

Ehrlichia muris in *Ixodes cookei* Ticks, Northeastern United States, 2016–2017

Technical Appendix

Technical Appendix Table 1. Primers and probes used in this study

Target gene	Application	Type	Sequences (5'→3')	Tm (C)	Reference
16S	Tick species PCR and confirmation	Forward	TGCTGTAGTATTTTGACTATACAAAGG	55	This paper
		Reverse	ATCCTAATCCAACATCGAGGTC		
ITS	<i>Ixodes scapularis</i> identification	Forward	TGCGTTTTCTTTGAGCAAATGCACGAG	60	This paper
		Reverse	GTACGGGATTTCCACAAACGGTATCCA		
ITS	<i>Ixodes pacificus</i> identification	Forward	FAM-TGCGCTTAACCAGTCTCTCTCTACGA-BHQ	60	This paper
		Reverse	CTCGGAGCAAGTACGGAGGTAG		
ITS	<i>Amblyomma americanum</i> identification	Forward	TTCCACAAAACGGTCGCCATC	60	This paper
		Reverse	Cy5-CTGAGCCAAGTCCTTCTACCCGGTTTG-BHQ		
ITS	<i>Dermacentor variabilis</i> identification	Forward	CGACCCGCCGAGGAAGG	60	(1)
		Reverse	CGTTTCTCGCAGCAGTTCGG		
gltA	<i>E. muris</i> PCR and confirmation	Forward	FAM-CCCCTGGCCCGCTACGTGT-BHQ	50	(2)
		Reverse	CTGAAGATTCTTTGCGAGGAGCGG		
groEL	<i>E. muris</i> PCR and confirmation	Forward	GCGTCAGCTCGGCCAAC	50	(2)
		Reverse	FAM-AGAAGGGCGTGCCCCGAAAGCGG-BHQ		
P13	EMLA detection	Forward	TACAGATTTCTCAAGAAATATA (outer)	60	This paper
		Reverse	ATGCAATGTTTTCTAATTCTAC (outer)		
gIpQ	<i>Borrelia miyamotoi</i> detection	Forward	TGACCAAACCCATTAATCTTG (inner)	60	(3)
		Reverse	GGATCCATTGGCTCTTGCTA (outer)		
ospA	<i>Borrelia burgdorferi</i> <i>Sensu Lato</i> detection	Forward	AAGGGATTCAAAGAATTGGATG (inner)	60	This paper
		Reverse	CCACCAACCTTTAAGACAGCA (outer)		
Tubulin	<i>Babesia</i> detection	Forward	CCACCAACCTTTAAGACAGCA (inner)	60	(4)
		Reverse	TACCTAATCTTCTCAAGAGATTGAGTTG		
MSP-2	<i>Anaplasma</i> detection	Forward	ATGATGATACTGCCAACAATAAGAG	60	(4)
		Reverse	Cy5-ATATTGATAAAAGAGTCAGTGTGATCCGTATGAGTTA		
gIpQ	<i>Borrelia miyamotoi</i> detection	Forward	GGGTT-BHQ	60	(3)
		Reverse	GACATAGTTCTAACAAAGGACAATATTCC		
ospA	<i>Borrelia burgdorferi</i> <i>Sensu Lato</i> detection	Forward	TCCGTTTTCTCTAGCTCGATTGG	60	This paper
		Reverse	HEX-TGCACGACCCAGAAATTGACACAACCACAA-BHQ		
Tubulin	<i>Babesia</i> detection	Forward	ATAGGTCTAATATTAGCCTTAATAGCAT	60	(4)
		Reverse	AGATCGTACTTGCCGTCTT		
MSP-2	<i>Anaplasma</i> detection	Forward	FAM-aagc+Aaa+Atgtt+Agc+Agccttga-BHQ (LNA)	60	(4)
		Reverse	GATTTGGAACCTGGCACCATG		
MSP-2	<i>Anaplasma</i> detection	Forward	AATGACCCTTAGCCCAATTATTTCC	60	(4)
		Reverse	FAM-ATCTGGCCCATACGGTGAATTGTTTCGC-BHQ		
MSP-2	<i>Anaplasma</i> detection	Forward	ATGGAAGGTAGTGTGGTTATGGTATT	60	(4)
		Reverse	TTGGTCTTGAAGCGCTCGTA		
MSP-2	<i>Anaplasma</i> detection	Forward	HEX-TGGTGCCAGGGTTGAGCTTGAGATTG-BHQ	60	(4)
		Reverse	HEX-TGGTGCCAGGGTTGAGCTTGAGATTG-BHQ		

*We use species-specific taqman qPCR identified *Amblyomma americanum* (243), *Dermacentor variabilis* (271), *Ixodes pacificus* (215), and *Ixodes scapularis* (7800). A fragment of 16S mtDNA was used to identify EMLA-positive *Ixodes scapularis* ticks (2), *Ixodes angustus* (14), *Ixodes cookei* (22), *Ixodes ricinus* (5), *Ixodes spinipalpis* (19) and *Dermacentor occidentalis* (3). The rest 114 ticks were identified by morphological characters or marked as unidentifiable because of poor sample conditions.

