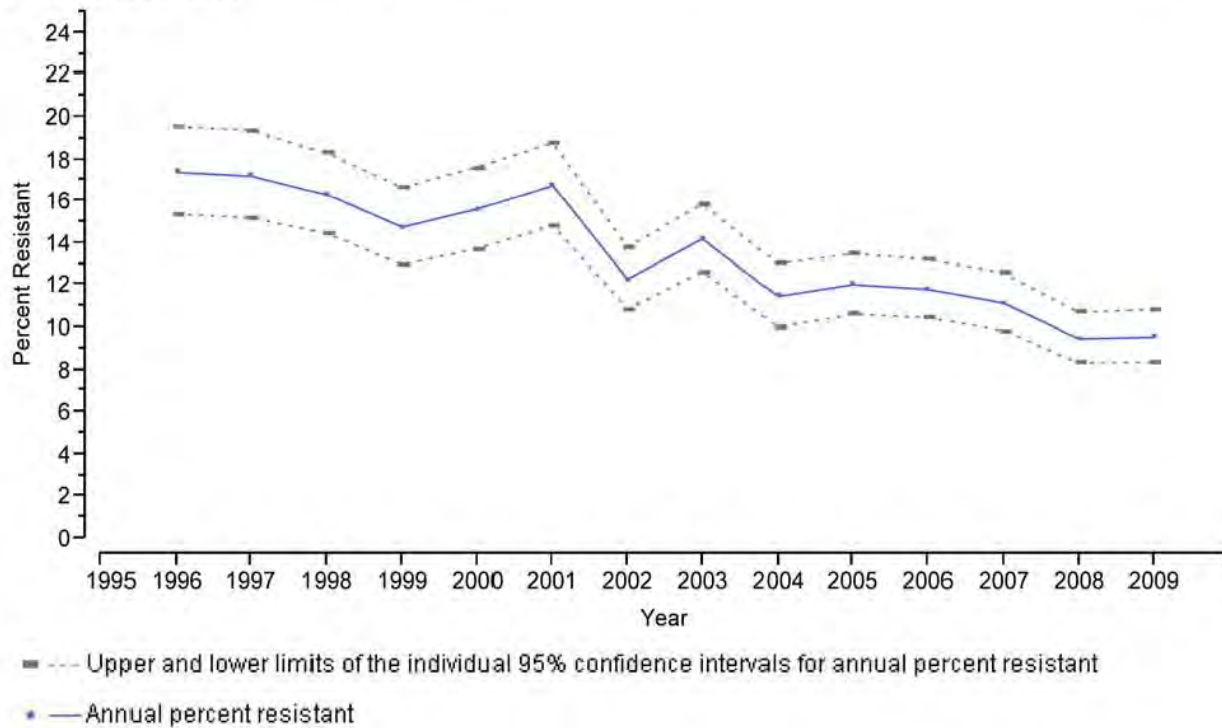
**Figure 6. Percentage of *Salmonella ser. Newport* isolates resistant to at least ampicillin, chloramphenicol, streptomycin, sulfonamide, tetracycline, amoxicillin-clavulanic acid, and ceftriaxone (ACSSuTAuCx), by year, 1996–2009**



**Figure 7. Percentage of non-typhoidal *Salmonella* isolates resistant to 1 or more antimicrobial classes, by year, 1996–2009**



**Figure 8. Percentage of non-typhoidal *Salmonella* isolates resistant to 3 or more antimicrobial classes, by year, 1996–2009**



**Figure 9. Percentage of *Salmonella ser. Typhi* isolates resistant to nalidixic acid, by year, 2000–2009**

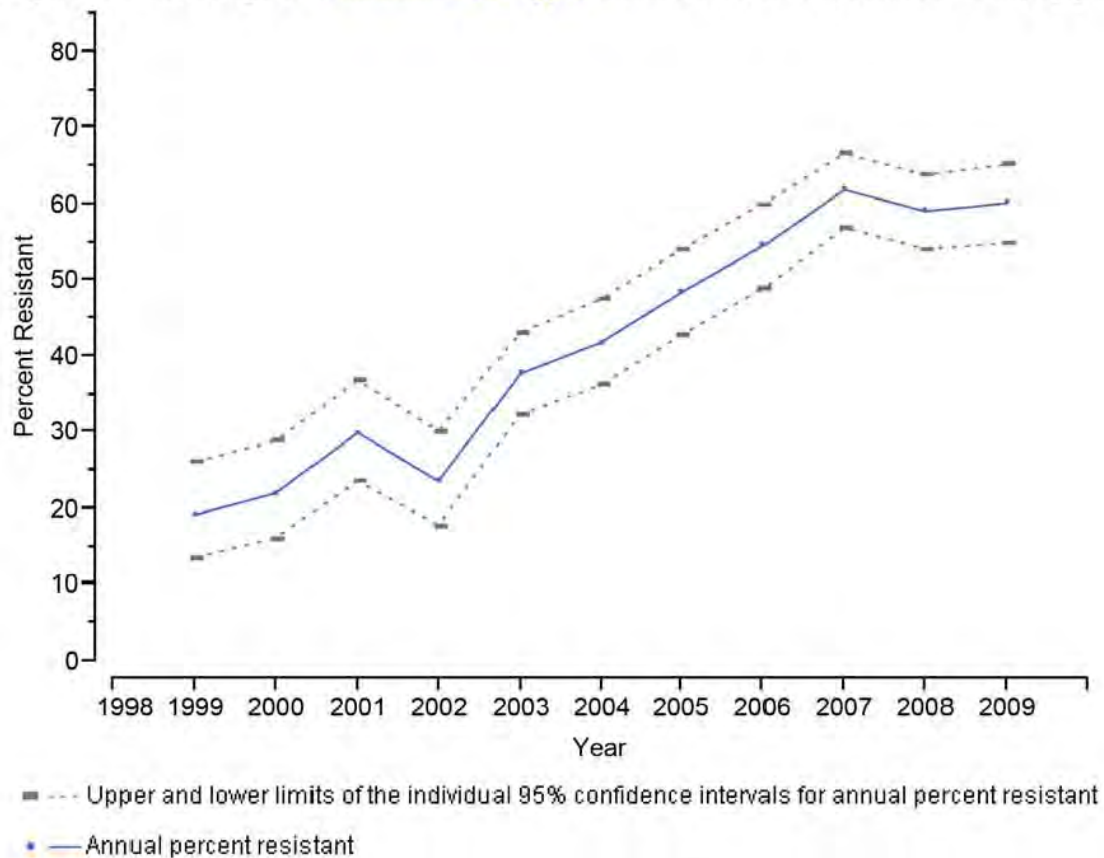
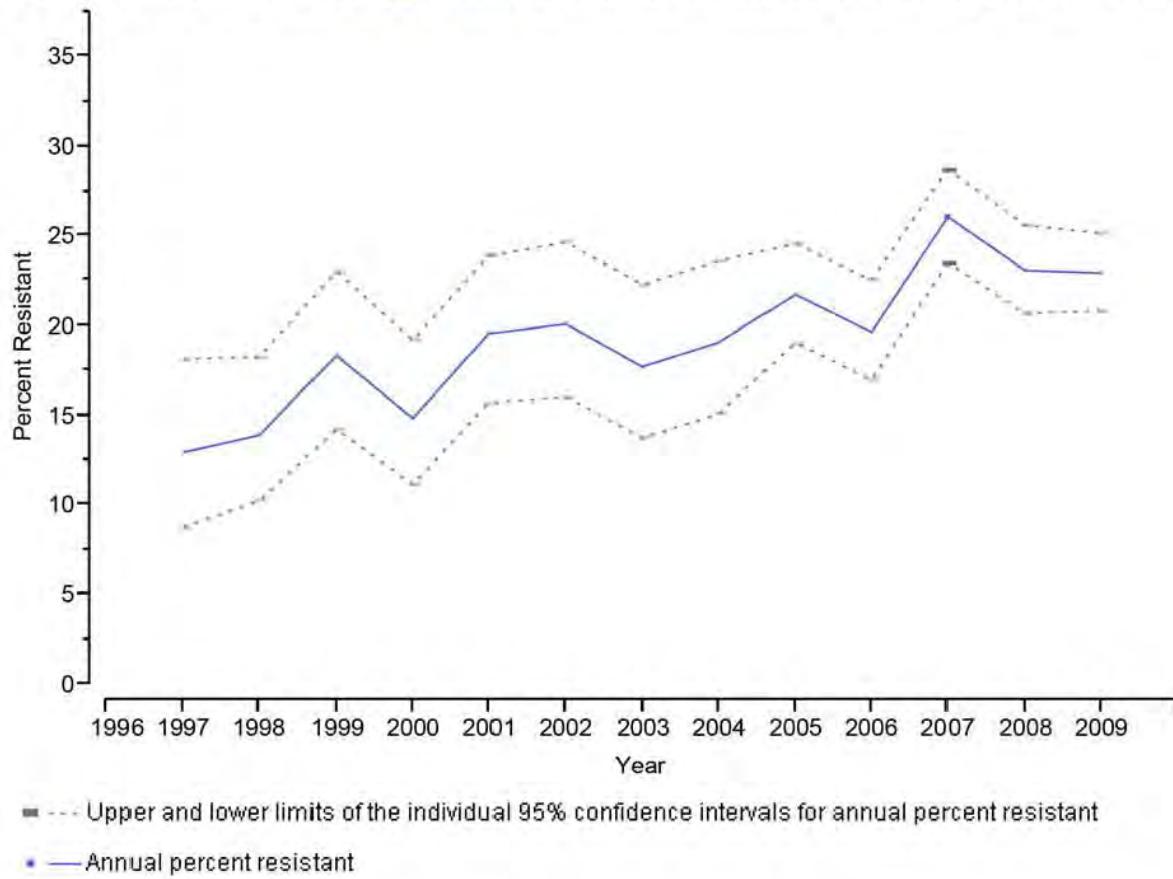


Figure 10. Percentage of *Campylobacter* isolates resistant to ciprofloxacin, by year, 1997–2009





**Table 2. Population size and number of isolates received and tested, NARMS, 2009**

State/Site	Population Size*	Non-typhoidal <i>Salmonella</i>		Typhoidal <i>Salmonella</i>		<i>Shigella</i>		<i>E. coli</i> O157		<i>Campylobacter</i> †	
		n	(%)	n	(%)	n	(%)	n	(%)	n	(%)
Alabama	4,708,708	65	(3.0%)	2	(0.4%)	9	(1.9%)	2	(1.1%)		
Alaska	698,473	4	(0.2%)	0	(0.0%)	1	(0.2%)	0	(0.0%)		
Arizona	6,595,778	56	(2.6%)	4	(0.9%)	27	(5.7%)	2	(1.1%)		
Arkansas	2,889,450	23	(1.0%)	0	(0.0%)	9	(1.9%)	2	(1.1%)		
California <sup>‡</sup>	27,113,653	184	(8.4%)	57	(12.3%)	2	(0.4%)	5	(2.7%)	76	(5.1%)
Colorado	5,024,748	34	(1.6%)	12	(2.6%)	6	(1.3%)	7	(3.7%)	57	(3.8%)
Connecticut	3,518,288	25	(1.1%)	5	(1.1%)	1	(0.2%)	4	(2.1%)	123	(8.2%)
Delaware	885,122	8	(0.4%)	0	(0.0%)	7	(1.5%)	0	(0.0%)		
District of Columbia	599,657	37	(1.7%)	3	(0.6%)	0	(0.0%)	0	(0.0%)		
Florida	18,537,969	17	(0.8%)	28	(6.0%)	0	(0.0%)	0	(0.0%)		
Georgia	9,829,211	133	(6.1%)	12	(2.6%)	29	(6.1%)	14	(7.4%)	473	(31.5%)
Hawaii	1,295,178	16	(0.7%)	6	(1.3%)	3	(0.6%)	1	(0.5%)		
Houston, Texas <sup>§</sup>	2,257,926	34	(1.6%)	8	(1.7%)	8	(1.7%)	0	(0.0%)		
Idaho	1,545,801	8	(0.4%)	2	(0.4%)	1	(0.2%)	2	(1.1%)		
Illinois	12,910,409	78	(3.6%)	15	(3.2%)	30	(6.3%)	17	(9.0%)		
Indiana	6,423,113	31	(1.4%)	4	(0.9%)	1	(0.2%)	2	(1.1%)		
Iowa	3,007,856	17	(0.8%)	0	(0.0%)	4	(0.8%)	4	(2.1%)		
Kansas	2,818,747	14	(0.6%)	0	(0.0%)	9	(1.9%)	1	(0.5%)		
Kentucky	4,314,113	21	(1.0%)	0	(0.0%)	4	(0.8%)	0	(0.0%)		
Los Angeles <sup>  </sup>	9,848,011	63	(2.9%)	6	(1.3%)	2	(0.4%)	0	(0.0%)		
Louisiana	4,492,076	30	(1.4%)	0	(0.0%)	0	(0.0%)	0	(0.0%)		
Maine	1,318,301	1	(0.0%)	0	(0.0%)	0	(0.0%)	0	(0.0%)		
Maryland	5,699,478	60	(2.7%)	18	(3.9%)	17	(3.6%)	24	(12.8%)	186	(12.4%)
Massachusetts	6,593,587	58	(2.6%)	16	(3.5%)	11	(2.3%)	4	(2.1%)		
Michigan	9,969,727	42	(1.9%)	15	(3.2%)	8	(1.7%)	2	(1.1%)		
Minnesota	5,266,214	28	(1.3%)	5	(1.1%)	4	(0.8%)	8	(4.3%)	166	(11.1%)
Mississippi	2,951,996	37	(1.7%)	1	(0.2%)	4	(0.8%)	0	(0.0%)		
Missouri	5,987,580	38	(1.7%)	1	(0.2%)	35	(7.4%)	6	(3.2%)		
Montana	974,989	7	(0.3%)	1	(0.2%)	1	(0.2%)	2	(1.1%)		
Nebraska	1,796,619	10	(0.5%)	0	(0.0%)	8	(1.7%)	3	(1.6%)		
Nevada	2,643,085	12	(0.5%)	3	(0.6%)	3	(0.6%)	1	(0.5%)		
New Hampshire	1,324,575	10	(0.5%)	0	(0.0%)	2	(0.4%)	2	(1.1%)		
New Jersey	8,707,739	57	(2.6%)	44	(9.5%)	15	(3.2%)	7	(3.7%)		
New Mexico	2,009,671	10	(0.5%)	1	(0.2%)	4	(0.8%)	2	(1.1%)	142	(9.5%)
New York <sup>¶</sup>	11,149,572	76	(3.5%)	19	(4.1%)	7	(1.5%)	6	(3.2%)	125	(8.3%)
New York City <sup>¶¶</sup>	8,391,881	74	(3.4%)	55	(11.9%)	18	(3.8%)	2	(1.1%)		
North Carolina	9,380,884	85	(3.9%)	11	(2.4%)	5	(1.1%)	2	(1.1%)		
North Dakota	646,844	6	(0.3%)	0	(0.0%)	2	(0.4%)	1	(0.5%)		
Ohio	11,542,645	81	(3.7%)	13	(2.8%)	20	(4.2%)	5	(2.7%)		
Oklahoma	3,687,050	34	(1.6%)	2	(0.4%)	12	(2.5%)	1	(0.5%)		
Oregon	3,825,657	26	(1.2%)	2	(0.4%)	3	(0.6%)	4	(2.1%)	113	(7.5%)
Pennsylvania	12,604,767	82	(3.7%)	22	(4.8%)	59	(12.4%)	1	(0.5%)		
Rhode Island	1,053,209	8	(0.4%)	1	(0.2%)	2	(0.4%)	1	(0.5%)		
South Carolina	4,561,242	58	(2.6%)	1	(0.2%)	4	(0.8%)	1	(0.5%)		
South Dakota	812,383	10	(0.5%)	2	(0.4%)	1	(0.2%)	3	(1.6%)		
Tennessee	6,296,254	41	(1.9%)	4	(0.9%)	16	(3.4%)	7	(3.7%)	41	(2.7%)
Texas <sup>§§</sup>	22,524,376	137	(6.3%)	16	(3.5%)	12	(2.5%)	2	(1.1%)		
Utah	2,784,572	23	(1.0%)	0	(0.0%)	1	(0.2%)	4	(2.1%)		
Vermont	621,760	1	(0.0%)	0	(0.0%)	0	(0.0%)	0	(0.0%)		
Virginia	7,882,590	60	(2.7%)	30	(6.5%)	7	(1.5%)	3	(1.6%)		
Washington	6,664,195	44	(2.0%)	5	(1.1%)	7	(1.5%)	7	(3.7%)		
West Virginia	1,819,777	35	(1.6%)	0	(0.0%)	10	(2.1%)	4	(2.1%)		
Wisconsin	5,654,774	34	(1.6%)	11	(2.4%)	22	(4.6%)	6	(3.2%)		
Wyoming	544,270	9	(0.4%)	0	(0.0%)	2	(0.4%)	4	(2.1%)		
<b>Total</b>	<b>307,006,550</b>	<b>2192</b>	<b>(100.0%)</b>	<b>463</b>	<b>(100.0%)</b>	<b>475</b>	<b>(100.0%)</b>	<b>188</b>	<b>(100.0%)</b>	<b>1502</b>	<b>(100.0%)</b>

\* US Census Bureau, 2009

† Typhoidal *Salmonella* includes Typhi, Paratyphi A, Paratyphi B, and Paratyphi C

‡ *Campylobacter* isolates are submitted only from FoodNet sites representing a total population 46,859,541. All *Campylobacter* isolates are received from Georgia, Maryland, New Mexico, Oregon, and Tennessee and every other isolate from California, Colorado, Connecticut, and New York; and every fifth isolate from Minnesota.

§ Excluding Los Angeles County

¶ Houston City

¶¶ Los Angeles County

|| Excluding New York City

¶¶ Five boroughs of New York City (Bronx, Brooklyn, Manhattan, Queens, Staten Island)

§§ Excluding Houston, Texas

### Surveillance Sites and Isolate Submissions

In 2009, NARMS conducted nationwide surveillance among approximately 307 million persons (2009 U.S. Census Bureau estimates). Public health laboratories systematically selected every 20<sup>th</sup> non-typhoidal *Salmonella*, *Shigella*, and *Escherichia coli* O157 isolate as well as every *Salmonella* ser. Typhi, *Salmonella* ser. Paratyphi A and *Salmonella* ser. Paratyphi C isolate received at their laboratories and forwarded these isolates to CDC for antimicrobial susceptibility testing. *Salmonella* ser. Paratyphi B was included in the every 20<sup>th</sup> sampling for non-typhoidal *Salmonella* because available laboratory methods do not always allow for consistent distinction between serotype Paratyphi B (which typically causes typhoidal illness) and serotype Paratyphi B var. L(+)/tartrate+ (which does not typically cause typhoidal illness).

