

Effectiveness of Naturally Acquired and Vaccine-Induced Immune Responses to SARS-CoV-2 Mu Variant

Edmilson F. de Oliveira-Filho,¹ Bladimiro Rincon-Orozco,¹ Natalia Jones-Cifuentes, Brigitte Peña-López, Barbara Mühlemann, Christian Drosten, Andres Moreira-Soto, Jan Felix Drexler

SARS-CoV-2 Mu variant emerged in Colombia in 2021 and spread globally. In 49 serum samples from vaccinees and COVID-19 survivors in Colombia, neutralization was significantly lower ($p < 0.0001$) for Mu than a parental strain and variants of concern. Only the Omicron variant of concern demonstrated higher immune evasion.

Diverse SARS-CoV-2 variants have arisen during the pandemic. As of May 4, 2022, there had been 2 recognized variants of concern (VOC), Delta and Omicron, in addition to earlier emerging VOCs Alpha, Beta, and Gamma and strains previously categorized as variants of interest (VOI). Many VOIs have been understudied in terms of pathogenesis, transmissibility, and potential for immune escape. Delta and Omicron illustrate how variants emerging in tropical settings can spread globally.

Mu was first reported as a VOI in early January 2021 in northern Colombia. While outcompeting other locally circulating variants, Mu spread to additional countries, such as Ecuador, United States, Mexico, and Spain; as of early 2022, it was still circulating at low levels in Colombia (1). Mu caused 70% of all COVID-19 cases in Colombia during May–July 2021 (Figure 1), a period which also accounted for the highest number of deaths in Colombia during the pandemic, suggesting substantial

pathogenicity of Mu (1). Mu was later outcompeted by Delta and Omicron, and the number of Mu-related cases gradually decreased through the end of 2021 (Figure 1).

Recent studies relying on data from spike-based pseudovirus testing suggested substantially lower neutralization of Mu compared with the parental B.1 virus in antiserum samples from persons in Japan and China who had received either the BNT162b2 (Pfizer-BioNTech, <https://www.pfizer.com>) or Sinovac (<http://www.sinovac.com>) vaccines or recovered from COVID-19 (2,3). Because of inherent limitations in pseudovirus-based systems for reproducing response variations based on natural infection (4), regional differences of immune responses (5), and different vaccines used in Colombia, we comparatively characterized the neutralization of Mu and VOCs using fully infectious viruses and serum samples from persons in Colombia. The study was approved by the Ethics Committee of the Universidad Industrial de Santander (protocol 4110) and by the Ethics Committee of the Charité-Universitätsmedizin Berlin (protocol EA2/031/22). All participants provided written informed consent.

The Study

By March 2022, ≈68% of the population of Colombia had been vaccinated, predominantly with spike-based mRNA (BNT162b2), vectored (AZD1222; AstraZeneca, <https://www.astrazeneca.com>), and chemically inactivated whole virus-based vaccines (CoronaVac) (Appendix Figure 1, <https://wwwnc.cdc.gov/EID/article/28/8/22-0584-App1.pdf>). To investigate the potency of natural and vaccine-derived immunity, we tested and compared the

Author affiliations: Charité-Universitätsmedizin Berlin, corporate member of Freie Universität Berlin and Humboldt-Universität zu Berlin, Institute of Virology, Berlin, Germany (E.F. de Oliveira-Filho, B. Mühlemann, C. Drosten, A. Moreira-Soto, J.F. Drexler); Universidad Industrial de Santander School of Medicine, Bucaramanga, Colombia (B. Rincon-Orozco, N. Jones-Cifuentes, B. Peña-López); German Centre for Infection Research, Berlin (B. Mühlemann, C. Drosten, J.F. Drexler)

DOI: <https://doi.org/10.3201/eid2808.220584>

¹These first authors contributed equally to this article.

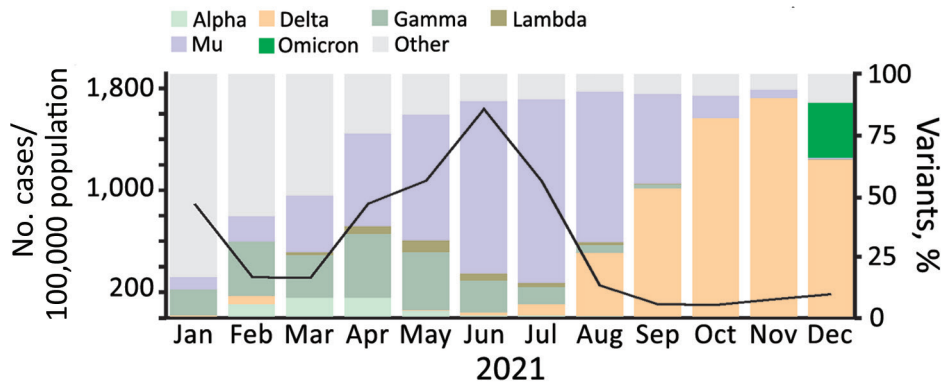


Figure 1. Incidence of SARS-CoV-2 and circulation of variants, by month, Colombia, 2021. Data on variant circulation was obtained from GISAID (<https://www.gisaid.org>) and data on the number of cases in Colombia from the Our World in Data database (<https://www.ourworldindata.org>).