Technical Appendix Table 2. DNA sequences used in this study

Organism	Gene	Sample or strain	GenBank accession no.*
<i>Ixodes scapularis</i>	16s rRNA	20170522–362	MG242324
<i>Ixodes scapularis</i>	16s rRNA	20170605–176	MG242325
<i>Ixodes cookei</i>	16s rRNA	20170731–52	MG242326
<i>Ixodes cookei</i>	16s rRNA	20170807–147	MG242327
<i>Ixodes cookei</i>	16s rRNA	20170918–13	MG242328
<i>E. muris</i>	gltA	20170522–362	MG242314
<i>E. muris</i>	gltA	20170605–176	MG242315
<i>E. muris</i>	gltA	20170731–52	MG242316
<i>E. muris</i>	gltA	20170807–147	MG242317
<i>E. muris</i>	gltA	20170918–13	MG242318
<i>E. muris</i>	groEL	20170522–362	MG242319
<i>E. muris</i>	groEL	20170605–176	MG242320
<i>E. muris</i>	groEL	20170731–52	MG242321
<i>E. muris</i>	groEL	20170807–147	MG242322
<i>E. muris</i>	groEL	20170918–13	MG242323
<i>E. muris</i>	gltA	WI22	HQ660494
<i>E. muris</i>	groEL	WI22	HQ660492
<i>E. muris</i>	gltA	WI975	HQ660497
<i>E. muris</i>	groEL	WI975	HQ660493
<i>E. muris</i>	gltA	EmCRT	LANU00000000
<i>E. muris</i>	groEL	EmCRT	LANU00000000
<i>E. muris</i>	gltA	AS145	CP006917
<i>E. muris</i>	groEL	AS145	CP006917
<i>E. muris</i>	groEL	Est7979	KU535864
<i>E. muris</i>	groEL	Nov-lp300	GU358687
<i>E. muris</i>	groEL	RUS/Nov14–2526-lpr	KX980049
<i>E. muris</i>	groEL	Est1709	KU535861
<i>E. muris</i>	groEL	IRK1/Warsaw	KF312362
<i>Ehrlichia spp.</i>	gltA	HF	NZ_CP007474
<i>Ehrlichia spp.</i>	groEL	HF	NZ_CP007474
<i>E. ovata</i>	groEL	Shizuoka	DQ672553
<i>E. chaffeensis</i>	gltA	Jax	CP007475
<i>E. chaffeensis</i>	groEL	Jax	CP007475
<i>E. chaffeensis</i>	gltA	Arkansas	CP000236
<i>E. chaffeensis</i>	groEL	Arkansas	CP000236
<i>E. canis</i>	gltA	Jake	NC_007354
<i>E. canis</i>	groEL	Jake	NC_007354
<i>E. ewingii</i>	gltA	Panola Mountain	DQ365879
<i>E. ewingii</i>	groEL		AF195273
<i>E. ruminantium</i>	gltA	Welgevonden	NC_005295
<i>E. ruminantium</i>	groEL	Welgevonden	NC_005295

*GenBank accession numbers MG242314 to MG242328 are new sequences in this study.

Technical Appendix Table 3. EMLA positive ticks in this study

Tick ID#	Tick species	Tick stage	Source	Location
20170522–362	<i>Ixodes scapularis</i>	Adult	Human	Laporte, MN 56461
20170605–176	<i>Ixodes scapularis</i>	Adult	Human	Eleva, WI 54738
20170731–52	<i>Ixodes cookei</i>	Adult	Human	Holden, ME 04429
20170807–147	<i>Ixodes cookei</i>	Adult	Human	Salamanca, NY 14779
20170918–13	<i>Ixodes cookei</i>	Adult	Human	Littleton, ME 04730

References

1. Shone SM, Dillon HJ, Hom SS, Delgado N. A novel real-time PCR assay for the speciation of medically important ticks. *Vector Borne Zoonotic Dis.* 2006;6:152–60. [PubMed](#)
<http://dx.doi.org/10.1089/vbz.2006.6.152>
2. Telford Iii SR, Goethert HK, Cunningham JA. Prevalence of *Ehrlichia muris* in Wisconsin Deer Ticks Collected During the Mid 1990s. *Open Microbiol J.* 2011;5:18–20. [PubMed](#)
<http://dx.doi.org/10.2174/1874285801105010018>
3. Krause PJ, Schwab J, Narasimhan S, Brancato J, Xu G, Rich SM. Hard Tick Relapsing Fever Caused by *Borrelia miyamotoi* in a Child. *Pediatr Infect Dis J.* 2016;35:1352–4. [PubMed](#)
<http://dx.doi.org/10.1097/INF.0000000000001330>
4. Xu G, Mather TN, Hollingsworth CS, Rich SM. Passive Surveillance of *Ixodes scapularis* (Say), Their Biting Activity, and Associated Pathogens in Massachusetts. *Vector Borne Zoonotic Dis.* 2016;16:520–7. [PubMed](#) <http://dx.doi.org/10.1089/vbz.2015.1912>