Since 2005, public health laboratories of the 10 state health departments that participated in CDC's Foodborne Diseases Active Surveillance Network (FoodNet) have forwarded a representative sample of *Campylobacter* isolates to CDC for susceptibility testing. The FoodNet sites, representing approximately 47 million persons (2009 U.S. Census Bureau estimates), include California, Colorado, Connecticut, Georgia, Maryland, Minnesota, New Mexico, New York, Oregon, and Tennessee. Depending on the burden of *Campylobacter* in each FoodNet site, one of the following three methods was used to obtain a representative sample of *Campylobacter* isolates: all isolates received by Georgia, Maryland, New Mexico, Oregon, and Tennessee; every other isolate from California, Colorado, Connecticut, and New York; and every fifth isolate from Minnesota. From 1997 to 2004, one *Campylobacter* isolate was submitted each week from participating FoodNet sites.

### Testing of *Salmonella*, *Shigella*, and *Escherichia coli* O157

#### Antimicrobial Susceptibility Testing

*Salmonella*, *Shigella*, and *E. coli* O157 isolates were tested using broth microdilution (Sensititre<sup>®</sup>, Trek Diagnostics, Cleveland, OH) according to manufacturer's instructions to determine the minimum inhibitory concentration (MIC) for each of 15 antimicrobial agents: amikacin, ampicillin, amoxicillin-clavulanic acid, cefoxitin, ceftiofur, ceftriaxone, chloramphenicol, ciprofloxacin, gentamicin, kanamycin, nalidixic acid, streptomycin, sulfisoxazole, tetracycline, and trimethoprim-sulfamethoxazole (Table 3). Before 2004, sulfamethoxazole was used instead of sulfisoxazole to represent the sulfonamides. Interpretive criteria defined by CLSI were used when available. The resistance breakpoint for amikacin, according to CLSI guidelines, is  $\geq 64$   $\mu\text{g}/\text{mL}$ . In 2002 and 2003, a truncated broth microdilution series was used for amikacin testing (0.5-4  $\mu\text{g}/\text{mL}$ ). For isolates that grew in all amikacin dilutions on the Sensititre panel (MIC > 4  $\mu\text{g}/\text{mL}$ ), ETest<sup>®</sup> (AB BIODISK, Solna, Sweden) was performed to determine amikacin MIC. The amikacin ETest<sup>®</sup> strip range of dilutions was 0.016-256  $\mu\text{g}/\text{mL}$ . Since 2004, amikacin had a full range of dilutions (0.5-64  $\mu\text{g}/\text{mL}$ ) on the Sensititre panel (CMV1AGNF).

In January 2010, CLSI published revised interpretive criteria for ceftriaxone and *Enterobacteriaceae*; the revised resistance breakpoint for ceftriaxone is MIC  $\geq 4$   $\mu\text{g}/\text{mL}$ . In this report, NARMS has applied the revised CLSI breakpoint for ceftriaxone resistance to data from all years.

**Table 3. Antimicrobial agents used for susceptibility testing for *Salmonella*, *Shigella*, and *Escherichia coli* O157 isolates, NARMS, 2009**

CLSI class	Antimicrobial Agent	Antimicrobial Agent Concentration Range ( $\mu\text{g/mL}$ )	MIC Interpretive Standard ( $\mu\text{g/mL}$ )		
			Susceptible	Intermediate	Resistant
Aminoglycosides	Amikacin	0.5–64	$\leq 16$	32	$\geq 64$
	Gentamicin	0.25–16	$\leq 4$	8	$\geq 16$
	Kanamycin	8–64	$\leq 16$	32	$\geq 64$
	Streptomycin*	32–64	$\leq 32$		$\geq 64$
$\beta$ -lactam / $\beta$ -lactamase inhibitor combinations	Amoxicillin-clavulanic acid	1/0.5–32/16	$\leq 8/4$	16/8	$\geq 32/16$
Cephems	Cefoxitin	0.5–32	$\leq 8$	16	$\geq 32$
	Ceftiofur	0.12–8	$\leq 2$	4	$\geq 8$
	Ceftriaxone <sup>†</sup>	0.25–64	$\leq 1$	2	$\geq 4$
	Cephalothin <sup>‡</sup>	2–32	$\leq 8$	16	$\geq 32$
Folate pathway inhibitors	Sulfamethoxazole <sup>§</sup>	16–512	$\leq 256$		$\geq 512$
	Sulfisoxazole	16–256	$\leq 256$		$\geq 512$
	Trimethoprim-sulfamethoxazole	0.12/2.38–4/76	$\leq 2/38$		$\geq 4/76$
Penicillins	Ampicillin	1–32	$\leq 8$	16	$\geq 32$
Phenicol	Chloramphenicol	2–32	$\leq 8$	16	$\geq 32$
Quinolones	Ciprofloxacin	0.015–4	$\leq 1$	2	$\geq 4$
	Nalidixic acid	0.5–32	$\leq 16$		$\geq 32$
Tetracyclines	Tetracycline	4–32	$\leq 4$	8	$\geq 16$

\* No CLSI breakpoints; resistance breakpoint used in NARMS is  $\geq 64 \mu\text{g/mL}$ .

<sup>†</sup> CLSI updated the ceftriaxone interpretive standards in January, 2010. Previous standards that were used for NARMS Human Isolate reports from 1996-2008 were susceptible  $\leq 8 \mu\text{g/mL}$ , intermediate 16-32  $\mu\text{g/mL}$ , and resistant  $\geq 64 \mu\text{g/mL}$ .

<sup>‡</sup> Cephalothin was tested from 1996 to 2003 for *Salmonella*, *Shigella*, and *E. coli* O157.

<sup>§</sup> Sulfamethoxazole, which was tested during 1996–2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.

## Additional Testing of *Salmonella* Strains

### Cephalosporin Retesting of Isolates from 1996-1998

Review of *Salmonella* isolates tested in NARMS during 1996 to 1998 gave conflicting cephalosporin susceptibility results. In particular, some isolates previously reported in NARMS as ceftiofur-resistant exhibited a low ceftriaxone MIC and, in some cases, did not exhibit an elevated MIC to other  $\beta$ -lactams. Because these findings suggested that some previously reported results were inaccurate, we retested, using the 2003 NARMS Sensititre<sup>®</sup> plate, isolates of *Salmonella* tested in NARMS during 1996 to 1998 that exhibited an MIC  $\geq 2$   $\mu\text{g/mL}$  to ceftiofur or ceftriaxone. The retest results have been included in the NARMS annual reports since 2003.

### Serotype Confirmation/Categorization

*Salmonella* serotype reported by the submitting laboratory was used for reporting with few exceptions. Serotype was confirmed by CDC for isolates that underwent subsequent molecular analysis for publication. Because of challenges associated with interpretation of tartrate fermentation assays, ability to ferment tartrate was confirmed for isolates reported as *Salmonella* ser. Paratyphi B by the submitting laboratory (serotype Paratyphi B is by definition unable to ferment L(+) tartrate). To distinguish *Salmonella* serotypes Paratyphi B and Paratyphi B var L(+) tartrate+ (formerly serotype Java), CDC performed Jordan's tartrate test and/or Kauffmann's tartrate test on all *Salmonella* ser. Paratyphi B isolates from 1996 to 2009 for which the tartrate result was not reported or was reported to be negative. Isolates negative for tartrate fermentation by both assays were categorized as serotype Paratyphi B. Isolates that were positive for tartrate fermentation by either assay were categorized as serotype Paratyphi B var L(+) tartrate+. Confirmation of other biochemical reactions or somatic and flagellar antigens was not performed at CDC.

Because of increased submissions of *Salmonella* ser. I 4,[5],12:i:- noted in previous years, and recognition of the possibility that this serotype may have been underreported in previous years, isolates reported as serogroup B and tested in NARMS during 1996 to 2009 were reviewed for additional information; isolates that could be clearly identified as serogroup B, first-phase flagellar antigen "1", second phase flagellar antigen absent were categorized in this report as *Salmonella* ser. I 4,[5],12:i:-.

### Testing of *Campylobacter*

#### Changes in Testing Methods in 2005

Starting in 2005, there were four changes in the methodology used for *Campylobacter*. First, a surveillance scheme for selecting a representative sample of *Campylobacter* isolates for submission by FoodNet sites was implemented. State public health laboratories within FoodNet sites receive *Campylobacter* isolates from reference and clinical laboratories within their state. In 2005, FoodNet sites changed from submitting the first isolate received each week to submitting every isolate (Georgia, Maryland, New Mexico, Oregon, and Tennessee), every other isolate (California, Colorado, Connecticut, and New York), or every fifth isolate received (Minnesota). The number of laboratories submitting isolates ranged from two to all. Second, the method of species identification was updated to parallel what is used by the CDC National *Campylobacter* Laboratory. Third, the susceptibility testing method changed from Etest<sup>®</sup> (AB bioMerieux, Solna, Sweden) to broth microdilution. Fourth, there were changes in the antimicrobial agents tested. Florfenicol replaced chloramphenicol as the phenicol class representative drug, and telithromycin was added to the NARMS panel of agents tested. These methods began in 2005 and continue through the current year's report.

### Identification/Speciation and Antimicrobial Susceptibility Testing

From 2005 through 2009, isolates were confirmed as *Campylobacter* by determination of typical morphology and motility using dark-field microscopy and a positive oxidase test reaction. Identification of *C. jejuni* was performed using the hippurate hydrolysis test. Hippurate-positive isolates were identified as *C. jejuni*. Hippurate-negative isolates were further characterized with polymerase chain reaction (PCR) assays with specific targets for *C. jejuni* (*mapA* or *hipO* gene), *C. coli*-specific *ceuE* gene (Linton *et al.* 1997, Gonzales *et al.* 1997, Pruckler *et al.* 2006), or other species specific primers. From 2003 to 2004, putative *Campylobacter* isolates were identified as *C. jejuni* or *C. coli* using BAX<sup>®</sup> System PCR Assay according to the manufacturer's instructions (DuPont Qualicon, Wilmington, DE). Isolates not identified as *C. jejuni* or *C. coli* were further characterized by other PCR assays

(Linton *et al.* 1996) or were characterized by the CDC National *Campylobacter* Reference Laboratory. From 1997 to 2002, methodology similar to that used from 2005 to 2009 was used.

The methods for susceptibility testing *Campylobacter* and criteria for interpreting the results have changed during the course of NARMS surveillance. Beginning in 2005, broth microdilution using the Sensititre® system (Trek Diagnostics, Cleveland, OH) was performed according to manufacturer's instructions to determine the MICs for nine antimicrobial agents: azithromycin, ciprofloxacin, clindamycin, erythromycin, florfenicol, gentamicin, nalidixic acid, telithromycin, and tetracycline (Table 4). CLSI recommendations for quality control were followed. From 1997 to 2004, Etest® (AB bioMerieux, Solna, Sweden) was used for susceptibility testing of *Campylobacter* isolates. *Campylobacter*-specific CLSI interpretive criteria were used for erythromycin, ciprofloxacin, and tetracycline beginning with the 2004 NARMS annual report. NARMS breakpoints were used when CLSI breakpoints were not available. Beginning in 2004, NARMS breakpoints were established based on the MIC distributions of NARMS isolates and the presence of known resistance genes or mutations. In pre-2004 annual reports, NARMS breakpoints used were based on those available for other organisms. Establishment of breakpoints based on MIC distributions resulted in higher MIC definitions for azithromycin and erythromycin resistance compared with those reported in pre-2004 annual reports. The breakpoints listed in Table 4 have been applied to MIC data collected for all years so that resistance prevalence is comparable over time.

**Table 4. Antimicrobial agents used for susceptibility testing of *Campylobacter* isolates, NARMS, 1997–2009**

CLSI class	Antimicrobial Agent	Antimicrobial Agent Concentration Range (µg/mL)	MIC Interpretive Standard (µg/mL)		
			Susceptible	Intermediate	Resistant
Aminoglycosides	Gentamicin	0.12–32 0.016–256*	≤2	4	≥8
Ketolides	Telithromycin†	0.015–8	≤4	8	≥16
Lincosamides	Clindamycin	0.03–16 0.016–256*	≤2	4	≥8
Macrolides	Azithromycin	0.015–64 0.016–256*	≤2	4	≥8
	Erythromycin	0.03–64 0.016–256*	≤8	16	≥32
Phenicol	Chloramphenicol‡	0.016–256*	≤8	16	≥32
	Florfenicol§	0.03–64	≤4	N/A	N/A
Quinolones	Ciprofloxacin	0.015–64 0.002–32*	≤1	2	≥4
	Nalidixic acid	4–64 0.016–256*	≤16	32	≥64
Tetracyclines	Tetracycline	0.06–64 0.016–256*	≤4	8	≥16

\* Etest dilution range used from 1997–2004.

† Telithromycin added to NARMS panel in 2005.

‡ Chloramphenicol, tested from 1997–2004, was replaced by florfenicol in 2005.

§ Currently only a susceptible breakpoint (≤4 µg/mL) has been established. In this report isolates with a MIC ≥8 µg/mL are categorized as resistant.

## Retesting

Known mechanisms of quinolone resistance in *Campylobacter* are expected to confer equivalent susceptibilities to nalidixic acid and ciprofloxacin. Similarly, known mechanisms of macrolide resistance are expected to confer equivalent susceptibilities to erythromycin and azithromycin. Confirmatory testing of isolates with conflicting results was performed by broth microdilution methods (Sensititre®, Trek Diagnostics, Cleveland, OH). Totals reported here reflect the retest results.

## Data Analysis

For all pathogens, MICs were categorized as resistant, intermediate (if applicable), or susceptible. Analysis was restricted to the first isolate received (per genus under surveillance) per patient in the calendar year. If two or more isolates were received for the same patient for *Salmonella* ser. Typhi, the first blood isolate collected would be included in analysis. If no blood isolates were submitted, the first isolate collected would be included in analysis. Where established, CLSI interpretive criteria were used; streptomycin resistance was defined as MIC  $\geq 64$   $\mu\text{g/mL}$  (Table 3). The 95% confidence intervals (CIs) for the percentage of resistant isolates are included in the MIC distribution tables. The 95% CIs were calculated using the Paulson-Camp-Pratt approximation method.

When describing results for several years, multidrug resistance for *Salmonella*, *Shigella*, and *E. coli* O157 isolates was limited to the eight CLSI classes (Table 3) tested in all years from 1996 through 2009 represented by 15 agents: amikacin, amoxicillin-clavulanic acid, ampicillin, cefoxitin, ceftiofur, ceftriaxone, chloramphenicol, ciprofloxacin, gentamicin, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline, and trimethoprim-sulfamethoxazole. When describing multidrug resistance for several years for *Campylobacter* isolates, multidrug resistance was limited to the five CLSI classes tested in all years from 1997 through 2009, represented by ciprofloxacin, chloramphenicol/florfenicol, clindamycin, erythromycin, nalidixic acid, and tetracycline.

Logistic regression was used to compare the prevalence of antimicrobial resistance among *Salmonella* and *Campylobacter* isolates tested in 2009 with the reference, which was the average prevalence of resistance in the first five years that NARMS surveillance was nationwide (2003–07). The analysis included the following:

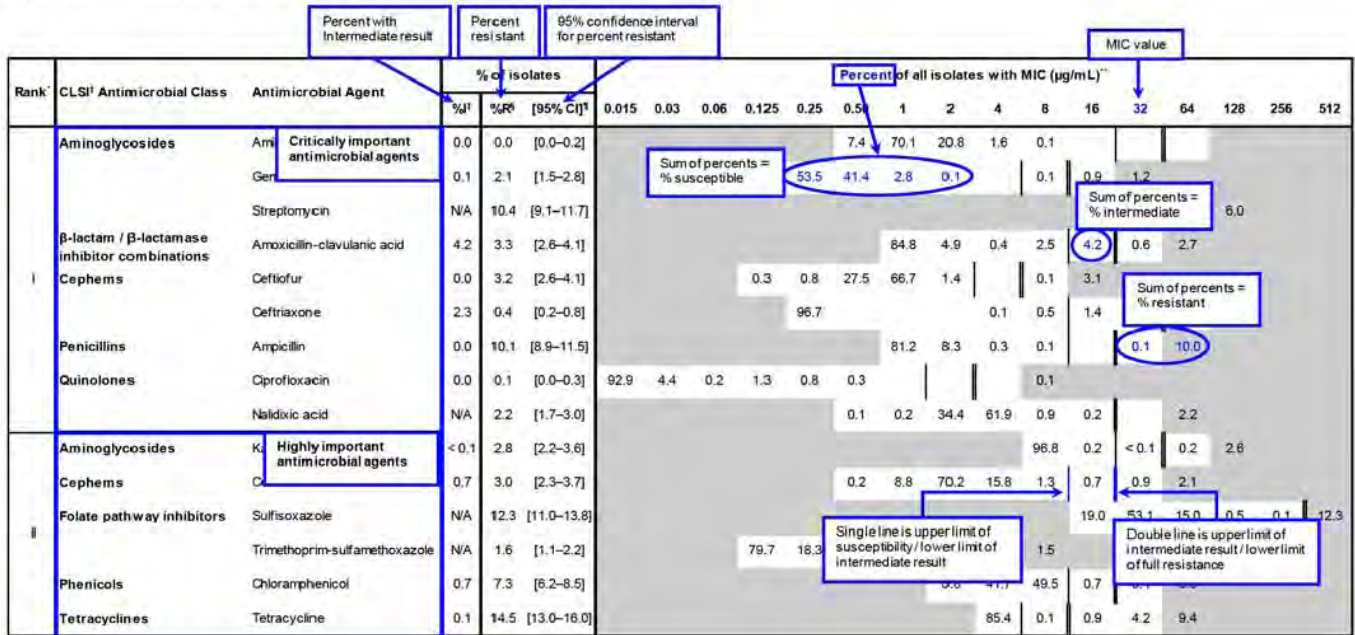
1. Non-typhoidal *Salmonella*: resistance to nalidixic acid, resistance to ceftriaxone, resistance to one or more CLSI classes, resistance to three more CLSI classes
2. *Salmonella* ser. Enteritidis: resistance to nalidixic acid
3. *Salmonella* ser. Typhimurium: resistance to at least ACSSuT (ampicillin, chloramphenicol, streptomycin, sulfonamide, and tetracycline)
4. *Salmonella* ser. Newport: resistance to at least ACSSuTAuCx (ACSSuT, amoxicillin-clavulanic acid, and ceftriaxone)
5. *Salmonella* ser. Typhi: resistance to nalidixic acid
6. *Campylobacter* species: resistance to ciprofloxacin
7. *Campylobacter jejuni*: resistance to ciprofloxacin

To account for site-to-site variation in the prevalence of antimicrobial resistance, we included main effects adjustments for site in the analysis. The final regression models for *Salmonella* adjusted for the submitting site using the nine geographic regions described in the Public Health Laboratory Information System (PHLIS): East North Central, East South Central, Mid-Atlantic, Mountain, New England, Pacific, South Atlantic, West North Central, and West South Central. For *Campylobacter*, the final regression models adjusted for the submitting FoodNet site. Odds ratios (ORs) and 95% confidence intervals (CIs) were calculated using unconditional maximum likelihood estimation. The adequacy of model fit was assessed in several ways. The significance of the main effect of year was assessed using the likelihood ratio test. The likelihood ratio test was also used to test for significance of interaction between site and year, although the power of the test to detect a single site-specific interaction was low. The Hosmer and Lemeshow goodness-of-fit test was also used. Finally, residual analysis was performed to examine the influence of individual observations. Having assessed that the main effect of year was significant, we reported ORs with 95% CIs (for 2009 compared with reference) that did not include 1.00 as statistically significant.