neutralization activity in 49 serum samples from vaccinated and naturally infected persons in Colombia. Among vaccinated persons, we tested serum from 32 persons sampled in October 2021. Of those, 10 vaccinated with BioNTech-Pfizer were tested a median 99.5 d (range 65–170) after completing vaccination, 7 vaccinated with AstraZeneca were tested a median 146.0 d (range 129–173) after completing vaccination, and 15 vaccinated with Sinovac were tested a median 46.0 d (range 28–131) after completing vaccination. We tested serum samples from 17 persons who tested positive for SARS-CoV-2 antibodies (MAGLUMI 2019-nCoV IgG; Snibe Diagnostic, <https://www.snibe.com>) (Table 1; Appendix Table 1) during a seroprevalence study conducted in November 2020. To control whether persons vaccinated with spike-based vaccines were not previously infected, serum samples were tested against the SARS-CoV-2 IgG nucleocapsid protein by ELISA (SARS-CoV-2 NCP kit; Euroimmun, <https://www.euroimmun.com>) (Table 2). We used 50% plaque reduction neutralization tests to obtain neutralizing titers against an early isolate and the Alpha, Beta, Delta, Gamma, Omicron BA.1, and Mu variants (Appendix).

Neutralizing antibody titers against Mu were significantly lower than those against the parental isolate ($p < 0.0001$ by Wilcoxon matched-pairs signed-rank test) in all serum samples tested in this study, irrespective of whether immune responses were elicited by vaccination or by natural infection. Vaccine-derived antibodies neutralized Mu on average 8.1-fold ($p < 0.0001$ by Wilcoxon test) less than the parental strain resembling the vaccine backbones (Figure 2, panels A–C; Appendix Figure 2). We found a similar 8.0-fold reduced neutralization of Mu ($p < 0.0001$ by Wilcoxon test) for the group of naturally infected persons (Figure 2, panel D). Despite the relatively lower neutralization potency observed in serum samples from persons immunized with the inactivated full

virus-based vaccine Sinovac, observed differences in the ability to neutralize Mu compared with the parental strain among the 3 vaccine groups were not statistically significant (range 7.7–11.4-fold; $p = 0.8298$ by Kruskal-Wallis test) (Figure 2).

Compared with other variants, neutralizing antibody titers from serum samples of both naturally infected persons and vaccinees were lower against Mu than against all VOCs except for Omicron (Figure 2, panels A and B). Therefore, our results provide strong evidence for immune evasion of the Mu VOI on the basis of results from robust neutralization testing using full viral isolates. Neutralization of Mu by vaccine-induced antibodies was significantly lower than for Beta ($p = 0.0083$ by Wilcoxon test), for which immune evasion properties led to the suspension of AstraZeneca usage in South Africa (6), and Gamma, which resulted in breakthrough infections in Latin America (7). Immune evasion of Mu is consistent with shared mutations in spike protein residues associated with immune evasion in Beta and Gamma, such as E484K (8). In addition, the mutation leading to the amino acid exchange R346K in Mu is known to be involved in the evasion of monoclonal antibody-mediated neutralization (9), and genomic exchanges occurring at 3 adjacent sites (Y144T, Y145S, and insertion of the amino acid asparagine [N] between spike residues 145 and 146) have been associated with the immune escape properties of Mu (10,11).

Table 1. Median age and days after the second dose of vaccinated persons, by vaccine type, at time of sampling among persons in Colombia*

Vaccine groups	Days after second dose (range)	Age, y (range)
AstraZeneca	146 (129–173)	66.0 (61–72)
Pfizer-BioNTech	99.5 (65–170)	44.6 (27–65)
Sinovac	46.0 (23–131)	44.5 (23–92)

*AstraZeneca (AZD1222), <https://www.astrazeneca.com>; Pfizer-BioNTech (BNT162b2), <https://www.pfizer.com>; Sinovac (CoronaVac), <http://www.sinovac.com>.

