# Newborn Screening Quality Assurance Program

# anti-HIV-1 Antibodies in Dried Blood Spots Proficiency Testing Program (HIVPT)

# November 4, 2017

## Introduction

This report is the summary of data reported within the specified period for Quarter 4, 2017, anti-HIV-1 Anti-bodies in dried blood spots (DBS) PT event. It is distributed to all participants, state laboratory directors, and program colleagues by request. The tables within this report provide certification profiles and distribution information for the HIVPT specimen panel, participant result information for screening methods, confirmatory methods, and final interpretations. An evaluation of your reported data is attached to this summary.

## Certification of PT Specimens

Method and laboratory performance are evaluated by challenging participants with DBS specimens representing HIV- negative and positive serostatuses. Anti-HIV-1 Antibody screening and confirmatory tests should identify all HIV-positive specimens, regardless of subtype. Expected screening and confirmatory results, and final clinical assessments, are provided in Table 1.

Table 1. Expected Results -EIA (OD), Western Blot (Band Detection) and Final Interpretation

Specimen	EIA—Avioq		Western Blot—Genetic Systems HIV-1 WB (Bio-Rad)								Final	
	OD	gp160	gp120	p65	P55/51	gp41	p40	p31	p24	p18	Interpretation	
41741	0.118	N	N	N	N	N	N	N	N	N	N	
41742	0.095	N	N	N	N	N	N	N	N	N	N	
41743	0.092	N	N	N	N	WP	N	N	N	N	N	
41744	1.726	Р	WP	WP	Р	WP	Р	WP	Р	WP	R	
41745	0.099	N	N	N	N	Ν	N	N	N	N	N	

# **Distribution of PT Specimens**

On October 2, 2017 a PT panel of five individual DBS specimens was distributed to 13 domestic laboratories and 15 international laboratories.

# **Participant Results**

## Screening Data

We received data reports from 23 of the 28 participating laboratories by the designated data reporting deadline. Each participant was asked to analyze the specimens for anti-HIV-1 Antibodies with the assay schemes they routinely use. Data submission must include the screening results, any confirmatory results performed based on presumptive positive screening results, and the analytic methods used for all testing.

Table 2 shows the number of laboratories using enzyme immunoassay (EIA) screening methods/kits both for the primary and secondary methods. Table 3 provides the overall statistics for the screening EIA methods where N>3.

Table 2. Number of EIA Screening Methods Reported; Includes Primary and Secondary Methods

Method Code	Kit Source	Primary *	Secondary
11	In House	1	
27	Tecnosuma (Cuba) UMELISA HIV 1+2	2	
40	Avioq HIV-1 Microeleisa Systems	9	2
41	Bio-Rad HIV-1/HIV-2 plus O EIA	1	
43	Murex® HIV-1.2.O. Diasorin	2	1
12	Other	4	3
	Total	19	6

<sup>\*</sup>Three laboratories did not report EIA data and reported final interpretations based on a Western Blot confirmatory test. One lab reported EIA results but no method.

Table 3. Over Statistics Screening Methods (N>3)

	0, 1, 1,	Specimen							
Method	Statistic	41741	41742	41743	41744	41745			
Avioq HIV-1 Microelisa System (N=9)	Mean	0.107	0.105	0.099	2.042	0.104			
	SD	0.027	0.013	0.016	0.777	0.023			
Gystein (N-9)	%CV	25.120	12.715	16.003	38.060	22.123			

# Confirmatory Data

Fifteen laboratories reported using Western Blot (WB) antibody analysis as either a screening or confirmatory method in the detection of anti-HIV-1 antibodies. Table 4 shows the number of laboratories using each WB kit source. Table 5 shows the Reported Frequency of Bands by WB for each of the PT specimens that tested positive by a primary screening method.