## MIC Distribution Tables and Proportional Figures

An explanation on “how to read a squashtogram” has been provided to assist the reader with the different parts of each table (Figure 11). A squashtogram shows the distribution of MICs for antimicrobial agents tested. Proportional figures visually display data from squashtograms for an immediate comparative summary of resistance in specific pathogens and serotypes. These figures are a categorical visual aid for the interpretation of MIC values. For most antimicrobial agents tested, three categories (susceptible, intermediate, and resistant) are used to interpret MICs. The proportion representing each category is shown in a horizontal proportional bar chart (Figure 12).

Figure 11. How to read a squashtogram









**Table 6. Percentage and number of non-typhoidal *Salmonella* isolates resistant to antimicrobial agents, 2000–2009**

Year		2000	2001	2002	2003	2004	2005	2006	2007	2008	2009	
Total Isolates		1372	1410	1998	1855	1782	2034	2173	2144	2380	2192	
Rank	CLSI <sup>†</sup> Antimicrobial Class	Antibiotic (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Gentamicin (MIC ≥ 16)	2.7%	1.9%	1.4%	1.4%	1.3%	2.2%	2.0%	2.1%	1.5%	1.3%
		Streptomycin (MIC ≥ 64)	16.3%	17.1%	13.2%	15.0%	12.0%	11.1%	10.7%	10.4%	10.0%	8.9%
	β lactam/β lactamase inhibitor combinations	Amoxicillin clavulanic acid (MIC ≥ 32/16)	3.9%	4.7%	5.3%	4.6%	3.7%	3.2%	3.7%	3.3%	3.1%	3.4%
			54	66	106	86	66	85	81	70	73	75
	Cephems	Ceftiofur (MIC ≥ 8)	3.2%	4.1%	4.4%	4.5%	3.4%	2.9%	3.6%	3.3%	3.0%	3.4%
		Ceftriaxone (MIC ≥ 64)	3.2%	3.7%	4.4%	4.4%	3.3%	2.9%	3.7%	3.3%	3.0%	3.4%
	Penicillins	Ampicillin (MIC ≥ 32)	15.9%	17.5%	13.0%	13.6%	12.1%	11.4%	11.0%	10.1%	9.7%	9.9%
			218	247	259	253	216	232	238	217	231	216
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.4%	0.2%	0.1%	0.2%	0.2%	0.0%	0.1%	0.1%	0.1%	0.0%
Nalidixic acid (MIC ≥ 32)		2.3%	2.3%	1.6%	1.9%	2.2%	1.9%	2.4%	2.3%	2.0%	1.8%	
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	5.6%	4.8%	3.8%	3.5%	2.8%	3.4%	2.9%	2.8%	2.1%	2.5%
			77	68	76	64	50	70	63	61	50	54
	Cephems	Cefoxitin (MIC ≥ 32)	3.2%	3.4%	4.3%	4.3%	3.4%	3.0%	3.5%	2.9%	3.0%	3.2%
		Cephalothin (MIC ≥ 32)	4.0%	4.0%	5.1%	5.3%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
	Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole <sup>‡</sup> (MIC ≥ 512)	17.1%	17.8%	12.9%	15.1%	13.3%	12.6%	12.1%	12.3%	10.1%	9.9%
		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	2.0%	2.0%	1.4%	1.9%	1.7%	1.7%	1.7%	1.5%	1.6%	1.7%
	Phenicol	Chloramphenicol (MIC ≥ 32)	10.1%	11.6%	8.6%	10.1%	7.6%	7.8%	6.4%	7.3%	6.2%	5.7%
			138	164	172	187	136	159	139	156	147	125
	Tetracyclines	Tetracycline (MIC ≥ 16)	18.7%	19.9%	14.9%	16.3%	13.6%	13.9%	13.5%	14.5%	11.6%	11.9%
			256	280	298	303	242	282	293	310	275	261

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.

**Table 7. Resistance patterns of non-typhoidal *Salmonella* isolates, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates	1372	1410	1998	1855	1782	2034	2173	2144	2380	2192
	%	%	%	%	%	%	%	%	%	%
	n	n	n	n	n	n	n	n	n	n
No resistance detected	74.5%	72.5%	79.1%	78.0%	80.0%	80.9%	80.5%	81.1%	84.0%	83.2%
	1022	1022	1580	1447	1425	1646	1749	1738	1999	1823
Resistance ≥ 1 CLSI class*	25.5%	27.5%	20.9%	22.0%	20.0%	19.1%	19.5%	18.9%	16.0%	16.8%
	350	388	418	408	357	388	424	406	381	369
Resistance ≥ 2 CLSI classes*	20.0%	22.1%	15.8%	17.5%	15.0%	14.8%	14.7%	14.2%	12.4%	13.0%
	275	311	315	325	267	302	319	305	295	284
Resistance ≥ 3 CLSI classes*	15.6%	16.7%	12.3%	14.2%	11.4%	12.0%	11.8%	11.1%	9.4%	9.5%
	214	236	245	263	204	244	256	239	224	209
Resistance ≥ 4 CLSI classes*	12.7%	13.5%	9.8%	11.4%	9.3%	9.1%	8.1%	8.2%	7.4%	7.3%
	174	191	195	211	165	185	177	176	177	159
Resistance ≥ 5 CLSI classes*	9.5%	10.3%	8.2%	9.8%	8.0%	7.2%	6.3%	6.9%	6.6%	6.3%
	131	145	164	182	142	146	137	149	157	137
At least ACSSuT <sup>†</sup>	8.9%	10.1%	7.8%	9.3%	7.2%	6.9%	5.6%	6.3%	5.8%	5.1%
	122	142	156	173	129	141	121	136	138	112
At least ACT/S <sup>‡</sup>	0.9%	0.5%	1.1%	1.2%	0.6%	0.9%	0.7%	0.7%	0.5%	0.7%
	13	7	21	23	10	18	15	16	11	15
At least ACSSuTAuCx <sup>§</sup>	2.6%	2.6%	3.4%	3.2%	2.4%	2.0%	2.0%	2.1%	1.8%	1.4%
	35	36	67	60	42	41	43	46	44	30
At least ceftriaxone and nalidixic acid resistant	0.1%	0.1%	0.2%	0.1%	0.1%	0.1%	0.1%	0.2%	0.0%	0.2%
	1	2	4	2	2	2	3	5	0	4

\* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

**Table 8. Twenty most common non-typhoidal *Salmonella* serotypes in NARMS, 2009**

NARMS		Isolates	
Rank	Serotype	n	(%)
1	Enteritidis	410	(18.7%)
2	Typhimurium	371	(16.9%)
3	Newport	236	(10.8%)
4	Javiana	105	(4.8%)
5	Heidelberg	86	(3.9%)
6	14,[5],12:i:-	72	(3.3%)
7	Oranienburg	64	(2.9%)
8	Saintpaul	57	(2.6%)
9	Montevideo	56	(2.6%)
10	Braenderup	46	(2.1%)
11	Infantis	44	(2.0%)
12	Muenchen	42	(1.9%)
13	Mississippi	28	(1.3%)
14	Thompson	27	(1.2%)
15	Agona	21	(1.0%)
16	Bareilly	20	(0.9%)
17	Litchfield	20	(0.9%)
18	Paratyphi B var. L(+) tartrate+	20	(0.9%)
19	Hadar	19	(0.9%)
20	Poona	16	(0.7%)
<b>Subtotal</b>		<b>1760</b>	<b>(80.3%)</b>
	All other serotypes	373	(17.0%)
	Unknown serotype	19	(0.9%)
	Partially serotyped	20	(0.9%)
	Rough/Nonmotile isolates	20	(0.9%)
<b>Subtotal</b>		<b>432</b>	<b>(19.7%)</b>
<b>Grand Total</b>		<b>2192</b>	<b>(100.0%)</b>



**Table 10. Percentage and number of *Salmonella ser. Enteritidis* isolates resistant to antimicrobial agents, 2000–2009**

Year			2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates			319	277	337	257	271	384	413	385	439	410
Rank*	CLSI† Antimicrobial Class	Antibiotic (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Gentamicin (MIC ≥ 16)	0.3%	0.0%	0.3%	0.4%	0.4%	0.8%	0.2%	0.0%	0.2%	0.0%
		Streptomycin (MIC ≥ 64)	0.0%	1.4%	1.5%	1.2%	2.2%	1.0%	1.2%	0.5%	0.5%	1.2%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	0.0%	1.4%	0.6%	0.0%	0.0%	0.8%	0.5%	0.5%	0.0%	0.0%
		Cephems	Ceftiofur (MIC ≥ 8)	0.0%	2.2%	0.0%	0.0%	0.0%	0.5%	0.5%	0.3%	0.0%
	Cephems	Ceftriaxone (MIC ≥ 64)	0.0%	1.4%	0.0%	0.0%	0.0%	0.3%	0.5%	0.3%	0.0%	0.0%
		Penicillins	Ampicillin (MIC ≥ 32)	7.5%	8.7%	6.8%	2.3%	4.1%	2.9%	4.4%	2.1%	3.6%
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Nalidixic acid (MIC ≥ 32)	2.2%	4.3%	3.9%	4.7%	6.6%	4.7%	7.0%	5.7%	6.6%	3.7%
	II	Aminoglycosides	Kanamycin (MIC ≥ 64)	0.3%	0.7%	0.3%	0.0%	0.7%	0.3%	0.2%	0.5%	0.0%
Cephems			Cefoxitin (MIC ≥ 32)	0.0%	0.4%	0.0%	0.0%	0.0%	1.0%	0.5%	0.3%	0.0%
Cephems		Cephalothin (MIC ≥ 32)	0.9%	1.1%	0.6%	1.2%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
		Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	0.9%	2.2%	1.5%	1.2%	1.8%	1.6%	1.5%	1.6%	1.1%
Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)			0.0%	0.7%	0.6%	0.8%	0.0%	0.5%	0.5%	1.0%	0.9%	0.7%
Phenicolis		Chloramphenicol (MIC ≥ 32)	0.0%	0.0%	0.3%	0.4%	0.4%	0.5%	0.0%	0.5%	0.5%	0.0%
		Tetracyclines	Tetracycline (MIC ≥ 16)	1.9%	1.8%	4.2%	1.6%	3.3%	2.3%	1.7%	3.9%	1.6%

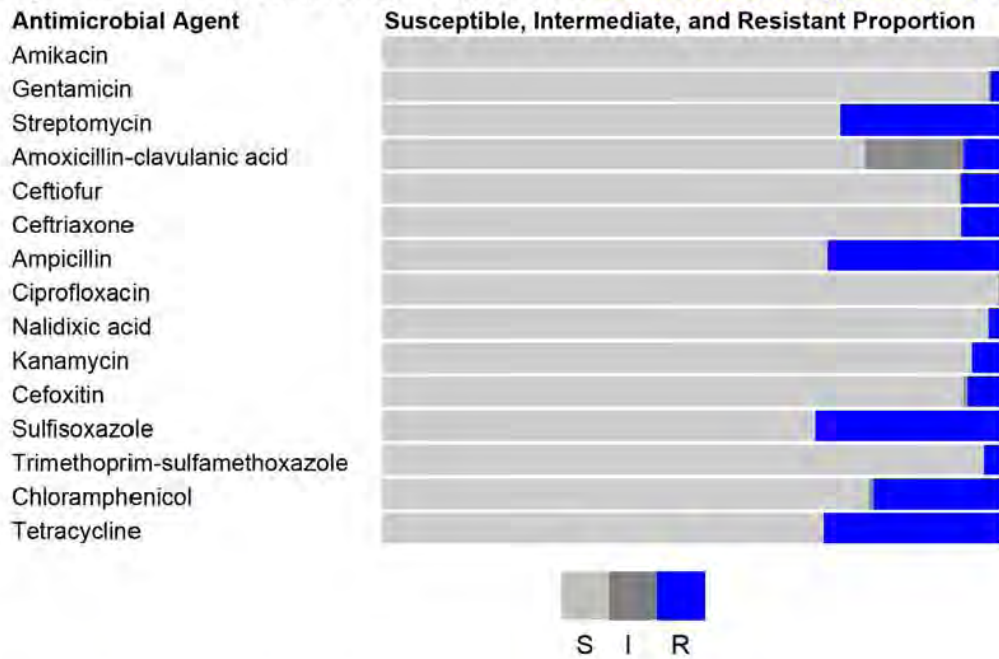
\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.



**Figure 15. Antimicrobial resistance pattern for *Salmonella ser. Typhimurium*, 2009**



**Table 13. Percentage and number of *Salmonella ser. Typhimurium* isolates resistant to antimicrobial agents, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009		
<b>Total Isolates</b>	<b>304</b>	<b>325</b>	<b>394</b>	<b>408</b>	<b>383</b>	<b>438</b>	<b>409</b>	<b>404</b>	<b>397</b>	<b>371</b>		
Rank <sup>*</sup>	CLSI <sup>†</sup> Antimicrobial Class	Antibiotic (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Gentamicin (MIC ≥ 16)	2.6%	1.5%	2.3%	2.0%	2.1%	1.8%	2.7%	2.5%	1.5%	1.9%
		Streptomycin (MIC ≥ 64)	39.5%	40.0%	32.0%	35.5%	31.9%	28.1%	29.3%	32.4%	28.5%	25.9%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	6.3%	6.2%	7.6%	5.6%	4.7%	3.2%	4.4%	6.7%	3.3%	6.2%
		Cephems	Ceftiofur (MIC ≥ 8)	3.6%	3.1%	4.3%	4.9%	4.4%	2.5%	4.2%	6.4%	3.3%
		Ceftriaxone (MIC ≥ 64)	3.3%	3.1%	4.3%	4.9%	4.4%	2.5%	4.2%	6.4%	3.3%	6.5%
	Penicillins	Ampicillin (MIC ≥ 32)	42.1%	42.5%	33.8%	36.3%	32.1%	29.0%	28.1%	31.7%	26.2%	28.0%
		Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.3%	0.0%	0.0%	0.0%	0.0%	0.2%	0.0%	0.0%
		Nalidixic acid (MIC ≥ 32)	1.3%	0.6%	1.3%	1.2%	0.5%	0.9%	0.7%	1.5%	1.3%	2.2%
	II	Aminoglycosides	Kanamycin (MIC ≥ 64)	13.2%	8.3%	7.6%	7.1%	5.7%	5.7%	5.1%	5.9%	2.3%
Cephems			Cefoxitin (MIC ≥ 32)	3.6%	3.1%	4.3%	4.4%	4.7%	2.5%	3.9%	5.7%	3.3%
		Cephalothin (MIC ≥ 32)	4.3%	3.1%	5.6%	6.1%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
Folate pathway inhibitors		Sulfamethoxazole/Sulfisoxazole <sup>‡</sup> (MIC ≥ 512)	45.4%	43.1%	32.2%	38.7%	36.0%	32.0%	33.3%	37.4%	30.2%	29.9%
		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	3.6%	2.5%	2.3%	3.4%	2.6%	2.7%	2.2%	2.5%	1.8%	3.0%
Phenicol		Chloramphenicol (MIC ≥ 32)	30.9%	31.7%	23.4%	28.2%	24.3%	24.4%	22.0%	25.5%	23.2%	20.5%
		Tetracyclines	Tetracycline (MIC ≥ 16)	43.4%	43.4%	32.0%	38.2%	30.3%	30.4%	31.5%	36.9%	27.5%

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

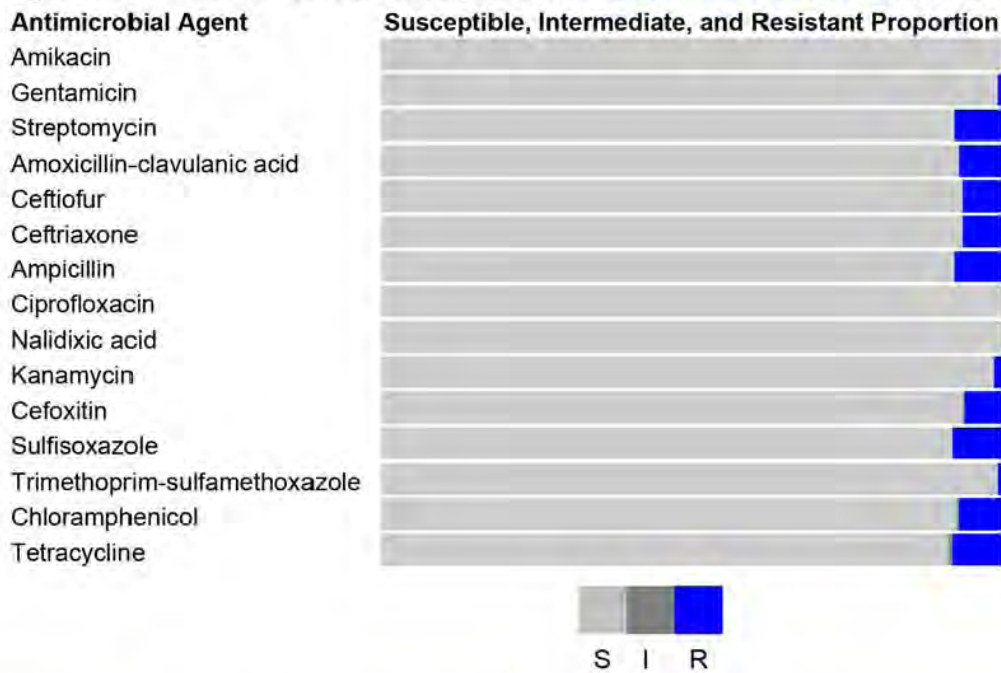
† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.