Antigenic cartography was recently employed to map the antigenic relationship between the SARS-CoV-2 Omicron and Delta VOCs and other previously circulating VOCs and VOIs (S.H. Wilks et al., unpub. data, <https://www.biorxiv.org/content/10.1101/2022.01.28.477987v1>). Among the serum samples from Colombia vaccinees, there was a high antigenic

distance between Mu and most variants from other serum samples, which clustered together with the parental strain and Alpha (Appendix Figure 3). Of note, antibody responses in naturally infected persons supported past infection with strains bearing similarities to early SARS-CoV-2 isolates and the Gamma variant (Figure 2, panel D). Antibody reactivity in naturally

Table 2. ELISA results and endpoint titers for vaccinee and naturally infected individual serum samples from persons in Colombia*

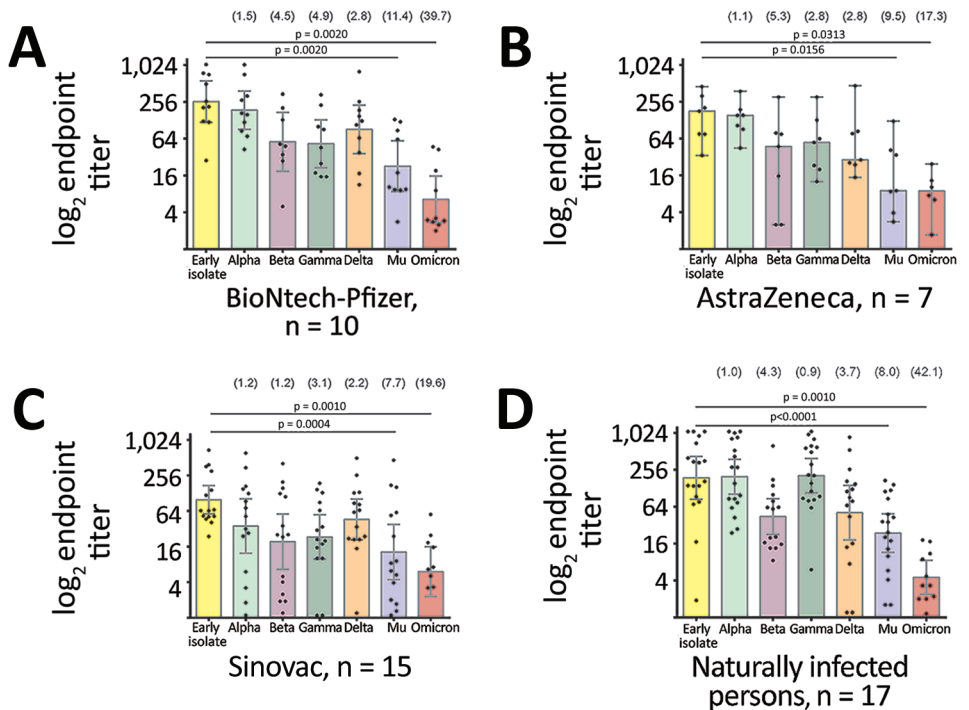
Group	Patient ID	Nucleocapsid		Neutralizing titer by PRNT ₅₀					
		IgG ELISA†	WT	Mu	Alpha	Beta	Gamma	Delta	Omicron
AstraZeneca	AZ2	0.15	204	41	154	79	64	77	13
	AZ3	0.07	453	123	381	305	306	470	25
	AZ4	0.11	75	3	91	16	20	24	6
	AZ5	0.12	76	9	104	3	13	29	2
	AZ6	0.13	34	4	45	3	23	15	0
	AZ9	0.08	179	35	189	75	128	84	10
	AZ10	0.07	319	9	153	47	55	26	8
Pfizer-BioNTech	PF1	0.14	119	9	85	1	18	38	3
	PF2	0.04	28	3	43	35	15	17	3
	PF3	0.15	262	62	158	130	101	149	19
	PF4	0.06	754	121	715	204	226	187	43
	PF5	0.05	501	87	320	48	91	259	9
	PF6	0.07	123	10	119	52	15	11	3
	PF7	0.19	214	9	70	5	0	125	3
	PF8	0.05	207	18	167	28	25	66	3
	PF9	0.09	715	10	273	0	46	108	2
	PF10	0.62	1043	132	1036	343	333	799	47
Sinovac	SVN1	2.96	51	54	72	36	66	83	0
	SVN2	1.68	47	9	24	23	21	22	0
	SVN3	0.46	41	6	1	1	18	1	0
	SVN4	2.43	118	61	151	111	89	87	25
	SVN7	0.97	363	162	347	407	188	259	56
	SVN8	0.81	303	5	93	26	30	61	5
	SVN9	0.69	53	0	32	3	15	35	0
	SVN10	1.61	65	4	27	0	10	66	1
	SVN12	0.29	52	8	52	1	20	21	0
	SVN13	0.39	387	24	126	126	35	130	7
	SVN15	2.81	145	175	168	197	147	133	19
	SVN16	0.40	67	2	6	25	10	21	3
	SVN17	0.07	24	1	1	5	0	15	3
SVN18	0.37	65	0	3	4	0	24	7	
SVN20	1.88	686	464	612	155	131	503	16	
Naturally infected	EA210	ND	696	146	825	595	167	177	2
	EA234	ND	142	4	86	83	67	9	0
	EA238	ND	1,080	48	1080	314	541	79	5
	EA245	ND	70	2	61	154	44	0	0
	EA332	ND	93	10	43	94	1	20	0
	EA334	ND	140	6	74	115	7	16	3
	EA340	ND	77	2	24	61	0	14	2
	EA352	ND	1,080	113	1,080	578	870	59	3
	EA354	ND	336	119	423	972	90	628	0
	EA380	ND	918	43	281	630	151	63	17
	EA396	ND	139	24	98	88	14	21	0
	EA413	ND	1,080	18	864	1,080	260	17	11
	EA422	ND	2	20	28	6	123	100	0
	EA439	ND	398	171	283	812	79	62	9
	EA485	ND	357	87	531	206	114	14	0
	EA501	ND	17	13	86	80	1	0	2
	EA520	ND	166	36	154	211	16	141	1