Table 4. Western Blot Confirmatory Methods Reported

Method Code	Kit Source	Total Participants		
16	Genetic Systems HIV-1 WB Kit (Bio-Rad)	10		
32	Cambridge Biotech HIV-1 WB Kit (Maxim)	3		
36	New LAV Blot I (Bio-Rad)	1		
37	Genelab Diagnostics HIV Blot Kit	1		
	Total	15		

Table 5. Frequency of Western Blot Bands for Reactive Specimens (All Methods)

Total	Number of Laboratories Finding Reactive Bands									
Specimen	pecimen Interpretation		gp120	p66	p55	p51	gp41	p31	p24	p18
Specimen 41744	Positive	15	8	7	13	6	8	10	15	2
	Weak Positive	0	4	5	1	6	1	3	0	7
	Negative	0	2	3	1	2	6	2	0	4
	Indeterminate	0	1	0	0	1	0	0	0	2

# Final Interpretations

A final interpretation for each specimen must be submitted to receive an evaluation. Table 6 provides the frequency of all participant interpretations.

Table 6. Frequency Distribution of Final Interpretations (23 Laboratories)

Specimen Number	Expected Value	Non-reactive	Reactive
41741	N	22	1
41742	N	22	1
41743	N	22	1
41744	R	0	23
41745	N	22	1

## **Evaluations**

Overall, participants reported four False-positive and no False-negative results.

# **Future Shipments**

The Newborn Screening Quality Assurance Program will ship next quarter's PT specimens for HIVPT in January 2018.

The content of this report may also be located on our website at: http://www.cdc.gov/labstandards/nsqap\_reports.html

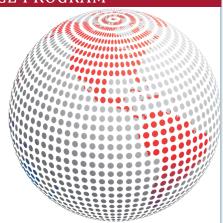
This program is co-sponsored by the Centers for Disease Control and Prevention (CDC) and The Association of Public Health Laboratories (APHL)

# NEWBORN SCREENING QUALITY ASSURANCE PROGRAM

Direct inquiries to:

Centers for Disease Control and Prevention 4770 Buford Highway NE, MS/F19 Atlanta, GA 30341-3724 Phone: 404-488-7945 Email: jvm0@cdc.gov

> <u>Editors</u> Joanne Mei Irene Williams



This NEWBORN SCREENING QUALITY ASSURANCE PROGRAM report is an internal publication distributed to program participants and selected program colleagues. The laboratory quality assurance program is a project cosponsored by the Centers for Disease Control and Prevention (CDC) and the Association of Public Health Laboratories.

## CENTERS FOR DISEASE CONTROL AND PREVENTION (CDC) ATLANTA, GA 30341

## **Acting Director**

Brenda Fitzgerald, M.D.

#### Director

National Center for Environmental Health

Patrick Breysse, Ph.D.

#### Director

Division of Laboratory Sciences James L. Pirkle, M.D., Ph.D.

#### Chief

Newborn Screening and Molecular Biology Branch Carla Cuthbert, Ph.D.

#### Contributors:

Carter Asef
John Bernstein
Quan Bui
Paul Dantonio

Daniel Mandel, Ph.D.
Joanne Mei, Ph.D.
Kristina Mercer
Gyliann Peña

Sharon Flores

Florab ath M. Hall

Konstantinos Petritis, Ph.D.

Sean Scott

Elizabeth M. Hall
Christopher Haynes, Ph.D.
Brandon Kenwood
Francis Lee, Ph.D.
Lixia Li, Ph.D.

Sean Scott
Robert Vogt, Ph.D.
Irene Williams
Sophia Winchester
Golriz Yazdanpanah

Timothy Lim, Ph.D. Sherri Zobel

Production:

Sarah Brown Kizzy Stewart

Kimberly Coulter

#### ASSOCIATION OF PUBLIC HEALTH LABORATORIES SILVER SPRING, MD 20910

## President

Ewa King, PhD

## Chairman, Newborn Screening and Genetics in Public Health Committee

Michele Caggana, Sc.D., FACMG

## Chairman, Newborn Screening Quality Assurance Quality Control Subcommittee

Patricia R. Hunt, B.A. and Joseph Orsini, Ph.D.

## Chairman, Newborn Screening Molecular Subcommittee

Rachel Lee, Ph.D.

#### **INQUIRIES TO:**

Irene Williams, Editor • Centers for Disease Control and Prevention (CDC) • Newborn Screening Quality Assurance Program Mailstop F-24 • 4770 Buford Highway, N.E. • Atlanta, GA 30341-3724

Phone (770) 488-4582 • NSQAPDMT@cdc.gov

E-mail: IWilliams1@cdc.gov