**Figure 16. Antimicrobial resistance pattern for *Salmonella ser. Newport*, 2009**



**Table 16. Percentage and number of *Salmonella ser. Newport* isolates resistant to antimicrobial agents, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009	
<b>Total Isolates</b>	<b>121</b>	<b>124</b>	<b>241</b>	<b>223</b>	<b>191</b>	<b>207</b>	<b>217</b>	<b>221</b>	<b>253</b>	<b>236</b>	
Rank*	CLSI† Antimicrobial Class	Antibiotic (Resistance breakpoint)									
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Gentamicin (MIC ≥ 16)	2.5%	3.2%	3.3%	3.1%	0.5%	1.0%	0.9%	0.9%	0.4%
		Streptomycin (MIC ≥ 64)	24.0%	31.5%	25.3%	24.2%	15.7%	14.0%	13.8%	10.4%	14.2%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	22.3%	26.6%	22.8%	21.5%	15.2%	12.8%	12.4%	8.1%	13.0%
		Cephems	Ceftiofur (MIC ≥ 8)	22.3%	27.4%	22.8%	22.0%	15.2%	12.6%	12.4%	8.1%
	Cephems	Ceftriaxone (MIC ≥ 64)	27	34	55	49	29	26	27	18	33
		Ampicillin (MIC ≥ 32)	23.1%	29.8%	24.9%	22.9%	15.7%	14.0%	15.2%	10.0%	15.0%
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Nalidixic acid (MIC ≥ 32)	0.8%	0.0%	0.8%	0.4%	0.5%	0.0%	0.5%	0.0%	0.4%
	II	Aminoglycosides	Kanamycin (MIC ≥ 64)	5.0%	7.3%	10.0%	4.5%	2.6%	1.9%	2.3%	0.9%
Cephems			Cefoxitin (MIC ≥ 32)	22.3%	25.8%	22.4%	21.5%	15.2%	12.6%	12.9%	8.1%
Cephems		Cephalothin (MIC ≥ 32)	27	32	54	48	29	26	28	18	33
		Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 5/12)	23.1%	32.3%	25.7%	24.7%	16.8%	15.5%	15.2%	10.4%
Folate pathway inhibitors		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	28	40	62	55	32	32	33	23	35
		Phenicol	Chloramphenicol (MIC ≥ 32)	4.1%	1.6%	4.1%	0.9%	2.1%	1.9%	3.2%	1.8%
Phenicol		Chloramphenicol (MIC ≥ 32)	5	2	10	2	4	4	7	4	8
		Tetracyclines	Tetracycline (MIC ≥ 16)	23.1%	28.2%	25.3%	22.4%	15.2%	13.5%	12.4%	9.5%
Tetracyclines		Tetracycline (MIC ≥ 16)	28	38	62	54	32	30	31	22	37

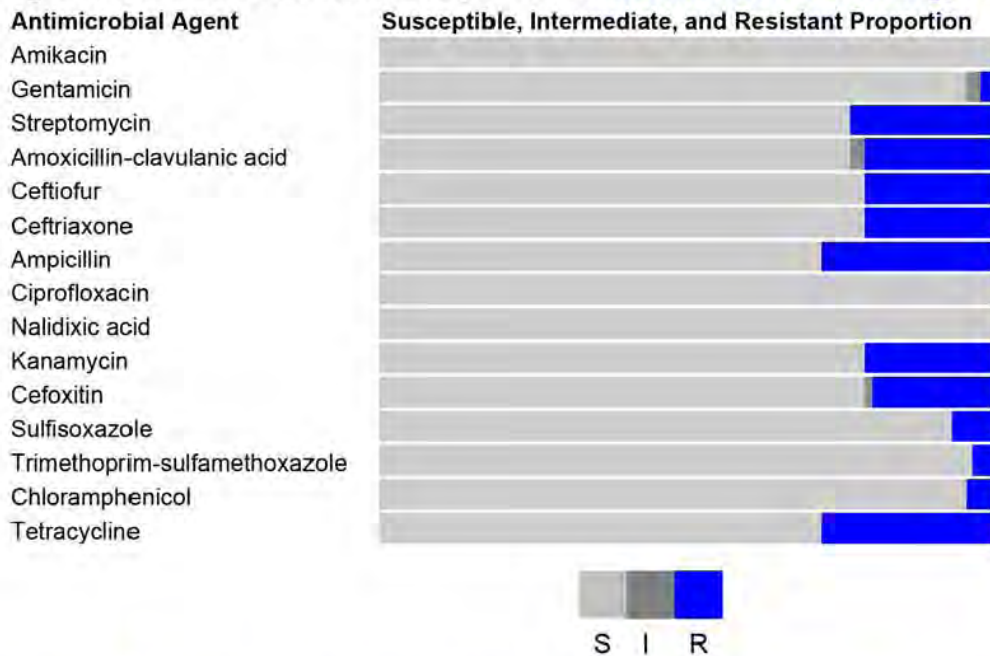
\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.



**Figure 17. Antimicrobial resistance pattern for *Salmonella ser. Heidelberg*, 2009**



**Table 19. Percentage and number of *Salmonella ser. Heidelberg* isolates resistant to antimicrobial agents, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009		
<b>Total Isolates</b>	<b>79</b>	<b>102</b>	<b>105</b>	<b>96</b>	<b>92</b>	<b>125</b>	<b>102</b>	<b>98</b>	<b>75</b>	<b>86</b>		
Rank*	CLSI† Antimicrobial Class	Antibiotic (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Gentamicin (MIC ≥ 16)	8.9%	7.8%	3.8%	5.2%	4.3%	6.4%	4.9%	16.3%	14.7%	2.3%
		Streptomycin (MIC ≥ 64)	22.8%	25.5%	17.1%	12.5%	15.2%	13.6%	11.8%	12.2%	30.7%	23.3%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	3.8%	2.9%	9.5%	5.2%	9.8%	8.8%	9.8%	7.1%	8.0%	20.9%
		Cephems	Ceftiofur (MIC ≥ 8)	3.8%	2.9%	7.6%	5.2%	8.7%	8.8%	9.8%	7.1%	8.0%
	Cephems	Ceftriaxone (MIC ≥ 64)	3.8%	2.9%	7.6%	5.2%	8.7%	8.8%	9.8%	7.1%	8.0%	20.9%
		Penicillins	Ampicillin (MIC ≥ 32)	10.1%	9.8%	12.4%	10.4%	25.0%	20.0%	18.6%	18.4%	28.0%
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Nalidixic acid (MIC ≥ 32)	1.3%	0.0%	0.0%	1.0%	0.0%	0.8%	0.0%	0.0%	0.0%	0.0%
	II	Aminoglycosides	Kanamycin (MIC ≥ 64)	15.2%	19.6%	10.5%	8.3%	8.7%	12.8%	8.8%	11.2%	26.7%
Cephems			Cefoxitin (MIC ≥ 32)	2.5%	2.9%	8.6%	5.2%	7.6%	8.8%	8.8%	7.1%	8.0%
Cephems		Cephalothin (MIC ≥ 32)	5.1%	3.9%	10.5%	7.3%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
		Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡	11.4%	8.8%	6.7%	7.3%	7.6%	8.0%	4.9%	18.4%	12.0%
Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)			1.3%	2.0%	1.0%	2.1%	0.0%	0.8%	0.0%	0.0%	2.7%	3.5%
Phenicol		Chloramphenicol (MIC ≥ 32)	1.3%	1.0%	1.0%	0.0%	1.1%	0.8%	0.0%	3.1%	1.3%	4.7%
		Tetracyclines	Tetracycline (MIC ≥ 16)	21.5%	24.5%	19.0%	16.7%	19.6%	18.4%	13.7%	22.4%	36.0%

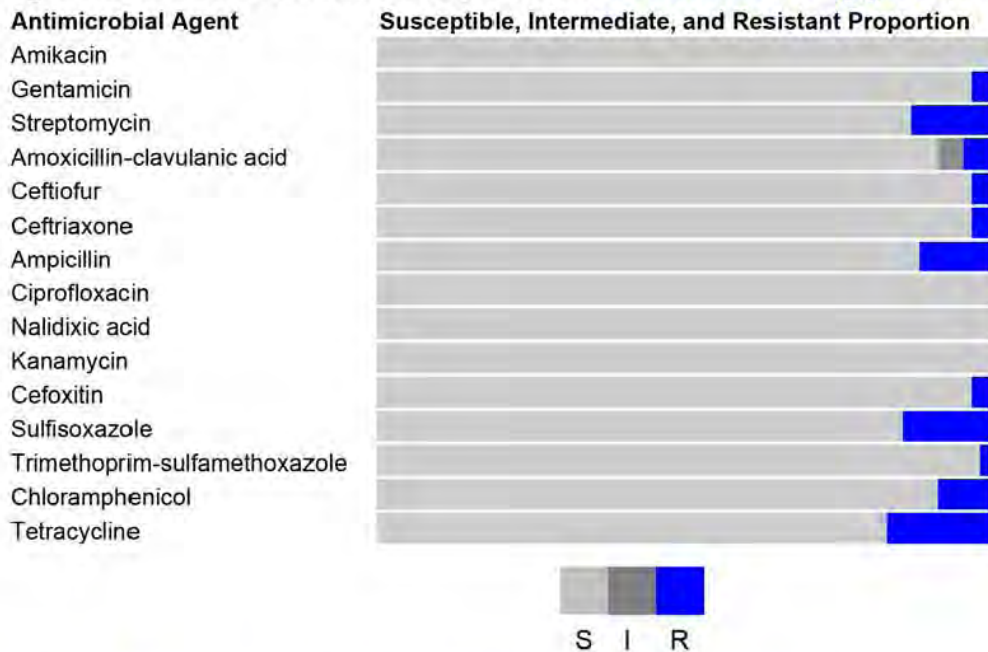
\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.



**Figure 18. Antimicrobial resistance pattern for *Salmonella ser. I 4,[5],12:i:-*, 2009**



**Table 22. Percentage and number of *Salmonella ser. I 4,[5],12:i:-* isolates resistant to antimicrobial agents, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009		
<b>Total Isolates</b>	<b>13</b>	<b>14</b>	<b>35</b>	<b>37</b>	<b>36</b>	<b>33</b>	<b>105</b>	<b>73</b>	<b>83</b>	<b>72</b>		
Rank*	CLSI† Antimicrobial Class	Antibiotic (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Gentamicin (MIC ≥ 16)	0.0%	7.1%	0.0%	5.4%	5.6%	0.0%	4.8%	1.4%	3.6%	2.8%
		Streptomycin (MIC ≥ 64)	7.7%	14.3%	2.9%	8.1%	5.6%	3.0%	3.8%	8.2%	10.8%	12.5%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	0.0%	0.0%	2.9%	5.4%	2.8%	3.0%	3.8%	1.4%	3.6%	4.2%
		Cephems	Ceftiofur (MIC ≥ 8)	0.0%	7.1%	2.9%	5.4%	2.8%	3.0%	3.8%	2.7%	3.6%
	Cephems	Ceftriaxone (MIC ≥ 64)	0.0%	0.0%	2.9%	5.4%	2.8%	3.0%	3.8%	2.7%	3.6%	2.8%
		Penicillins	Ampicillin (MIC ≥ 32)	7.7%	7.1%	8.6%	8.1%	5.6%	6.1%	6.7%	5.5%	8.4%
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Nalidixic acid (MIC ≥ 32)	0.0%	0.0%	0.0%	2.7%	2.8%	0.0%	1.0%	1.4%	1.2%	0.0%
	II	Aminoglycosides	Kanamycin (MIC ≥ 64)	0.0%	7.1%	0.0%	0.0%	0.0%	0.0%	1.4%	1.2%	0.0%
Cephems			Cefoxitin (MIC ≥ 32)	Not Tested	0.0%	2.9%	5.4%	2.8%	3.0%	3.8%	1.4%	3.6%
Cephems		Cephalothin (MIC ≥ 32)	0.0%	7.1%	2.9%	5.4%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
		Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	0.0%	14.3%	2.9%	5.4%	11.1%	0.0%	8.6%	4.1%	13.3%
Folate pathway inhibitors		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	0.0%	7.1%	2.9%	0.0%	2.8%	0.0%	0.0%	1.4%	4.8%	1.4%
		Phenicol	Chloramphenicol (MIC ≥ 32)	0.0%	7.1%	2.9%	0.0%	2.8%	0.0%	1.9%	1.4%	6.0%
Tetracyclines		Tetracycline (MIC ≥ 16)	7.7%	7.1%	5.7%	0.0%	11.1%	3.0%	8.6%	9.6%	16.9%	16.7%

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.

**Table 23. Resistance patterns of *Salmonella* ser. I 4,[5],12:i:- isolates, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates	13	14	35	37	36	33	105	73	83	72
	%	%	%	%	%	%	%	%	%	%
	n	n	n	n	n	n	n	n	n	n
No resistance detected	92.3%	78.6%	91.4%	78.4%	80.6%	87.9%	85.7%	82.2%	77.1%	76.4%
	12	11	32	29	29	29	90	60	64	55
Resistance ≥ 1 CLSI class*	7.7%	21.4%	8.6%	21.6%	19.4%	12.1%	14.3%	17.8%	22.9%	23.6%
	1	3	3	8	7	4	15	13	19	17
Resistance ≥ 2 CLSI classes*	7.7%	14.3%	8.6%	10.8%	13.9%	3.0%	11.4%	6.8%	16.9%	16.7%
	1	2	3	4	5	1	12	5	14	12
Resistance ≥ 3 CLSI classes*	7.7%	7.1%	5.7%	5.4%	8.3%	3.0%	9.5%	5.5%	9.6%	12.5%
	1	1	2	2	3	1	10	4	8	9
Resistance ≥ 4 CLSI classes*	0.0%	7.1%	2.9%	0.0%	2.8%	0.0%	3.8%	2.7%	7.2%	9.7%
	0	1	1	0	1	0	4	2	6	7
Resistance ≥ 5 CLSI classes*	0.0%	7.1%	2.9%	0.0%	2.8%	0.0%	2.9%	1.4%	4.8%	6.9%
	0	1	1	0	1	0	3	1	4	5
At least ACSSuT <sup>†</sup>	0.0%	7.1%	2.9%	0.0%	2.8%	0.0%	1.9%	1.4%	3.6%	6.9%
	0	1	1	0	1	0	2	1	3	5
At least ACT/S <sup>‡</sup>	0.0%	7.1%	2.9%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	0	1	1	0	0	0	0	0	0	0
At least ACSSuTAuCx <sup>§</sup>	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	2.4%	0.0%
	0	0	0	0	0	0	0	0	2	0
At least ceftriaxone and nalidixic acid resistant	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	0	0	0	0	0	0	0	0	0	0

\* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

## F. Specific Drug Resistance Phenotypes

**Table 24. Number and percentage of isolates with resistance to at least ACSSuT, ACSSuTAuCx, nalidixic acid, and ceftriaxone among the 20 most common non-typhoidal *Salmonella* serotypes isolated in NARMS, 2009**

Rank	Serotype	N	ACSSuT*		ACSSuTAuCx†		Nalidixic Acid		Ceftriaxone	
			n	(%)	n	(%)	n	(%)	n	(%)
1	Enteritidis	410	0	(0.0%)	0	(0.0%)	15	(38.5%)	0	(0.0%)
2	Typhimurium	371	72	(64.3%)	6	(20.0%)	8	(20.5%)	24	(32.0%)
3	Newport	236	15	(13.4%)	15	(50.0%)	0	(0.0%)	15	(20.0%)
4	Javiana	105	0	(0.0%)	0	(0.0%)	1	(2.6%)	1	(1.3%)
5	Heidelberg	86	3	(2.7%)	1	(3.3%)	0	(0.0%)	18	(24.0%)
6	14,[5],12:i:-	72	5	(4.5%)	0	(0.0%)	0	(0.0%)	2	(2.7%)
7	Oranienburg	64	0	(0.0%)	0	(0.0%)	0	(0.0%)	0	(0.0%)
8	Saintpaul	57	1	(0.9%)	0	(0.0%)	1	(2.6%)	0	(0.0%)
9	Montevideo	56	0	(0.0%)	0	(0.0%)	0	(0.0%)	0	(0.0%)
10	Braenderup	55	0	(0.0%)	0	(0.0%)	0	(0.0%)	0	(0.0%)
11	Infantis	44	2	(1.8%)	2	(6.7%)	1	(2.6%)	5	(6.7%)
12	Muenchen	42	0	(0.0%)	0	(0.0%)	0	(0.0%)	0	(0.0%)
13	Mississippi	28	1	(0.9%)	0	(0.0%)	0	(0.0%)	0	(0.0%)
14	Thompson	27	0	(0.0%)	0	(0.0%)	0	(0.0%)	0	(0.0%)
15	Agona	21	2	(1.8%)	2	(6.7%)	1	(2.6%)	2	(2.7%)
16	Bareilly	20	0	(0.0%)	0	(0.0%)	0	(0.0%)	0	(0.0%)
17	Litchfield	20	0	(0.0%)	0	(0.0%)	0	(0.0%)	0	(0.0%)
18	Paratyphi B var. L(+) tartrate+	20	2	(1.8%)	0	(0.0%)	0	(0.0%)	1	(1.3%)
19	Hadar	19	0	(0.0%)	0	(0.0%)	1	(2.6%)	0	(0.0%)
20	Poona	16	0	(0.0%)	0	(0.0%)	0	(0.0%)	0	(0.0%)
<b>Subtotal</b>		<b>1769</b>	<b>103</b>	<b>(92.0%)</b>	<b>26</b>	<b>(86.7%)</b>	<b>28</b>	<b>(71.8%)</b>	<b>68</b>	<b>(90.7%)</b>
	All other serotypes	364	5	(4.5%)	3	(10.0%)	8	(20.5%)	6	(8.0%)
	Unknown serotype	19	1	(0.9%)	1	(3.3%)	2	(5.1%)	1	(1.3%)
	Partially serotyped	20	2	(1.8%)	0	(0.0%)	1	(2.6%)	0	(0.0%)
	Rough/Nonmotile isolates	20	1	(0.9%)	0	(0.0%)	0	(0.0%)	0	(0.0%)
<b>Total</b>		<b>2192</b>	<b>112</b>	<b>(100.0%)</b>	<b>30</b>	<b>(100.0%)</b>	<b>39</b>	<b>(100.0%)</b>	<b>75</b>	<b>(100.0%)</b>

\* ACSSuT: at least resistant to ampicillin, chloramphenicol, streptomycin, sulfisoxazole, tetracycline

† ACSSuTAuCx: at least resistant to ACSSuT, amoxicillin-clavulanic acid, and ceftriaxone

## Box 2. Four major multidrug-resistant patterns among non-typhoidal *Salmonella* isolates, 1996–2009

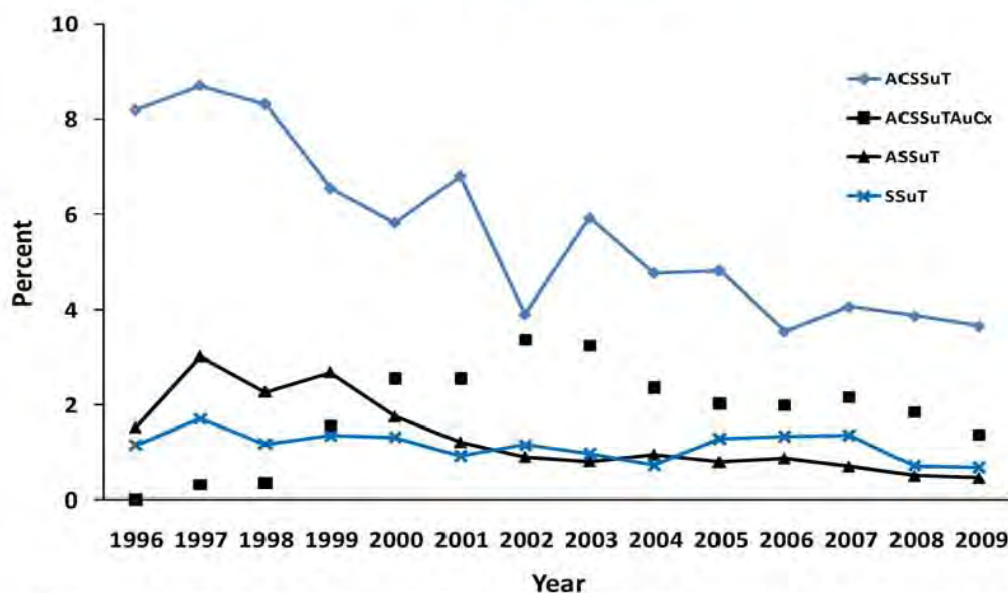
During 1996–2009, 3,243 (13.0%) of 24,903 non-typhoidal *Salmonella* isolates tested in NARMS were resistant to  $\geq 3$  classes of antimicrobial agents. Among the 3,243 isolates, we identified the 4 most common multidrug-resistant (MDR) patterns based on resistance to 7 of the 15 agents currently tested in NARMS: ampicillin (A), chloramphenicol (C), streptomycin (S), sulfonamide (Su), tetracycline (T), amoxicillin-clavulanic acid (Au), and ceftriaxone (Cx). Unlike MDR criteria used for tables in the rest of this report, which defined “at least ACSSuT” as resistant to at least A, C, S, Su, and T, we used mutually exclusive criteria in this section: ACSSuT was defined as resistant to A, C, S, Su, and T, not resistant to Au and Cx; ACSSuTAuCx as resistant to all 7 agents; ASSuT as resistant to A, S, Su, and T, not resistant to C, Au, and Cx; and SSuT as resistant to S, Su, and T, not resistant to A, C, Au, and Cx. Use of mutually exclusive criteria is important in monitoring major and emerging patterns, which may be driven by different resistance mechanisms.