*AstraZeneca (AZD1222), <https://www.astrazeneca.com>; Pfizer-BioNTech (BNT162b2), <https://www.pfizer.com>; Sinovac (CoronaVac), <http://www.sinovac.com>. ND, not determined; PRNT₅₀, 50% plaque reduction neutralization test; WT, wild-type.

†Cut-off ≥ 0.8 was considered positive.

Figure 2. Comparative neutralization of the Mu SARS-CoV-2 variant in Colombia.

A–C) Neutralization of SARS-CoV-2 variants from serum samples from persons fully immunized with BNT162b2 (Pfizer-BioNTech, <https://www.pfizer.com>) (A), AZD1222 (AstraZeneca, <https://www.astrazeneca.com>) (B), or CoronaVac (Sinovac, <http://www.sinovac.com>) (C). D) Neutralization of SARS-CoV-2 variants by serum samples from naturally infected persons who tested positive for SARS-CoV-2 antibodies during a seroprevalence study in November 2020. For all panels, each point represents the reciprocal plaque reduction neutralization test endpoint titer of 1 tested serum sample for different SARS-CoV-2 variants; colored bars indicate geometric mean titers, and error bars represent 95% CIs. Values in parentheses above bars represent reduction compared to the parental strain. Statistical significance was determined by the Wilcoxon matched signed-rank test; p values are indicated. For clarity of presentation, only significant values between the early isolate and the Mu variant are shown.



infected persons was thus in concordance with the circulation of SARS-CoV-2 variants in South America during the time of sampling in late 2020 (12), supporting the robustness of our data.

Our study was limited by different time points for sampling of vaccinees and the lack of information on natural infections altering immune responses in vaccinees. However, lack of detectable N-protein antibody responses and the absence of clinical records suggestive of COVID-19 infection in vaccinees immunized with spike-based vaccines supports the robustness of our data despite the vaccinees' unclear infection histories.

Conclusions

Our data highlight the importance of continuous monitoring for the emergence of new SARS-CoV-2 variants and strains and the timely identification of those variants with potential to evade naturally elicited and vaccine-derived immune responses, using local sampling specimens in the context of regional epidemiologic conditions. Moreover, our data confirmed the potential of Mu to partially evade immune responses, which may affect the efficacy of vaccination programs in southern America and other areas (7,13). Further studies are warranted to evaluate the

pathogenicity of and cell-mediated immunity against Mu and the ability of immune responses associated with Mu to neutralize other SARS-CoV-2 variants. However, because vaccination boosters still provide some degree of protection against severe disease from Omicron (3,14), which shows more immunity evasion than Mu, vaccination will likely still provide protection against severe disease from Mu.

Acknowledgments

We thank Victor Carvalho Urbieto, Ana María Arboleda, Karina Freyle, and Arne Kühne for their technical support. The Gamma and the Omicron SARS-CoV-2 isolates were obtained from the European Virus Archive) and provided by Dr. Chantal Reusken from the National Institute for Public Health and the Environment.

This work was supported by the Deutsche Gesellschaft für Internationale Zusammenarbeit GmbH (project nos. 88114108 and 81263203), Universidad Industrial de Santander, and MinCiencias-SGR (project no. BPIN 2020000100126).

About the Author

Dr. Oliveira-Filho is a virologist at the Institute of Virology, Charité Universitätsmedizin Berlin. His research interests include the epidemiology and evolution of emerging viruses.

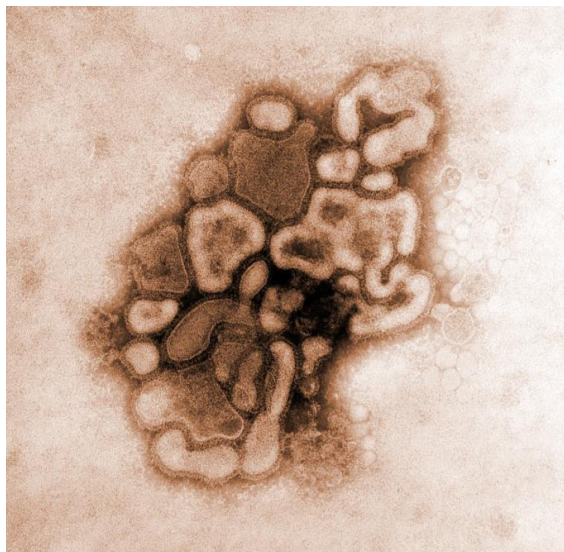
References

- Laiton-Donato K, Franco-Muñoz C, Álvarez-Díaz DA, Ruiz-Moreno HA, Usme-Ciro JA, Prada DA, et al. Characterization of the emerging B.1.621 variant of interest of SARS-CoV-2. *Infect Genet Evol.* 2021;95:105038. <https://doi.org/10.1016/j.meegid.2021.105038>
- Uriu K, Kimura I, Shirakawa K, Takaori-Kondo A, Nakada TA, Kaneda A, et al.; Genotype to Phenotype Japan (G2P-Japan) Consortium. Neutralization of the SARS-CoV-2 Mu variant by convalescent and vaccine serum. *N Engl J Med.* 2021;385:2397–9. <https://doi.org/10.1056/NEJMc2114706>
- Wang Y, Ma Y, Xu Y, Liu J, Li X, Chen Y, et al. Resistance of SARS-CoV-2 Omicron variant to convalescent and CoronaVac vaccine plasma. *Emerg Microbes Infect.* 2022; 11:424–7. <https://doi.org/10.1080/22221751.2022.2027219>
- Chen M, Zhang XE. Construction and applications of SARS-CoV-2 pseudoviruses: a mini review. *Int J Biol Sci.* 2021;17:1574–80. <https://doi.org/10.7150/ijbs.59184>
- Kollmann TR. Variation between populations in the innate immune response to vaccine adjuvants. *Front Immunol.* 2013;4:81. <https://doi.org/10.3389/fimmu.2013.00081>
- Madhi SA, Izu A, Pollard AJ. ChAdOx1 nCoV-19 vaccine efficacy against the B.1.351 variant. [Reply]. *N Engl J Med.* 2021;385:571–2. <https://doi.org/10.1056/NEJMc2110093>
- Vignier N, Bérout V, Bonnave N, Peugny S, Ballet M, Jacoud E, et al. Breakthrough infections of SARS-CoV-2 Gamma variant in fully vaccinated gold miners, French Guiana, 2021. *Emerg Infect Dis.* 2021;27:2673–6. <https://doi.org/10.3201/eid2710.211427>
- Cai Y, Zhang J, Xiao T, Lavine CL, Rawson S, Peng H, et al. Structural basis for enhanced infectivity and immune evasion of SARS-CoV-2 variants. *Science.* 2021;373:642–8. <https://doi.org/10.1126/science.abi9745>
- McCallum M, Czudnochowski N, Rosen LE, Zepeda SK, Bowen JE, Walls AC, et al. Structural basis of SARS-CoV-2 Omicron immune evasion and receptor engagement. *Science.* 2022;375:864–8. <https://doi.org/10.1126/1659science.abn8652>
- Hossain MJ, Rabaan AA, Mutair AA, Alhumaid S, Emran TB, Saikumar G, et al. Strategies to tackle SARS-CoV-2 Mu, a newly classified variant of interest likely to resist currently available COVID-19 vaccines. *Hum Vaccin Immunother.* 2022;18:2027197. <https://doi.org/10.1080/21645515.2022.2027197>
- Uriu K, Cardenas P, Munoz E, Barragan V, Kosugi Y, Shirakawa K, et al. Characterization of the immune resistance of SARS-CoV-2 Mu variant and the robust immunity induced by Mu infection. *J Infect Dis.* 2022;jiac053.
- Gutierrez B, Marquez S, Prado-Vivar B, Becerra-Wong M, Guadalupe JJ, Candido DDS, et al. Genomic epidemiology of SARS-CoV-2 transmission lineages in Ecuador. *Virus Evol.* 2021;7:veab051. 13. Collie S, Champion J, Moultrie H, Bekker LG, Gray G. Effectiveness of BNT162b2 vaccine against Omicron variant in South Africa. *N Engl J Med.* 2022;386:494–6. <https://doi.org/10.1056/NEJMc2119270>
- Khoury DS, Cromer D, Reynaldi A, Schlub TE, Wheatley AK, Juno JA, et al. Neutralizing antibody levels are highly predictive of immune protection from symptomatic SARS-CoV-2 infection. *Nat Med.* 2021;27:1205–11. <https://doi.org/10.1038/s41591-021-01377-8>

Address for correspondence: Jan Felix Drexler, Institute of Virology, Charitéplatz 1, 10117 Berlin, Germany; email: felix.drexler@charite.de

EID Podcast

Farmer Infected with Avian-Like Swine Influenza



Viruses are constantly mutating, and with those mutations can come shifts in their abilities to infect different hosts. Sometimes these mutations allow a virus to “jump” from one species to another, such as an avian influenza virus adapting to pigs.