ACSSuT, ACSSuTAuCx, ASSuT, and SSuT isolates accounted for 73.1% of the 3,243 isolates resistant to  $\geq 3$  classes: 1,323 (40.8%) were ACSSuT, 476 (14.7%) ACSSuTAuCx, 295 (9.1%) ASSuT, and 275 (8.5%) SSuT. Serotype Typhimurium accounted for 90.0% of ACSSuT and 74.6% of ASSuT isolates. Of the ACSSuTAuCx isolates, 67.0% were serotype Newport and 19.8% were serotype Typhimurium. More than half of SSuT isolates were serotypes Typhimurium (26.6%), Stanley (12.7%), Derby (10.2%), and Heidelberg (9.8%).

Figure 1 below shows the percentage of the 4 MDR patterns by year among all non-typhoidal *Salmonella* isolates tested from 1996 through 2009. ACSSuT declined from 8.2% in 1996 to 3.7% in 2009. ACSSuTAuCx, first detected in 1997, peaked at 3.4% in 2002 before declining to 1.4% in 2009. ASSuT was  $\leq 3.0\%$  and SSuT was  $\leq 1.7\%$  from 1996 through 2009.

Refer to Table 7 in the Results section of this report for the percentage of “at least ACSSuT” by year (2000–09) among non-typhoidal *Salmonella* isolates, which includes ACSSuT and ACSSuTAuCx. Since “at least ACSSuTAuCx” in Table 7 includes resistance to all 7 agents, percentages are the same as in Figure 1.

**Figure 1. Percentage of 4 major MDR patterns\* among non-typhoidal *Salmonella* isolates, by year, 1996–2009**



\*Four most common multidrug-resistant (MDR) patterns that include resistance to  $\geq 3$  antimicrobial classes and any of the following agents: ampicillin (A), chloramphenicol (C), streptomycin (S), sulfonamide (Su), tetracycline (T), amoxicillin-clavulanic acid (Au), and ceftriaxone (Cx).





**Table 26. Percentage and number of *Salmonella* ser. Typhi isolates resistant to antimicrobial agents, 2000–2009**

Year			2000	2001	2002	2003	2004	2005	2006	2007	2008	2009		
Total Isolates			177	197	195	332	304	318	322	401	410	361		
Rank <sup>†</sup>	CLSI <sup>†</sup> Antimicrobial Class	Antibiotic (Resistance breakpoint)												
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Gentamicin (MIC ≥ 16)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Streptomycin (MIC ≥ 64)	9.0%	20.3%	7.2%	14.5%	11.8%	13.2%	18.9%	15.7%	11.5%	10.5%	10.5%	
	β-lactam / β-lactamase inhibitor combinations	Amoxicillin-Clavulanic Acid (MIC ≥ 32)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.3%	0.2%	0.0%	0.3%	0.3%	
		Cephems	Ceftiofur (MIC ≥ 8)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	Penicillins	Ceftriaxone (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Ampicillin (MIC ≥ 32)	9.0%	20.3%	5.6%	16%	11.8%	13.2%	20.5%	17%	13.2%	12.2%	12.2%	
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.3%	0.0%	0.3%	0.9%	1.0%	0.0%	3.3%	3.3%	
		Nalidixic Acid (MIC ≥ 32)	22%	29.9%	23.6%	37.7%	41.8%	48.4%	54.7%	61.8%	59.0%	60.1%	60.1%	
			39	59	46	125	127	154	176	248	242	217	217	
	II	Aminoglycosides	Kanamycin (MIC ≥ 64)	0.0%	0.5%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
			Cephems	Cefoxitin (MIC ≥ 32)	0.6%	0.5%	0.0%	0.0%	0.0%	0.0%	0.3%	0.5%	0.0%	0.0%
Folate pathway inhibitors		Cephalothin (MIC ≥ 32)	1.1%	0.5%	1.5%	0.0%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	
		Sulfisoxazole (MIC ≥ 512)	11.3%	20.8%	6.2%	16.9%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
Phenicol		Trimethoprim-Sulfamethoxazole (MIC ≥ 4)	9.0%	20.8%	6.7%	16.9%	13.2%	14.5%	20.8%	16.2%	12.7%	12.2%	12.2%	
		Chloramphenicol (MIC ≥ 32)	10.7%	20.8%	6.2%	16.6%	13.2%	13.2%	19.6%	15.7%	12.9%	11.4%	11.4%	
Tetracyclines		Tetracycline (MIC ≥ 16)	9.6%	20.8%	6.7%	15.4%	8.9%	10.1%	8.4%	6.2%	4.6%	5.5%	5.5%	
			17	41	13	51	27	32	27	25	19	20	20	

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table 1): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.

**Table 27. Resistance patterns of *Salmonella* ser. Typhi isolates, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates	177	197	195	332	304	318	323	401	410	361
	%	%	%	%	%	%	%	%	%	%
	n	n	n	n	n	n	n	n	n	n
No resistance detected	72.3%	58.9%	74.4%	56.6%	56.6%	48.1%	40.2%	35.7%	38.0%	37.7%
	128	116	145	188	172	153	130	143	156	136
Resistance ≥ 1 CLSI class*	27.7%	41.1%	25.6%	43.4%	43.4%	51.9%	59.8%	64.3%	62.0%	62.3%
	49	81	50	144	132	165	193	258	254	225
Resistance ≥ 2 CLSI classes*	10.7%	22.8%	7.2%	17.5%	13.2%	14.5%	21.7%	18.0%	14.4%	14.1%
	19	45	14	58	40	46	70	72	59	51
Resistance ≥ 3 CLSI classes*	9.6%	21.8%	6.7%	16.6%	12.8%	13.8%	20.7%	17.5%	13.4%	12.7%
	17	43	13	55	39	44	67	70	55	46
Resistance ≥ 4 CLSI classes*	9.0%	21.3%	6.2%	16.3%	12.5%	12.9%	19.2%	17.0%	12.9%	12.2%
	16	42	12	54	38	41	62	68	53	44
Resistance ≥ 5 CLSI classes*	7.9%	16.8%	5.6%	14.2%	11.8%	11.9%	16.7%	14.7%	10.7%	10.0%
	14	33	11	47	36	38	54	59	44	36
At least ACSSuT <sup>†</sup>	7.9%	16.8%	5.6%	12.7%	7.9%	9.1%	5.9%	3.7%	2.4%	2.5%
	14	33	11	42	24	29	19	15	10	9
At least ACT/S <sup>‡</sup>	9.0%	17.8%	5.6%	15.7%	11.8%	12.9%	18.6%	15.2%	12.2%	10.5%
	16	35	11	52	36	41	60	61	50	38
At least ACSSuTAuCx <sup>§</sup>	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	0	0	0	0	0	0	0	0	0	0
At least ceftriaxone and nalidixic acid resistant	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	0	0	0	0	0	0	0	0	0	0

\* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone



**Table 30. Percentage and number of *Salmonella* ser. Paratyphi A, Paratyphi B, and Paratyphi C isolates resistant to antimicrobial agents, 2000–2009**

Year			2000	2001	2002	2003	2004	2005	2006	2007	2008	2009	
Total Isolates			5	9	10	8	11	18	16	17	92	102	
Rank*	CLSI† Antimicrobial Class	Antibiotic (Resistance breakpoint)											
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Gentamicin (MIC ≥ 16)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Streptomycin (MIC ≥ 64)	20.0%	0.0%	10.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	1.0%
	β lactam/β lactamase inhibitor combinations	Amoxicillin clavulanic acid (MIC ≥ 32/16)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	0.0%
			0	0	0	0	0	0	0	0	0	1	0
	Cephems	Ceftiofur (MIC ≥ 8)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	0.0%
		Ceftriaxone (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	0.0%
	Penicillins	Ampicillin (MIC ≥ 32)	20.0%	0.0%	0.0%	12.5%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	1.0%
			1	0	0	1	0	0	0	0	0	1	1
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
Nalidixic acid (MIC ≥ 32)		40.0%	55.6%	40.0%	75.0%	72.7%	66.7%	50.0%	94.1%	85.9%	86.3%		
		2	5	4	6	8	12	8	16	79	88		
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
			0	0	0	0	0	0	0	0	0	0	
	Cephems	Cefoxitin (MIC ≥ 32)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	0.0%
		Cephalothin (MIC ≥ 32)	0.0%	0.0%	0.0%	0.0%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
	Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	20.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	1.0%
		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	20.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.0%
			1	0	0	0	0	0	0	0	0	1	
	Phenicol	Chloramphenicol (MIC ≥ 32)	20.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	1.0%
			1	0	0	0	0	0	0	0	0	1	1
	Tetracyclines	Tetracycline (MIC ≥ 16)	0.0%	0.0%	10.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	2.2%	1.0%
		0	0	1	0	0	0	0	0	0	2	1	

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.

**Table 31. Resistance patterns of *Salmonella* ser. Paratyphi A, Paratyphi B, and Paratyphi C isolates, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates	5	9	10	8	11	18	16	17	92	102
	%	%	%	%	%	%	%	%	%	%
	n	n	n	n	n	n	n	n	n	n
No resistance detected	40.0%	44.4%	50.0%	12.5%	27.3%	33.3%	50.0%	5.9%	12.0%	12.7%
	2	4	5	1	3	6	8	1	11	13
Resistance ≥ 1 CLSI class*	60.0%	55.6%	50.0%	87.5%	72.7%	66.7%	50.0%	94.1%	88.0%	87.3%
	3	5	5	7	8	12	8	16	81	89
Resistance ≥ 2 CLSI classes*	20.0%	0.0%	10.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	1.0%
	1	0	1	0	0	0	0	0	1	1
Resistance ≥ 3 CLSI classes*	20.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	1.0%
	1	0	0	0	0	0	0	0	1	1
Resistance ≥ 4 CLSI classes*	20.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	1.0%
	1	0	0	0	0	0	0	0	1	1
Resistance ≥ 5 CLSI classes*	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	1.0%
	0	0	0	0	0	0	0	0	1	1
At least ACSSuT†	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	1.0%
	0	0	0	0	0	0	0	0	1	1
At least ACT/S‡	20.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.0%
	1	0	0	0	0	0	0	0	0	1
At least ACSSuTAuCx§	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.1%	0.0%
	0	0	0	0	0	0	0	0	1	0
At least ceftriaxone and nalidixic acid resistant	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	0	0	0	0	0	0	0	0	0	0

\* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

### Box 3. Molecular characterization of Non-Typhi *Salmonella enterica* that show decreased susceptibility to extended-spectrum cephalosporins

Although most *Salmonella* infections are self-limited and treated symptomatically, antimicrobial therapy is necessary for the management of invasive infections. The recommended regimen used to include either amoxicillin or trimethoprim-sulphamethoxazole, but due to increased resistance levels to these drugs, current recommendations suggest using a fluoroquinolone, such as ciprofloxacin, or an extended-spectrum cephalosporin, such as ceftriaxone.

Among 4,236 isolates of non-typhi *Salmonella* (NTS) submitted to NARMS in 2005 and 2006, 175 (4.1%) displayed decreased susceptibility (MIC  $\geq$  2 mg/L) to the extended-spectrum cephalosporins ceftriaxone or ceftiofur. Among these, thirty different serotypes were represented and the three most prevalent serotypes were Newport (33.1%), Typhimurium (13.7%) and Heidelberg (13.1%).

Among the 175 isolates, 172 were available for molecular analysis. For each isolate, genomic DNA was prepared and the presence of  $\beta$ -lactamase genes investigated by polymerase chain reaction (PCR) amplification targeting six different genes: *bla*<sub>TEM</sub>, *bla*<sub>OXA</sub>, *bla*<sub>SHV</sub>, *bla*<sub>CTX-M</sub>, *bla*<sub>PSE</sub> and *bla*<sub>CMY</sub>.

One or more  $\beta$ -lactamase genes were detected in 139 (80.8%) isolates. The most prevalent resistance mechanisms were AmpC  $\beta$ -lactamase genes (*bla*<sub>CMY</sub>) which were identified in 133 (95.7%) of the 139  $\beta$ -lactamase-carrying isolates. The ceftriaxone MIC-values for the *bla*<sub>CMY</sub>-containing isolates ranged from 4 to 64 mg/L; all *bla*<sub>CMY</sub>-bearing isolates were thereby classified as ceftriaxone resistant according to current CLSI guidelines.

Other  $\beta$ -lactamase genes detected included eleven *bla*<sub>TEM-1</sub>, three *bla*<sub>PSE-1</sub>, two *bla*<sub>OXA-1</sub> and one *bla*<sub>CTX-M-15</sub>. The latter is an extended-spectrum  $\beta$ -lactamase (ESBL) that exhibits increased activity against cefotaxime and confers cross-resistance to the fourth generation cephalosporin cefepime. The *bla*<sub>CTX-M-15</sub> gene was found in an isolate of serotype Concord and is the second CTX-M-producing *Salmonella* identified in the NARMS human isolate collection (the first case was a CTX-M-5-producing *Salmonella* ser. Typhimurium isolated from a 3-month old child in 2003). The source of the CTX-M-15-producing *Salmonella* remains unknown, but it is likely that the infection was acquired abroad since the patient reported international travel to Ethiopia in conjunction with illness onset.

In conclusion, among NTS isolated from humans in 2005-2006, CMY  $\beta$ -lactamases were the predominant cause of decreased susceptibility and resistance to extended-spectrum cephalosporins. To limit further spread of these genes, prudent use of antimicrobial agents in both human and veterinary medicine will be crucial. Continued surveillance for cephalosporin-resistant bacteria among humans remains critical.

#### Box 4. Reduced azithromycin susceptibility in *Shigella sonnei*, United States

*Shigella* spp. are a major cause of gastroenteritis in the United States, transmitted most commonly by the fecal-oral route. It occurs most often in children 5 years old and younger in child care settings. While treatment with antimicrobial agents may shorten the duration of clinical symptoms for this usually self-limiting disease, emergence of antimicrobial resistance has recently made drug selection difficult. Previous drugs of choice for treatment of shigellosis were ampicillin and trimethoprim-sulfamethoxazole, but resistance to these drugs is high, especially in *Shigella sonnei*. Because of resistance to these agents, the macrolide azithromycin has been recommended by the American Academy of Pediatrics for treatment of shigellosis in children. Azithromycin resistance in *Shigella sonnei* is now being reported, but there are currently no CLSI criteria for interpretation of azithromycin susceptibility test results for *Shigella*. To explore azithromycin susceptibility in the United States, we determined MIC distributions for *Shigella sonnei* isolated from humans in the United States.

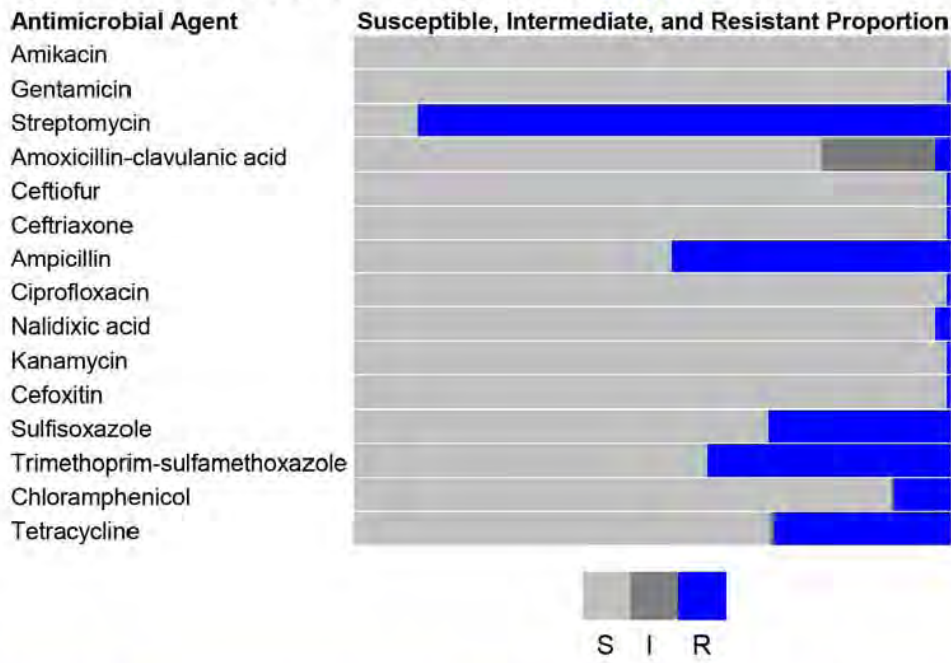
Azithromycin MICs were determined by broth microdilution for *Shigella sonnei* isolates from 11 recognized outbreaks in 10 states in 2006-2007 (n=56) and for those submitted routinely to NARMS in 2005 (the most recently completed year at the time, n=336) from 44 states. Five isolates collected from 1969-1974 (before approval of azithromycin) were also tested for historical perspective, but were not included in the MIC distribution analysis. Each isolate that demonstrated an azithromycin MIC > 64 mg/L was screened for the *mphA* gene by PCR, and the gene was sequenced to confirm positives.

The resulting distribution was log-normal and spanned three doubling dilution steps (4-16 mg/L) with an MIC<sub>50</sub> and MIC<sub>90</sub> of 8 mg/L. Five isolates collected from 1969-1974 exhibited azithromycin MICs of either 4 mg/L (n=1) or 8 mg/L (n=4). One outbreak isolate and two routine isolates showed MICs at least four-fold higher than the MIC<sub>90</sub> for the overall group of isolates; the outbreak isolate displayed an MIC of 256 mg/L and the routine isolates showed MICs of 256 and 128 mg/L. These isolates were PCR-positive for *mphA*, which encodes a macrolide-2'-phosphotransferase. In all three isolates, the gene was on a transferrable plasmid, with plasmid sizes ranging from 15-150kb.

Decreased azithromycin susceptibility for *Shigella sonnei* in the face of high levels of ampicillin and trimethoprim-sulfamethoxazole resistance is particularly worrisome for children because the remaining options for treatment are ceftriaxone, which is administered by injection and fluoroquinolones such as ciprofloxacin, which are not approved for use in children. Continued surveillance will be needed to examine antimicrobial use practices for *Shigella* infections and to determine whether azithromycin use leads to decreased susceptibility to the agent. Clinical outcome studies are needed to establish clinical breakpoints for azithromycin in treating *Shigella*.



**Figure 21. Antimicrobial resistance pattern for *Shigella*, 2009**



**Table 34. Percentage and number of *Shigella* isolates resistant to antimicrobial agents, 2000–2009**

Year			2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates			450	344	620	495	316	396	402	480	549	475
Rank*	CLSI† Antimicrobial Class	Antibiotic (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Gentamicin (MIC ≥ 16)	0.2%	0.0%	0.2%	0.0%	0.0%	1.0%	0.2%	0.8%	0.4%	0.6%
		Streptomycin (MIC ≥ 64)	57.1%	53.2%	54.4%	57.0%	59.8%	68.7%	60.7%	73.3%	80.7%	89.1%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	2.2%	4.4%	2.6%	1.4%	1.6%	1.0%	1.5%	0.4%	3.3%	2.1%
		Cephems										
	Cephems	Ceftiofur (MIC ≥ 8)	0.0%	0.0%	0.2%	0.2%	0.3%	0.5%	0.2%	0.0%	0.0%	0.6%
		Ceftriaxone (MIC ≥ 64)	0.0%	0.0%	0.2%	0.2%	0.3%	0.5%	0.2%	0.0%	0.0%	0.6%
	Penicillins	Ampicillin (MIC ≥ 32)	79.1%	79.7%	76.6%	79.4%	77.5%	70.7%	62.4%	63.8%	62.5%	46.3%
		Quinolones										
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.3%	0.0%	0.0%	0.0%	0.0%	0.2%	0.2%	0.7%	0.6%
Nalidixic acid (MIC ≥ 32)		0.9%	1.7%	1.6%	1.0%	1.6%	1.5%	3.5%	1.7%	1.6%	2.1%	
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	1.3%	0.6%	0.8%	0.4%	0.0%	0.8%	0.0%	0.2%	0.5%	0.4%
		Cephems										
	Cephems	Cefoxitin (MIC ≥ 32)	0.2%	1.2%	0.3%	0.0%	0.3%	0.3%	0.0%	0.0%	0.0%	0.6%
		Cephalothin (MIC ≥ 32)	8.0%	9.0%	6.6%	9.3%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
	Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	55.8%	56.4%	31.8%	33.0%	52.5%	57.6%	40.3%	25.8%	28.6%	30.5%
		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	52.9%	46.8%	37.3%	38.6%	46.8%	53.3%	46.0%	25.8%	31.3%	40.4%
	Phenicol	Chloramphenicol (MIC ≥ 32)	14.0%	21.5%	7.6%	8.5%	15.2%	10.9%	10.9%	8.3%	6.9%	9.3%
		Tetracyclines										
	Tetracyclines	Tetracycline (MIC ≥ 16)	44.9%	59.3%	30.6%	29.1%	49.4%	38.4%	34.6%	25.6%	24.2%	29.5%

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.



**Table 35. Resistance patterns of *Shigella* isolates, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates	450	344	620	495	316	396	402	480	549	475
	%	%	%	%	%	%	%	%	%	%
	n	n	n	n	n	n	n	n	n	n
No resistance detected	7.3%	4.9%	8.2%	8.5%	4.7%	4.5%	6.5%	7.1%	4.4%	4.0%
	33	17	51	42	15	18	26	34	24	19
Resistance ≥ 1 CLSI class*	92.7%	95.1%	91.8%	91.5%	95.3%	95.5%	93.5%	92.9%	95.6%	96.0%
	417	327	569	453	301	378	376	446	525	456
Resistance ≥ 2 CLSI classes*	64.7%	68.6%	55.2%	57.8%	64.2%	71.7%	64.7%	65.4%	68.3%	67.8%
	291	236	342	286	203	284	260	314	375	322
Resistance ≥ 3 CLSI classes*	61.3%	59.9%	41.5%	40.2%	59.2%	58.6%	43.8%	27.7%	35.2%	36.4%
	276	206	257	199	187	232	176	133	193	173
Resistance ≥ 4 CLSI classes*	31.8%	45.3%	24.4%	24.8%	32.9%	19.4%	15.4%	11.7%	10.4%	13.3%
	143	156	151	123	104	77	62	56	57	63
Resistance ≥ 5 CLSI classes*	6.4%	7.6%	2.7%	3.6%	7.0%	4.8%	5.2%	4.6%	2.7%	6.3%
	29	26	17	18	22	19	21	22	15	30
At least ACSSuT†	5.6%	6.4%	1.8%	3.2%	6.0%	4.0%	5.0%	3.8%	2.2%	5.9%
	25	22	11	16	19	16	20	18	12	28
At least ACT/S‡	6.9%	7.0%	2.7%	3.6%	6.6%	6.3%	6.0%	4.0%	2.9%	6.7%
	31	24	17	18	21	25	24	19	16	32
At least AT/S§	44.4%	37.5%	29.8%	33.7%	34.5%	35.6%	26.6%	12.9%	16.0%	17.5%
	200	129	185	167	109	141	107	62	88	83
At least ANT/S¶	0.0%	0.6%	0.3%	0.8%	0.6%	0.5%	0.5%	0.8%	0.0%	0.2%
	0	2	2	4	2	2	2	4	0	1
At least ACSSuTAuCx**	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	0	0	0	0	0	0	0	0	0	0
At least ceftriaxone and nalidixic acid resistant	0.0%	0.0%	0.0%	0.2%	0.3%	0.3%	0.2%	0.0%	0.0%	0.0%
	0	0	0	1	1	1	1	0	0	0

\* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ AT/S: resistance to ampicillin, trimethoprim-sulfamethoxazole

¶ ANT/S: resistance to AT/S, nalidixic acid

\*\* ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

**Table 36. Minimum inhibitory concentrations (MICs) and resistance of *Shigella sonnei* isolates to antimicrobial agents, 2009 (N=410)**

Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent	% of isolates			Percent of all isolates with MIC (µg/mL)‡																		
			%I§	%R¶	[95% CI]¶	0.015	0.03	0.06	0.125	0.25	0.50	1	2	4	8	16	32	64	128	256	512			
I	Aminoglycosides	Amikacin	0.0	0.0	[0.0 - 0.9]						0.5	2.0	35.1	59.5	2.9									
		Gentamicin	0.0	0.7	[0.1 - 2.1]					1.5	14.9	78.0	4.4	0.5		0.2	0.5							
		Streptomycin	N/A	91.5	[88.3 - 94.0]												8.5	48.3	43.2					
	β-lactam / β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid	14.6	2.0	[0.8 - 3.6]							2.4	2.7	50.5	27.8	14.6	1.5	0.5						
		Cephems							5.6	83.4	8.3	2.2			0.5									
		Ceftriaxone	0.0	0.5	[0.05 - 1.7]					99.3	0.2				0.5									
		Penicillins										4.4	47.8	4.1	0.5				0.2	42.9				
	Quinolones							98.0	0.2	1.0	0.2	0.5												
		Nalidixic acid	N/A	1.7	[0.7 - 3.5]						4.6	83.7	8.3	1.7				1.0	0.7					
II	Aminoglycosides	Kanamycin	0.0	0.2	[0.00 - 1.3]										99.5	0.2						0.2		
	Cephems	Cefoxitin	0.0	0.7	[0.1 - 2.1]						0.5	18.8	73.7	5.9	0.5			0.5	0.2					
	Folate pathway inhibitors	Sulfisoxazole	N/A	23.9	[19.9 - 28.3]											65.6	7.1	2.2	1.2				23.9	
		Trimethoprim-sulfamethoxazole	N/A	36.1	[31.4 - 41.0]				5.6	1.5	5.9	21.5	29.5	10.2	25.9									
		Phenolics	Chloramphenicol	0.0	1.2	[0.4 - 2.8]								20.2	76.3	2.2					1.2			
	Tetracyclines	Tetracycline	0.5	20.7	[18.9 - 25.0]										78.8	0.5	0.2	9.0	11.5					

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

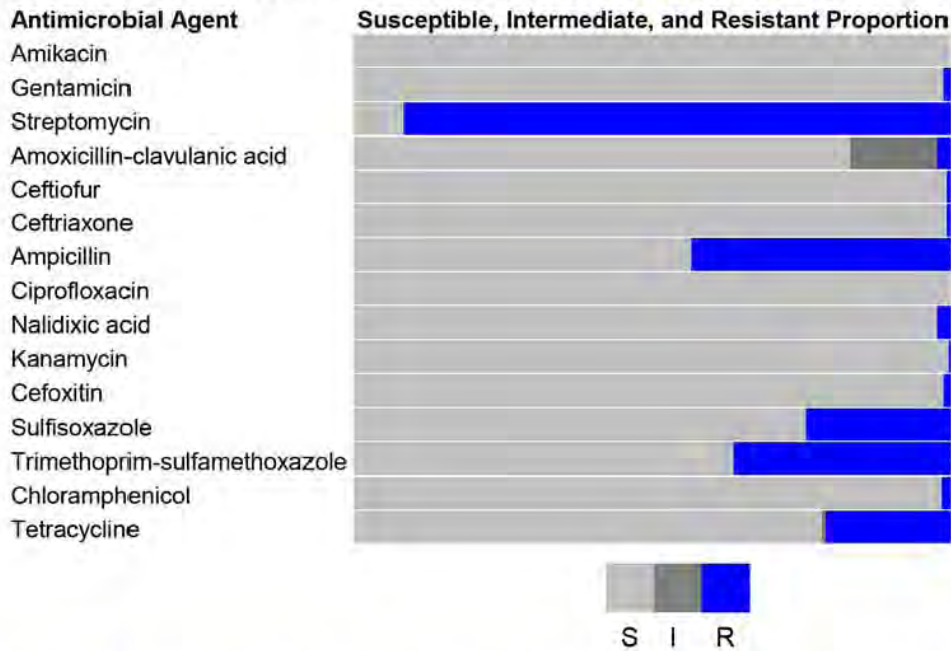
‡ Percent of isolates with intermediate susceptibility, N/A if no MIC range of intermediate susceptibility exists

§ Percent of isolates that were resistant

¶ The 95% confidence intervals (CI) for percent resistant (%R) were calculated using the Poisson-Camp-Pratt approximation to the Copper-Harison exact method. The 95% CI is presented to summarize uncertainty in the observed resistance (R%).

\*\* The unshaded areas indicate the dilution range of the Sensititre plates used to test isolates. Single vertical bars indicate the breakpoints for susceptibility, while double vertical bars indicate breakpoints for resistance. Numbers in the shaded areas indicate the percentages of isolates with MICs greater than the highest concentrations on the Sensititre plate. Numbers listed for the lowest tested concentrations represent the percentages of isolates with MICs equal to or less than the lowest tested concentration. CLSI breakpoints were used when available.

**Figure 22. Antimicrobial resistance pattern for *Shigella sonnei*, 2009**



**Table 37. Percentage and number of *Shigella sonnei* isolates resistant to antimicrobial agents, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009		
<b>Total Isolates</b>	<b>366</b>	<b>239</b>	<b>536</b>	<b>434</b>	<b>241</b>	<b>340</b>	<b>321</b>	<b>414</b>	<b>496</b>	<b>410</b>		
Rank*	CLSI† Antimicrobial Class	Antibiotic (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Gentamicin (MIC ≥ 16)	0.3%	0.0%	0.0%	0.0%	0.0%	1.2%	0.0%	1.0%	0.4%	
		Streptomycin (MIC ≥ 64)	56.0%	54.0%	55.4%	56.5%	56.8%	70.3%	61.7%	76.8%	82.5%	
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	1.9%	4.6%	2.2%	1.4%	1.7%	1.2%	1.9%	0.5%	3.2%	
		Cephems	Ceftiofur (MIC ≥ 8)	0.0%	0.0%	0.0%	0.0%	0.4%	0.6%	0.0%	0.0%	0.5%
	Ceftriaxone (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.4%	0.6%	0.0%	0.0%	0.0%	0.5%	
		Penicillins	Ampicillin (MIC ≥ 32)	80.6%	82.8%	77.6%	79.7%	79.3%	70.6%	62.8%	64.0%	61.5%
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.6%	0.0%	
		Nalidixic acid (MIC ≥ 32)	1.1%	0.8%	1.5%	0.5%	1.7%	1.2%	2.8%	1.2%	1.6%	
	II	Aminoglycosides	Kanamycin (MIC ≥ 64)	1.6%	0.4%	0.4%	0.0%	0.0%	0.0%	0.0%	0.2%	0.6%
			Cephems	Cefoxitin (MIC ≥ 32)	0.3%	1.7%	0.4%	0.0%	0.4%	0.3%	0.0%	0.0%
		Cephalothin (MIC ≥ 32)	8.7%	12.6%	7.3%	10.1%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
			Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	56.0%	54.4%	29.9%	31.3%	49.0%	57.9%	33.3%	20.0%
Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)		54.9%		50.6%	37.9%	38.5%	46.9%	55.0%	42.7%	22.0%	29.4%	
Phenicol		Chloramphenicol (MIC ≥ 32)		2.7%	1.3%	0.2%	1.2%	2.5%	2.4%	0.9%	1.2%	1.0%
Tetracyclines		Tetracycline (MIC ≥ 16)	34.4%	44.8%	23.5%	22.1%	36.1%	29.4%	22.7%	16.2%	17.3%	
		126	107	126	96	87	100	73	87	86	85	

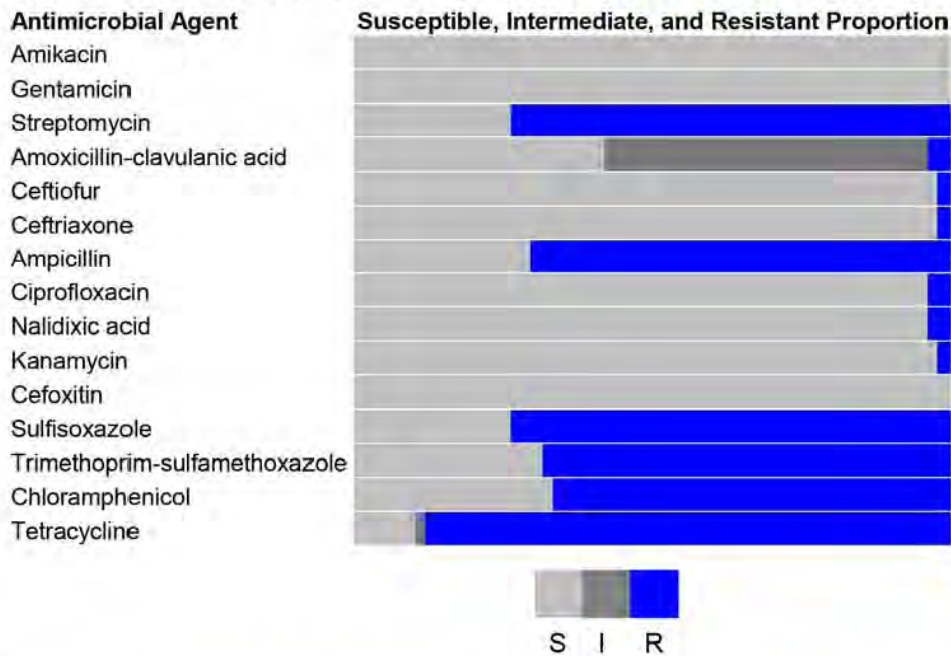
\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.



**Figure 23. Antimicrobial resistance pattern for *Shigella flexneri*, 2009**



**Table 40. Percentage and number of *Shigella flexneri* isolates resistant to antimicrobial agents, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009		
<b>Total Isolates</b>	<b>75</b>	<b>91</b>	<b>73</b>	<b>51</b>	<b>62</b>	<b>52</b>	<b>74</b>	<b>61</b>	<b>45</b>	<b>57</b>		
Rank* CLSI† Antimicrobial Class Antibiotic (Resistance breakpoint)												
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Gentamicin (MIC ≥ 16)	0.0%	0.0%	1.4%	0.0%	0.0%	0.0%	1.4%	0.0%	0.0%	
		Streptomycin (MIC ≥ 64)	61.3%	47.3%	43.8%	60.8%	71.0%	57.7%	58.1%	52.5%	62.2%	73.7%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	4.0%	4.4%	5.5%	2.0%	1.6%	0.0%	0.0%	0.0%	4.4%	3.5%
		Cephems	Ceftiofur (MIC ≥ 8)	0.0%	0.0%	1.4%	2.0%	0.0%	0.0%	1.4%	0.0%	0.0%
		Ceftriaxone (MIC ≥ 64)	0.0%	0.0%	1.4%	2.0%	0.0%	0.0%	1.4%	0.0%	0.0%	1.8%
	Penicillins	Ampicillin (MIC ≥ 32)	77.3%	72.5%	75.3%	84.3%	80.6%	75.0%	63.5%	63.9%	75.6%	70.2%
		Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	1.1%	0.0%	0.0%	0.0%	0.0%	1.4%	1.6%	2.2%
		Nalidixic acid (MIC ≥ 32)	0.0%	3.3%	2.7%	5.9%	1.6%	3.8%	5.4%	4.9%	2.2%	3.5%
	II	Aminoglycosides	Kanamycin (MIC ≥ 64)	0.0%	1.1%	4.1%	3.9%	0.0%	3.8%	0.0%	0.0%	0.0%
Cephems			Cefoxitin (MIC ≥ 32)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Cephalothin (MIC ≥ 32)	2.7%	1.1%	2.7%	3.9%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
Folate pathway inhibitors		Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	53.3%	57.1%	41.1%	52.9%	66.1%	55.8%	68.9%	62.3%	62.2%	73.7%
		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	42.7%	34.1%	28.8%	39.2%	46.8%	44.2%	59.5%	49.2%	48.9%	68.4%
Phenicol		Chloramphenicol (MIC ≥ 32)	69.3%	74.7%	63.0%	68.6%	61.3%	65.4%	54.1%	55.7%	68.9%	66.7%
		Tetracyclines	Tetracycline (MIC ≥ 16)	92.0%	94.5%	78.1%	82.4%	95.2%	94.2%	83.8%	83.6%	86.7%

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.