Zoonotic transmission can have catastrophic effects on global and environmental health. Researchers document and study these events, prepare for them, and if possible, minimize the risk for zoonotic transmission in the first place.

In this EID podcast, Dr. Kristien Van Reeth, a professor of virology at Ghent University in Belgium, tells the events of how an avian-like influenza virus infected a pig farmer in the Netherlands.

Visit our website to listen:
<https://go.usa.gov/xHgBx>

**EMERGING
INFECTIOUS DISEASES®**

Effectiveness of Naturally Acquired and Vaccine-Induced Immune Responses to SARS-CoV-2 Mu Variant

Appendix

Participant Recruitment and Sampling

The study was approved by the Ethics Committee of the Universidad Industrial de Santander (protocol 4110) and by the Ethics Committee of the Charité Universitätsmedizin-Berlin (Protocol EA2/031/22). All patients provided written informed consent. To control whether the vaccinated persons also had been naturally infected, subjects were followed up and did not report clinical symptoms or direct contact with persons who tested positive until sampling. In addition to that, all persons vaccinated with the spike-based vaccines from Pfizer and AstraZeneca tested negative for antibodies against the SARS-CoV-2 N protein, suggesting lack of natural infection and consistent with recording of clinical symptoms.

50% Plaque Reduction Neutralization Tests

We used a parental SARS-CoV-2 B.1 lineage strain (Pango version 3.1.17) sampled in January 2020 (Munich/ChVir929/2020 strain, GISAID accession: EPI_ISL_406862), containing one mutation (D614G) in the spike-encoding gene only compared to the SARS-CoV-2 reference sequence used for vaccine production (Isolate Wuhan-Hu-1 GenBank accession number: NC045512). We used the following SARS-CoV-2 variants: Alpha (ChVir21652/2020, GISAID accession: EPI_ISL_802995), Beta (ChVir22131/2021, GISAID accession: EPI_ISL_862149), Gamma (NH-RIVM_10915/2021, GISAID accession: EPI_ISL_943045), Delta (454236/2021, GISAID accession: EPI_ISL_4566914), Mu (H3/2021, GISAID accession: EPI_ISL_6665693) and Omicron (hCoV-19/Netherlands/NH-RIVM-71076/2021, GISAID accession: EPI_ISL_6841611.2; Pango lineage: BA.1.17.2). A total of 60 plaque-forming units were incubated with serum dilutions of 1:40, 1:120, 1:360, and 1:1080 for 1 h, and afterwards added onto a monolayer containing 1.8×10^5 Vero E6 cells per well in a 12-well plate. After 1 h of

incubation, an overlay containing DMEM with 1% FCS and 2% Avicell was added, and cells were further incubated for 3 d for Mu and 2 d for the other variants. The overlay medium was removed, and cells were fixated with 6% paraformaldehyde and stained with crystal violet. PRNT50 endpoint titers were calculated using a logistic regression function in GraphPad prism6 (www.graphpad.com).

Antigenic Cartography

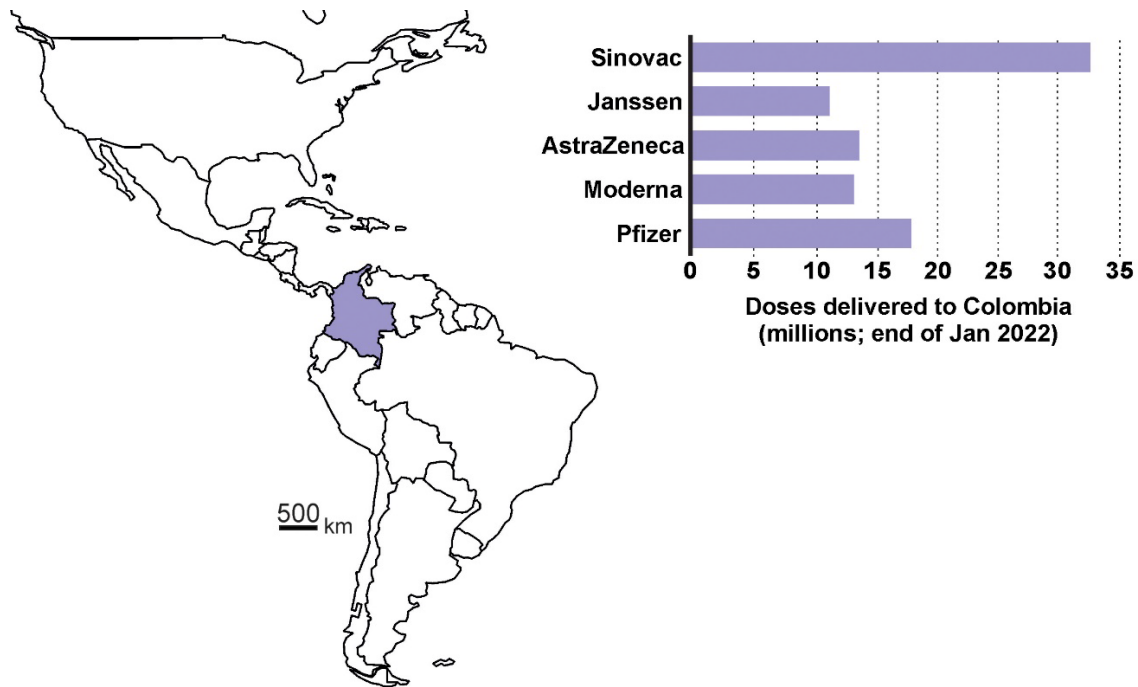
Antigenic cartography was done using the R package Racmacs (on <https://acorg.github.io/Racmacs>) as described elsewhere (S.H. Wilks; unpub. data, <https://www.biorxiv.org/content/10.1101/2022.01.28.477987v1>). Comparative neutralization of SARS-CoV-2 variants by serum samples from persons fully immunized with the different vaccines (BioNTech-Pfizer BNT162b2, AstraZeneca AZD1222, and CoronaVac).

Appendix Table. List of samples and reciprocal PRNT₅₀ endpoint titers of serum samples for vaccinated persons*†

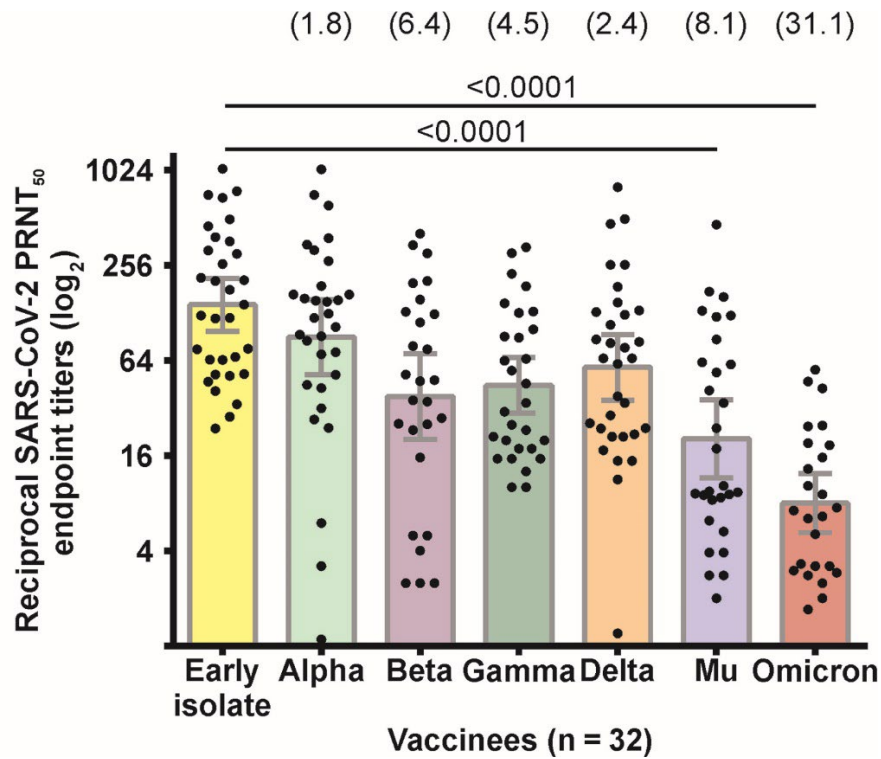
Sample ID	Early isolate	Mu	Alpha	Beta	Gamma	Delta	Omicron	D after 2nd dose	Age	Vaccine
AZ2	204	41	154	79	64	77	13	129	64	AstraZeneca
AZ3	453	123	381	305	306	470	25	173	69	AstraZeneca
AZ4	75	3	91	16	20	24	6	173	72	AstraZeneca
AZ5	76	9	104	3	13	29	2	146	61	AstraZeneca
AZ6	34	4	45	3	23	15	0	165	72	AstraZeneca
AZ9	179	35	189	75	128	84	10	142	61	AstraZeneca
AZ10	319	9	153	47	55	26	8	142	63	AstraZeneca
PF1	119	9	85	1	18	38	3	113	65	BioNTech
PF2	28	3	43	35	15	17	3	170	42	BioNTech
PF3	262	62	158	130	101	149	19	110	46	BioNTech
PF4	754	121	715	204	226	187	43	110	27	BioNTech
PF5	501	87	320	48	91	259	9	121	29	BioNTech
PF6	123	10	119	52	15	11	3	80	46	BioNTech
PF7	214	9	70	5	0	125	3	88	54	BioNTech
PF8	207	18	167	28	25	66	3	66	35	BioNTech
PF9	715	10	273	0	46	108	2	65	49	BioNTech
PF10	1043	132	1,036	343	333	799	47	89	53	BioNTech
SVN1	51	54	72	36	66	83	0	89	56	CoronaVac
SVN2	47	9	24	23	21	22	0	33	23	CoronaVac
SVN3	41	6	1	1	18	1	0	31	30	CoronaVac
SVN4	118	61	151	111	89	87	25	27	28	CoronaVac
SVN7	363	162	347	407	188	259	56	37	27	CoronaVac
SVN8	303	5	93	26	30	61	5	37	54	CoronaVac
SVN9	53	0	32	3	15	35	0	41	25	CoronaVac
SVN10	65	4	27	0	10	66	1	46	26	CoronaVac
SVN12	52	8	52	1	20	21	0	105	60	CoronaVac
SVN13	387	24	126	126	35	130	7	90	57	CoronaVac
SVN15	145	175	168	197	147	133	19	131	92	CoronaVac
SVN16	67	2	6	25	10	21	3	90	54	CoronaVac
SVN17	24	1	1	5	0	15	3	97	56	CoronaVac
SVN18	65	0	3	4	0	24	7	81	52	CoronaVac
SVN20	686	464	612	155	131	503	16	46	28	CoronaVac