**Table 41. Resistance patterns of *Shigella flexneri* isolates, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates	75	91	73	51	62	52	74	61	45	57
	%	%	%	%	%	%	%	%	%	%
	n	n	n	n	n	n	n	n	n	n
No resistance detected	4.0%	3.3%	15.1%	7.8%	0.0%	5.8%	5.4%	9.8%	4.4%	5.3%
	3	3	11	4	0	3	4	6	2	3
Resistance ≥ 1 CLSI class*	96.0%	96.7%	84.9%	92.2%	100.0%	94.2%	94.6%	90.2%	95.6%	94.7%
	72	88	62	47	62	49	70	55	43	54
Resistance ≥ 2 CLSI classes*	82.7%	89.0%	76.7%	86.3%	93.5%	80.8%	85.1%	80.3%	93.3%	86.0%
	62	81	56	44	58	42	63	49	42	49
Resistance ≥ 3 CLSI classes*	81.3%	79.1%	75.3%	80.4%	90.3%	78.8%	75.7%	68.9%	84.4%	82.5%
	61	72	55	41	56	41	56	42	38	47
Resistance ≥ 4 CLSI classes*	64.0%	62.6%	57.5%	62.7%	64.5%	65.4%	47.3%	55.7%	57.8%	63.2%
	48	57	42	32	40	34	35	34	26	36
Resistance ≥ 5 CLSI classes*	32.0%	25.3%	19.2%	31.4%	29.0%	30.8%	28.4%	27.9%	28.9%	49.1%
	24	23	14	16	18	16	21	17	13	28
At least ACSSuT <sup>†</sup>	29.3%	22.0%	15.1%	29.4%	27.4%	28.8%	27.0%	26.2%	24.4%	47.4%
	22	20	11	15	17	15	20	16	11	27
At least ACT/S <sup>‡</sup>	32.0%	23.1%	21.9%	27.5%	24.2%	32.7%	28.4%	26.2%	26.7%	47.4%
	24	21	16	14	15	17	21	16	12	27
At least AT/S <sup>§</sup>	38.7%	25.3%	27.4%	37.3%	35.5%	38.5%	43.2%	36.1%	33.3%	52.6%
	29	23	20	19	22	20	32	22	15	30
At least ANT/S <sup>¶</sup>	0.0%	1.1%	1.4%	5.9%	0.0%	1.9%	2.7%	1.6%	0.0%	1.8%
	0	1	1	3	0	1	2	1	0	1
At least ACSSuTAuCx <sup>**</sup>	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	0	0	0	0	0	0	0	0	0	0
At least ceftriaxone and nalidixic acid resistant	0.0%	0.0%	0.0%	2.0%	0.0%	0.0%	1.4%	0.0%	0.0%	0.0%
	0	0	0	1	0	0	1	0	0	0

\* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ AT/S: resistance to ampicillin, trimethoprim-sulfamethoxazole

¶ ANT/S: resistance to AT/S, nalidixic acid

\*\* ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone



**Table 43. Percentage and number of *Escherichia coli* O157 isolates resistant to antimicrobial agents, 2000–2009**

Year			2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates			407	277	399	158	169	194	233	190	159	188
Rank*	CLSI† Antimicrobial Class	Antibiotic (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Gentamicin (MIC ≥ 16)	0.5%	0.4%	0.0%	0.0%	0.6%	0.5%	0.0%	0.0%	1.3%	0.5%
		Streptomycin (MIC ≥ 64)	5.2%	1.8%	2.3%	1.9%	1.8%	2.1%	2.6%	2.1%	1.9%	4.8%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	1.0%	0.7%	0.0%	1.3%	0.0%	0.0%	1.3%	0.5%	0.6%	0.5%
		Cephems										
	Cephems	Ceftiofur (MIC ≥ 8)	1.0%	1.1%	0.0%	1.3%	0.0%	0.0%	1.3%	0.0%	0.6%	0.0%
		Ceftriaxone (MIC ≥ 64)	1.0%	0.7%	0.0%	1.3%	0.0%	0.0%	1.3%	0.0%	0.6%	0.0%
	Penicillins	Ampicillin (MIC ≥ 32)	2.7%	2.2%	1.5%	3.2%	1.2%	4.1%	2.6%	2.1%	3.8%	4.3%
		Quinolones										
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.4%	0.5%	0.0%	0.5%
Nalidixic acid (MIC ≥ 32)		0.5%	1.1%	1.0%	0.6%	1.8%	1.5%	2.1%	2.1%	1.3%	2.1%	
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	1.0%	0.0%	0.5%	0.0%	0.0%	0.5%	0.4%	0.0%	0.0%	0.5%
		Cephems										
	Cephems	Cefoxitin (MIC ≥ 32)	1.0%	0.7%	0.0%	1.3%	0.6%	0.0%	1.3%	0.0%	1.3%	0.5%
		Cephalothin (MIC ≥ 32)	1.2%	1.4%	1.5%	3.2%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
	Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	5.9%	5.1%	3.5%	3.8%	1.8%	6.7%	3.0%	2.6%	3.1%	6.4%
		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	0.7%	0.7%	0.5%	0.6%	0.0%	0.5%	0.4%	1.1%	1.3%	4.3%
	Phenicol	Chloramphenicol (MIC ≥ 32)	3.7%	1.4%	1.3%	1.3%	0.6%	1.0%	1.3%	0.5%	0.6%	1.1%
		Tetracyclines										
	Tetracyclines	Tetracycline (MIC ≥ 16)	7.1%	5.4%	3.0%	5.7%	1.8%	8.8%	4.7%	4.7%	1.9%	7.4%

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

**Table 44. Resistance patterns of *Escherichia coli* O157 isolates, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates	407	277	399	158	169	194	233	190	159	188
	%	%	%	%	%	%	%	%	%	%
	n	n	n	n	n	n	n	n	n	n
No resistance detected	90.4%	91.3%	94.0%	90.5%	94.7%	87.6%	91.8%	92.1%	91.8%	89.9%
	368	253	375	143	160	170	214	175	146	169
Resistance ≥ 1 CLSI class*	9.6%	8.7%	6.0%	9.5%	5.3%	12.4%	8.2%	7.9%	8.2%	10.1%
	39	24	24	15	9	24	19	15	13	19
Resistance ≥ 2 CLSI classes*	6.6%	5.4%	3.8%	5.1%	2.4%	6.7%	4.7%	3.2%	3.1%	7.4%
	27	15	15	8	4	13	11	6	5	14
Resistance ≥ 3 CLSI classes*	4.7%	2.2%	2.0%	3.2%	1.2%	5.2%	3.4%	2.1%	2.5%	5.9%
	19	6	8	5	2	10	8	4	4	11
Resistance ≥ 4 CLSI classes*	3.4%	1.4%	0.8%	1.3%	0.6%	1.0%	2.1%	1.1%	1.3%	4.3%
	14	4	3	2	1	2	5	2	2	8
Resistance ≥ 5 CLSI classes*	1.2%	0.4%	0.0%	0.0%	0.0%	0.0%	0.9%	0.5%	0.0%	0.5%
	5	1	0	0	0	0	2	1	0	1
At least ACSSuT†	1.2%	0.4%	0.0%	0.0%	0.0%	0.0%	0.9%	0.0%	0.0%	0.0%
	5	1	0	0	0	0	2	0	0	0
At least ACT/S‡	0.2%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.6%	0.0%
	1	0	0	0	0	0	0	0	1	0
At least ACSSuTAuCx§	1.0%	0.4%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	4	1	0	0	0	0	0	0	0	0
At least ceftriaxone and nalidixic acid resistant	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.4%	0.0%	0.0%	0.0%
	0	0	0	0	0	0	1	0	0	0

\* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

## 5. Campylobacter

**Table 45. Frequency of *Campylobacter* species isolated in NARMS, 2009**

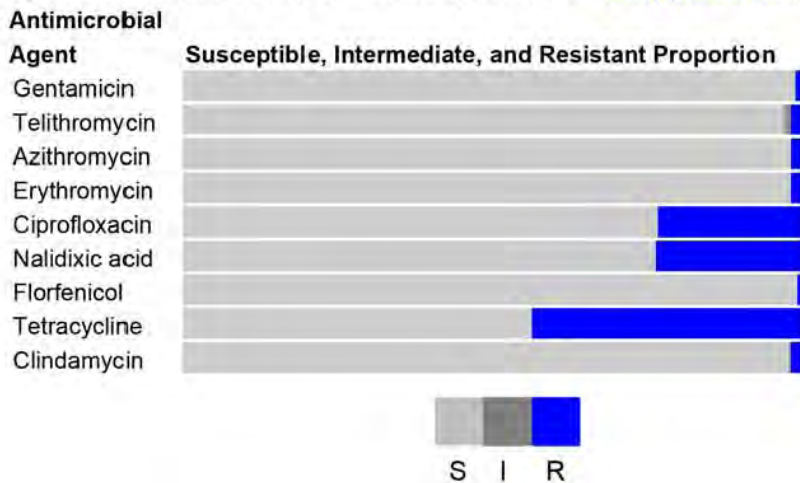
Species	2009	
	N	(%)
<i>Campylobacter jejuni</i>	1355	(90.2%)
<i>Campylobacter coli</i>	143	(9.5%)
Other	4	(0.3%)
<b>Total</b>	<b>1502</b>	<b>(100.0%)</b>

**Table 46. Minimum inhibition concentrations (MICs) and resistance of *Campylobacter* isolates to antimicrobial agents, 2009 (N=1502)**

Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent	% of isolates			Percent of all isolates with MIC (µg/mL)‡														
			%I‡	%R§	[95% CI]¶	0.015	0.03	0.06	0.125	0.25	0.50	1	2	4	8	16	32	64	128	256
I	Aminoglycosides	Gentamicin	0.0	0.9	[0.5 - 1.5]	[Shaded area: 0.015-0.125; Vertical bars at 0.06, 0.125, 0.25, 0.50, 1, 2, 4, 8, 16, 32, 64, 128, 256, 512; Unshaded area: 0.125-0.512]														
		Telithromycin	0.9	1.5	[0.9 - 2.2]	[Shaded area: 0.015-0.125; Vertical bars at 0.06, 0.125, 0.25, 0.50, 1, 2, 4, 8, 16, 32, 64, 128, 256, 512; Unshaded area: 0.125-0.512]														
	Macrolides	Azithromycin	0.0	1.7	[1.1 - 2.4]	[Shaded area: 0.015-0.125; Vertical bars at 0.06, 0.125, 0.25, 0.50, 1, 2, 4, 8, 16, 32, 64, 128, 256, 512; Unshaded area: 0.125-0.512]														
		Erythromycin	0.0	1.7	[1.1 - 2.4]	[Shaded area: 0.015-0.125; Vertical bars at 0.06, 0.125, 0.25, 0.50, 1, 2, 4, 8, 16, 32, 64, 128, 256, 512; Unshaded area: 0.125-0.512]														
	Quinolones	Ciprofloxacin	< 0.1	22.9	[20.8 - 25.1]	[Shaded area: 0.015-0.125; Vertical bars at 0.06, 0.125, 0.25, 0.50, 1, 2, 4, 8, 16, 32, 64, 128, 256, 512; Unshaded area: 0.125-0.512]														
		Nalidixic acid	< 0.1	23.2	[21.1 - 25.4]	[Shaded area: 0.015-0.125; Vertical bars at 0.06, 0.125, 0.25, 0.50, 1, 2, 4, 8, 16, 32, 64, 128, 256, 512; Unshaded area: 0.125-0.512]														
II	Phenicol	Florfenicol††	0.0	0.5	[0.2 - 1.0]	[Shaded area: 0.015-0.125; Vertical bars at 0.06, 0.125, 0.25, 0.50, 1, 2, 4, 8, 16, 32, 64, 128, 256, 512; Unshaded area: 0.125-0.512]														
		Tetracyclines	Tetracycline	0.1	43.5	[40.9 - 46.0]	[Shaded area: 0.015-0.125; Vertical bars at 0.06, 0.125, 0.25, 0.50, 1, 2, 4, 8, 16, 32, 64, 128, 256, 512; Unshaded area: 0.125-0.512]													
III	Lincosamides	Clindamycin	0.3	1.4	[0.9 - 2.1]	[Shaded area: 0.015-0.125; Vertical bars at 0.06, 0.125, 0.25, 0.50, 1, 2, 4, 8, 16, 32, 64, 128, 256, 512; Unshaded area: 0.125-0.512]														

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically important; Rank 2, Highly important; Rank 3, Important  
 † CLSI: Clinical and Laboratory Standards Institute  
 ‡ Percent of isolates with intermediate susceptibility. NA if no MIC range of intermediate susceptibility exists  
 § Percent of isolates that were resistant  
 ¶ The 95% confidence intervals (CI) for percent resistant (%R) were calculated using the Paulson-Camp-Pratt approximation to the Copper-Pearson exact method. The 95% CI is presented to summarize uncertainty in the observed resistance (R%).  
 \*\* The unshaded areas indicate the dilution range of the Sensititre plates used to test isolates. Single vertical bars indicate the breakpoints for susceptibility, while double vertical bars indicate breakpoints for resistance. Numbers in the shaded areas indicate the percentages of isolates with MICs greater than the highest concentrations on the Sensititre plate. Numbers listed for the lowest tested concentrations represent the percentages of isolates with MICs equal to or less than the lowest tested concentration. CLSI breakpoints were used when available.  
 †† Only a susceptible breakpoint ( $\leq 4$  µg/ml) has been established. In this report, isolates with an MIC  $\geq 8$  µg/ml are categorized as resistant.

**Figure 25. Antimicrobial resistance pattern for *Campylobacter*, 2009**





**Table 47. Percentage and number of *Campylobacter* isolates resistant to antimicrobial agents, 2000–2009**

Year			2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates			324	384	354	328	347	890	816	1100	1159	1502
Rank <sup>*</sup>	CLSI <sup>†</sup> Antimicrobial Class	Antibiotic (Resistance breakpoint)										
I	Aminoglycosides	Gentamicin (MIC ≥ 8)	0.3%	0.0%	0.0%	0.3%	0.3%	0.7%	0.1%	0.6%	1.1%	0.9%
			1	0	0	1	1	6	1	7	13	13
	Ketolides	Telithromycin (MIC ≥ 16)	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	1.0%	1.6%	1.5%	2.5%	1.5%
								9	13	16	29	22
	Macrolides	Azithromycin (MIC ≥ 8)	1.9%	2.1%	2.0%	0.9%	0.6%	1.9%	1.7%	2.0%	3.0%	1.7%
			6	8	7	3	2	17	14	22	35	25
Quinolones	Erythromycin (MIC ≥ 32)	1.2%	2.1%	1.4%	0.9%	0.3%	1.8%	1.7%	2.0%	3.0%	1.7%	
		4	8	5	3	1	16	14	22	35	25	
		Ciprofloxacin (MIC ≥ 4)	14.8%	19.5%	20.1%	17.7%	19.0%	21.7%	19.6%	26.0%	23.0%	22.9%
			48	75	71	58	66	193	160	286	267	344
		Nalidixic acid (MIC ≥ 64)	16.7%	20.3%	20.6%	18.9%	19.6%	22.4%	20.1%	26.5%	23.6%	23.2%
			54	78	73	62	68	199	164	291	273	348
II	Phenicol	Chloramphenicol (MIC ≥ 32)	0.0%	0.3%	0.3%	0.0%	1.4%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
			0	1	1	0	5					
		Florfenicol <sup>‡</sup>	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	0.6%	0.0%	0.0%	0.5%	0.5%
		Susceptible breakpoint: (MIC ≤ 4)	Tested	Tested	Tested	Tested	Tested	5	0	0	6	8
	Tetracyclines	Tetracycline (MIC ≥ 16)	38.3%	40.9%	41.2%	38.4%	46.1%	40.6%	46.0%	44.4%	43.7%	43.5%
			124	157	146	126	160	361	375	488	507	653
III	Lincosamides	Clindamycin (MIC ≥ 8)	0.9%	2.1%	2.0%	0.6%	2.0%	1.5%	2.0%	1.7%	2.8%	1.4%
			3	8	7	2	7	13	16	19	32	21

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important; Rank 3, Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Only a susceptible breakpoint (≤ 4 µg/ml) has been established. In this report, isolates with an MIC ≥ 8 µg/ml are categorized as resistant

**Table 48. Resistance patterns of *Campylobacter* isolates, 2000–2009**

Year	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates	324	384	354	328	347	890	816	1100	1159	1502
	%	%	%	%	%	%	%	%	%	%
	n	n	n	n	n	n	n	n	n	n
No resistance detected	52.2%	49.2%	48.3%	50.9%	46.1%	48.4%	43.9%	45.2%	45.8%	46.4%
	169	189	171	167	160	431	358	497	531	697
Resistance ≥ 1 CLSI class*	47.8%	50.8%	51.7%	49.1%	53.9%	51.6%	56.1%	54.8%	54.2%	53.6%
	155	195	183	161	187	459	458	603	628	805
Resistance ≥ 2 CLSI classes*	8.0%	13.3%	12.7%	8.5%	14.1%	13.6%	12.0%	17.5%	15.6%	13.8%
	26	51	45	28	49	121	98	192	181	207
Resistance ≥ 3 CLSI classes*	0.9%	1.6%	1.1%	0.9%	1.2%	1.5%	1.5%	1.7%	2.5%	1.6%
	3	6	4	3	4	13	12	19	29	24
Resistance ≥ 4 CLSI classes*	0.3%	0.3%	0.0%	0.3%	0.3%	0.3%	0.5%	0.9%	1.1%	1.0%
	1	1	0	1	1	3	4	10	13	15
Resistance ≥ 5 CLSI classes*	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.3%	0.3%
	0	0	0	0	0	0	0	0	3	4

\* CLSI: Clinical and Laboratory Standards Institute



**Table 50. Percentage and number of *Campylobacter jejuni* isolates resistant to antimicrobial agents, 2000–2009**

Year	Total Isolates		2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
			306	365	329	303	320	791	709	992	1046	1355
Rank*	CLSI† Antimicrobial Class	Antibiotic (Resistance breakpoint)										
I	Aminoglycosides	Gentamicin (MIC ≥ 8)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.3% 1	0.5% 4	0.0% 0	0.7% 7	1.1% 12	0.7% 9
	Ketolides	Telithromycin (MIC ≥ 16)	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	0.6% 5	0.8% 6	1.0% 10	2.2% 23	1.4% 19
	Macrolides	Azithromycin (MIC ≥ 8)	1.6% 5	1.9% 7	1.8% 6	0.3% 1	0.6% 2	1.8% 14	0.8% 6	1.6% 16	2.3% 24	1.5% 21
		Erythromycin (MIC ≥ 32)	1.0% 3	1.9% 7	1.2% 4	0.3% 1	0.3% 1	1.6% 13	0.8% 6	1.6% 16	2.3% 24	1.5% 21
	Quinolones	Ciprofloxacin (MIC ≥ 4)	14.7% 45	18.4% 67	20.7% 68	17.2% 52	18.1% 58	21.5% 170	19.5% 138	25.8% 256	22.4% 234	23.0% 312
		Nalidixic acid (MIC ≥ 64)	16.0% 49	18.9% 69	21.3% 70	17.8% 54	18.4% 59	21.9% 173	19.0% 135	26.1% 259	22.8% 239	23.2% 314
II	Phenicol	Chloramphenicol (MIC ≥ 8)	0.0% 0	0.3% 1	0.3% 1	0.0% 0	1.6% 5	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
		Florfenicol‡ (Susceptible breakpoint: (MIC ≤ 4))	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	0.5% 4	0.0% 0	0.0% 0	0.6% 6	0.6% 8
	Tetracyclines	Tetracycline (MIC ≥ 16)	39.2% 120	40.3% 147	41.3% 136	38.3% 116	46.9% 150	41.8% 331	47.4% 336	44.8% 444	44.3% 463	43.4% 588
III	Lincosamides	Clindamycin (MIC ≥ 8)	0.7% 2	1.9% 7	1.8% 6	0.0% 0	2.2% 7	1.1% 9	1.0% 7	1.3% 13	2.1% 22	1.3% 18