*AZ, AstraZeneca; PF, Pfizer; SVN, Sinovac

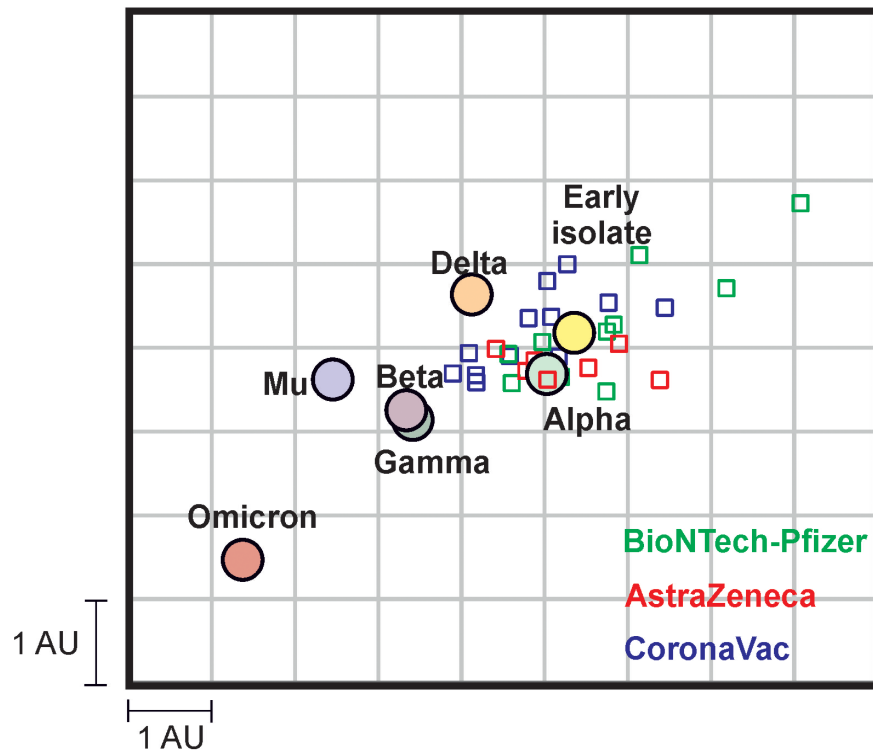
†Because we followed the Colombian vaccination program, it was not possible to collect samples the same time after completing the recommended vaccination scheme and the subjects' age were variable.



Appendix Figure 1. Vaccines doses delivered to Colombia as of January 2022 (<https://www.minsalud.gov.co>).



Appendix Figure 2. Comparative neutralization of SARS-CoV-2 variants by serum samples from persons fully immunized with the different vaccines (BioNTech-Pfizer BNT162b2, AstraZeneca AZD1222, and CoronaVac). Each point represents 50% plaque reduction neutralization test endpoint titers of 1 tested serum using different SARS-CoV-2 variants; the bars indicate the geometric mean titers, and the grey error bars represent 95% CI. Statistical significance was determined by the Wilcoxon matched signed-rank test and p-values are indicated on top. For clarity of presentation, only significant values between the early isolate and the Mu and the Omicron variants are shown.



Appendix Figure 3. Antigenic cartography of SARS-CoV-2 variants based on serum samples used in Figure 2 A–C. Each square corresponds to a serum sample tested. The colored circles indicate the tested SARS-CoV-2 variants. One grid square (1 antigenic unit) corresponds to a 2-fold serum dilution in the PRNT₅₀ assay. To decrease uncertainty in the antigenic cartography, PRNT₅₀ all endpoint titers <10 were considered as exactly <10. Antigenic mapping was not done for naturally infected persons because we could not rule out infection with multiple SARS-CoV-2 variants leading to heterogeneous antibody responses preventing a meaningful antigenic map.