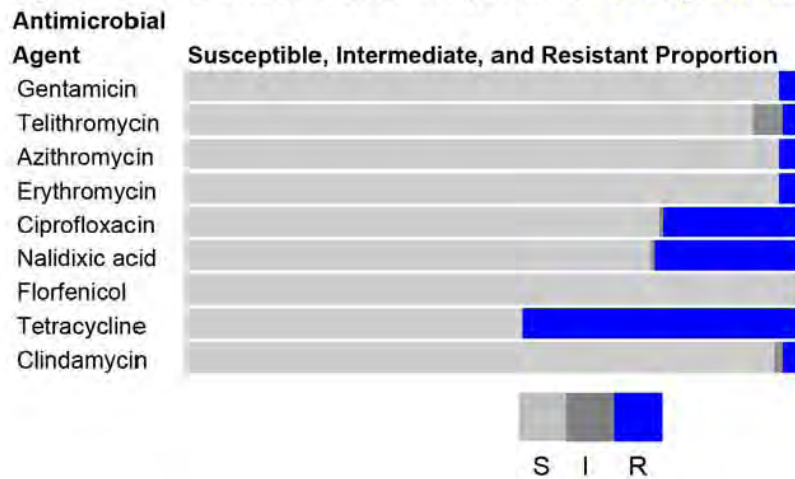
\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important; Rank 3, Important  
 † CLSI: Clinical and Laboratory Standards Institute  
 ‡ Only a susceptible breakpoint (≤ 4 µg/ml) has been established. In this report, isolates with an MIC ≥ 8 µg/ml are categorized as resistant

**Table 51. Minimum inhibitory concentrations (MICs) and resistance of *Campylobacter coli* isolates to antimicrobial agents, 2009 (N=143)**

Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent	% of isolates			Percent of all isolates with MIC (µg/mL)‡																
			%I§	%R¶	[95% CI]¶	0.015	0.03	0.06	0.125	0.25	0.50	1	2	4	8	16	32	64	128	256	512	
I	Aminoglycosides	Gentamicin	0.0	2.8	[0.8 - 7.0]				0.7	5.6	48.3	39.9	2.8								2.8	
	Ketolide	Telithromycin	4.8	2.1	[0.4 - 6.0]			0.7	0.7	11.2	18.9	21.0	21.0	19.8	4.9	2.1						
	Macrolides	Azithromycin	0.0	2.8	[0.8 - 7.0]	0.7	2.1	18.2	47.6	25.2	3.5											2.8
		Erythromycin	0.0	2.8	[0.8 - 7.0]			0.7	0.7	5.6	34.3	29.4	16.8	9.1	0.7							0.7
	Quinolones	Ciprofloxacin	0.7	21.7	[15.2 - 29.3]			15.4	37.8	15.4	8.4	0.7	0.7	3.5	5.6	9.8	1.4	1.4				
Nalidixic acid		0.7	23.1	[16.4 - 30.9]									43.4	28.0	4.9	0.7	7.0	16.1				
II	Phenicol	Florfenicol††	0.0	0.0	[0.0 - 2.5]					7.0	56.6	29.4	7.0									
	Tetracyclines	Tetracycline	0.0	44.8	[36.4 - 53.3]			1.4	14.7	17.5	12.6	8.4	0.7								3.5	41.3
III	Lincosamides	Clindamycin	1.4	2.1	[0.4 - 6.0]			0.7	5.6	16.8	28.7	29.4	11.9	3.5	1.4	1.4	0.7					

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically important; Rank 2, Highly important; Rank 3, Important  
 † CLSI: Clinical and Laboratory Standards Institute  
 ‡ Percent of isolates with intermediate susceptibility, N/A if no MIC range of intermediate susceptibility exists  
 § Percent of isolates that were resistant  
 ¶ The 95% confidence intervals (CI) for percent resistant (%R) were calculated using the Paulson-Camp-Pratt approximation to the Copper-Pearson exact method. The 95% CI is presented to summarize uncertainty in the observed resistance (R%).  
 \*\* The unshaded areas indicate the dilution range of the Sensititre plates used to test isolates. Single vertical bars indicate the breakpoints for susceptibility, while double vertical bars indicate breakpoints for resistance. Numbers in the shaded areas indicate the percentages of isolates with MICs greater than the highest concentrations on the Sensititre plate. Numbers listed for the lowest tested concentrations represent the percentages of isolates with MICs equal to or less than the lowest tested concentration. CLSI breakpoints were used when available.  
 †† Only a susceptible breakpoint (≤ 4 µg/ml) has been established. In this report, isolates with an MIC ≥ 8 µg/ml are categorized as resistant.

**Figure 27. Antimicrobial resistance pattern for *Campylobacter coli*, 2009**



**Table 52. Percentage and number of *Campylobacter coli* isolates resistant to antimicrobial agents, 2000–2009**

Year			2000	2001	2002	2003	2004	2005	2006	2007	2008	2009
Total Isolates			12	17	25	22	26	98	97	105	110	143
Rank*	CLSI† Antimicrobial Class	Antibiotic (Resistance breakpoint)										
I	Aminoglycosides	Gentamicin (MIC ≥ 8)	8.3% 1	0.0% 0	0.0% 0	4.5% 1	0.0% 0	2.0% 2	1.0% 1	0.0% 0	0.9% 1	2.8% 4
	Ketolides	Telithromycin (MIC ≥ 16)	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	4.1% 4	7.2% 7	5.7% 6	5.5% 6	2.1% 3
		Macrolides	Azithromycin (MIC ≥ 8)	8.3% 1	5.9% 1	4.0% 1	9.1% 2	0.0% 0	3.1% 3	8.2% 8	5.7% 6	10.0% 11
			Erythromycin (MIC ≥ 32)	8.3% 1	5.9% 1	4.0% 1	9.1% 2	0.0% 0	3.1% 3	8.2% 8	5.7% 6	10.0% 11
	Quinolones	Ciprofloxacin (MIC ≥ 4)	25.0% 3	47.1% 8	12.0% 3	22.7% 5	30.8% 8	23.5% 23	21.6% 21	28.6% 30	30.0% 33	21.7% 31
			Nalidixic acid (MIC ≥ 64)	25.0% 3	47.1% 8	12.0% 3	22.7% 5	34.6% 9	26.5% 26	23.7% 23	30.5% 32	30.0% 33
II	Phenicol	Chloramphenicol (MIC ≥ 32)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
		Florfenicol†	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	1.0% 1	0.0% 0	0.0% 0	0.0% 0	0.0% 0
		Susceptible breakpoint: (MIC ≤ 4)	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	1	0	0	0	0
	Tetracyclines	Tetracycline (MIC ≥ 16)	25.0% 3	58.8% 10	40.0% 10	45.5% 10	38.5% 10	30.6% 30	39.2% 38	41.9% 44	40.0% 44	44.8% 64
III	Lincosamides	Clindamycin (MIC ≥ 8)	8.3% 1	5.9% 1	4.0% 1	9.1% 2	0.0% 0	4.1% 4	9.3% 9	5.7% 6	9.1% 10	2.1% 3

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important; Rank 3, Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Only a susceptible breakpoint (≤ 4 µg/ml) has been established. In this report, isolates with an MIC ≥ 8 µg/ml are categorized as resistant

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## Appendix A

### Summary of *Escherichia coli* Resistance Surveillance Pilot Study, 2009

#### ***Escherichia coli* working group**

#### **Centers for Disease Control and Prevention**

Frederick Angulo, Ezra Barzilay, Patricia Griffin, Amy Krueger, Rebecca Howie, Kathryn Lupoli, Andre McCullough, Kevin Joyce, Felicita Medalla  
Foodborne and Diarrheal Diseases Branch, Division of Bacterial and Mycotic Diseases  
National Center for Infectious Diseases

#### ***Michigan Department of Community Health and William Beaumont Hospital***

Sue Donabedian, Mary Beth Perry, Mary Thill, Mark Zervos

## INTRODUCTION

*Escherichia coli* is a Gram–negative coccobacillus bacterium that is part of the intestinal flora of humans and other animals. Because antimicrobial resistance genes commonly reside in mobile genetic elements that can be transferred horizontally to other bacteria, antimicrobial–resistant bacteria of the intestinal flora, including *E. coli*, constitute an important reservoir of resistance genes for pathogenic bacteria of humans and other animals. Furthermore, when introduced into a normally sterile site, *E. coli* is an important cause of infections, including septicemia, urinary tract infections, and wound infections. The human intestinal tract is the predominant source of *E. coli* causing these infections. Antimicrobial resistance among *E. coli* causing such infections complicates treatment options.

The use of antimicrobial agents creates a selective pressure for the emergence and dissemination of resistant bacteria. Use of antimicrobial agents in food animals selects resistant bacteria, including resistant *E. coli* in the intestinal tract of food animals. These resistant bacteria can be transmitted to humans through the food supply. Therefore, monitoring resistance in *E. coli* isolated from the intestinal flora of humans and animals is important to determining the role of these bacteria as human pathogens and as reservoirs of resistance determinants for human pathogens. The *E. coli* Resistance Surveillance Pilot is designed to determine the prevalence of resistance to clinically important antimicrobial agents among *E. coli* isolated from persons in the community.

## SUMMARY OF 2009 SURVEILLANCE DATA

### Background

Beginning in 2004, NARMS began to prospectively monitor the prevalence of antimicrobial resistance of *E. coli* isolated from human stool samples in two sites: Maryland and Michigan.

### SURVEILLANCE AND LABORATORY TESTING METHODS

In 2009, Michigan was the sole participant in the study. Michigan cultured 10 human stool samples, from outpatients, each month for *E. coli* using Eosin Methylene Blue agar and subsequent biochemical confirmation. One *E. coli* isolate, if present, from each stool sample was sent to CDC for susceptibility testing to antimicrobial agents using broth microdilution (Sensititre®) to determine the minimum inhibitory concentration (MIC) for each of

15 antimicrobial agents: amikacin, ampicillin, amoxicillin-clavulanic acid, ceftiofur, ceftriaxone, chloramphenicol, ciprofloxacin, gentamicin, kanamycin, nalidixic acid, streptomycin, sulfonamides, tetracycline, and trimethoprim-sulfamethoxazole.

Interpretive criteria from the Clinical and Laboratory Standards Institute (CLSI) were used when available ([Table 53](#)). The 95% CIs for the percentage of resistant isolates calculated using the Clopper-Pearson exact method, are included in the MIC distribution tables. Similarly, multiclass resistance by CLSI antimicrobial class was defined as resistance to two or more classes.

## RESULTS

In 2009, CDC received 45 isolates; of these, 45 (100.0%) were viable *E. coli* isolates. MIC was determined for *E. coli* isolates for 15 antimicrobial agents ([Table 54](#)). Of the 45 *E. coli* isolates, 22.2% (10/45) were resistant to ampicillin, 17.8% (8/45) to sulfisoxazole, 17.8% (8/45) to tetracycline, and 8.9% (4/45) to nalidixic acid ([Table 55](#)).

### Multidrug-Resistant *E. coli*

Multidrug resistance is described in NARMS by the number of antimicrobial classes and also by specific coresistant phenotypes. Antimicrobial classes of agents defined by CLSI are used in this report.

- 11.1% (5/45) of *E. coli* isolates were resistant to three or more classes of antimicrobial agents ([Table 56](#)).
- 8.9% (4/45) of *E. coli* isolates were resistant to five or more classes of antimicrobial agents ([Table 56](#)).

### Clinically Important Resistance

Antimicrobial agents commonly used to treat serious *E. coli* infections in humans include third-generation cephalosporins and fluoroquinolones.

- 2.2% (1/45) of *E. coli* isolates were resistant to ceftriaxone ([Table 55](#)).
- 8.9% (4/45) of *E. coli* isolates were resistant to ciprofloxacin ([Table 55](#)).

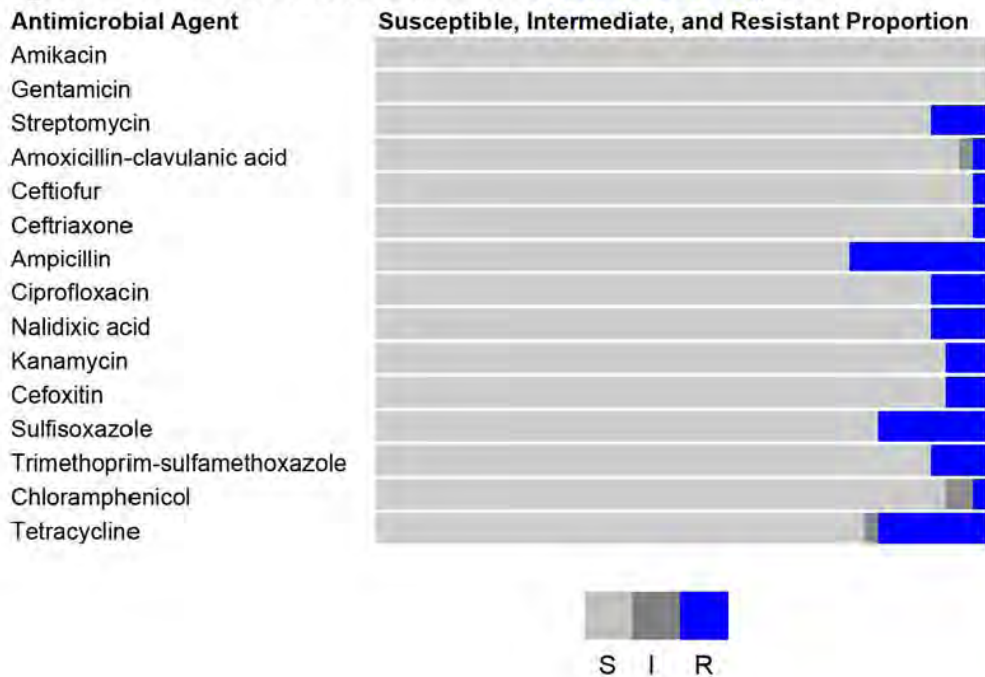
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**Figure 28. Antibiotic resistance pattern for *Escherichia coli*, 2009**



**Table 55. Percentage and number of *Escherichia coli* isolates resistant to antimicrobial agents, 2004–2009**

Year			2004	2005	2006	2007	2008	2009
Total Isolates			151	119	82	66	57	45
Rank*	CLSI† Antimicrobial Class	Antibiotic (Resistance breakpoint)						
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Gentamicin (MIC ≥ 16)	2.0%	3.4%	3.7%	3.0%	0.0%	0.0%
		Streptomycin (MIC ≥ 64)	10.6%	14.3%	7.3%	13.6%	8.8%	8.9%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32)	2.6%	4.2%	3.7%	0.0%	3.5%	2.2%
		Cephems						
	Cephems	Ceftiofur (MIC ≥ 8)	0.0%	0.8%	0.0%	0.0%	1.8%	2.2%
		Ceftriaxone (MIC ≥ 64)	0.0%	0.8%	0.0%	0.0%	1.8%	2.2%
	Penicillins	Ampicillin (MIC ≥ 32)	24.5%	26.1%	28.0%	21.2%	26.3%	22.2%
	Quinolones	Ciprofloxacin (MIC ≥ 4)	3.3%	7.6%	4.9%	7.6%	10.5%	8.9%
		Nalidixic Acid (MIC ≥ 32)	9.3%	9.2%	11.0%	10.6%	12.3%	8.9%
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	2.0%	0.0%	0.0%	1.5%	1.8%	6.7%
		Cephems						
	Folate pathway inhibitors	Sulfisoxazole‡ (MIC ≥ 512)	17.9%	18.4%	17.1%	24.2%	14.0%	17.8%
		Trimethoprim-sulfamethoxazole (MIC ≥ 4)	11.3%	14.9%	12.2%	15.2%	12.3%	8.9%
	Phenicols	Chloramphenicol (MIC ≥ 32)	1.3%	2.5%	3.7%	3.0%	5.3%	2.2%
	Tetracyclines	Tetracycline (MIC ≥ 16)	13.2%	19.3%	14.6%	21.2%	14.0%	17.8%

\* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important  
 † CLSI: Clinical and Laboratory Standards Institute  
 ‡ Results unavailable for 5 isolates

**Table 56. Resistance patterns of *Escherichia coli* isolates, 2004–2009**

Year	2004	2005	2006	2007	2008	2009
<b>Total Isolates</b>	<b>151</b>	<b>119</b>	<b>82</b>	<b>66</b>	<b>57</b>	<b>45</b>
	%	%	%	%	%	%
	n	n	n	n	n	n
No resistance detected	62.9%	63.0%	63.4%	65.2%	64.9%	68.9%
	95	75	52	43	37	31
Resistance ≥1CLSI class*	37.7%	37.0%	36.6%	34.8%	35.1%	31.1%
	57	44	30	23	20	14
Resistance ≥2 CLSI classes*	17.9%	22.7%	22.0%	21.2%	21.1%	13.3%
	27	27	18	14	12	6
Resistance ≥3 CLSI classes*	9.9%	14.3%	15.9%	15.2%	12.3%	11.1%
	15	17	13	10	7	5
Resistance ≥4 CLSI classes*	5.3%	9.2%	8.5%	9.1%	8.8%	8.9%
	8	11	7	6	5	4
Resistance ≥5 CLSI classes*	3.3%	7.6%	1.2%	4.5%	7.0%	8.9%
	5	9	1	3	4	4
At least ACSSuT <sup>†</sup>	1.3%	0.8%	0.0%	0.0%	1.8%	0.0%
	2	1	0	0	1	0
At least ACT/S <sup>‡</sup>	1.3%	0.8%	1.2%	1.5%	3.5%	0.0%
	2	1	1	1	2	0
At least ACSSuTAuCx <sup>§</sup>	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	0	0	0	0	0	0
At least ceftriaxone and nalidixic acid resistant	0.0%	0.0%	0.0%	0.0%	1.8%	2.2%
	0	0	0	0	1	1

\* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone