
Highly Pathogenic Avian Influenza A(H5N6) in Domestic Cats, South Korea

KyungHyun Lee,¹ Eun-Kyoung Lee,¹
HyunKyoung Lee, Gyeong-Beom Heo, Yu-Na Lee,
Ji-Youl Jung, You-chan Bae, ByungJae So,
Youn-Jeong Lee, Eun-Jin Choi

In December 2016, highly pathogenic avian influenza (HPAI) infection with systemic pathologic lesions was found in cats in South Korea. Genetic analyses indicated that the feline isolates were similar to HPAI H5N6 viruses isolated in chicken farms nearby. This finding highlights the need for monitoring of domestic mammals during HPAI outbreaks.

Highly pathogenic avian influenza (HPAI) H5N6 has spread across Asia, Europe, and Africa. Since a novel influenza A(H5N6) virus emerged in China in late 2013 (1), H5N6 viruses have been subsequently reported in Southeast Asia. In China, HPAI A(H5N6) virus caused the earliest reported human infection in 2014 and became one of the dominant subtypes in poultry farms and live poultry markets (2). These viruses caused a potential threat to other mammals, including pigs and cats (3,4). We report H5N6 virus infection in cats during 2016–17 HPAI outbreaks in domestic poultry in South Korea (5).

The Study

The 2016–17 winter season saw epidemics of HPAI A(H5N6) in domestic poultry and wild birds in South Korea (5). At the end of December 2016, three carcasses of cats were submitted from areas near H5N6 virus-infected chicken farms in Pocheon. The cats had shown sudden clinical signs of salivation, lethargy, convulsion, and bloody discharge around the mouth and jaws and died within 4 days after illness onset despite antimicrobial drug treatment. After necropsy, we processed representative tissues for histopathology and immunohistochemistry (online Technical Appendix, <https://wwwnc.cdc.gov/EID/article/24/12/18-0290-Techapp1.pdf>). The necropsy findings included bloody nasal discharge (Figure 1, panel A), severe pulmonary congestion and edema, and white-colored foci in the liver (Figure 1, panel B). The lungs were red and yellow in color and incompletely collapsed and had accumulated a small amount of frothy fluid. The spleen was

enlarged 2-fold. The pancreas showed spotty hemorrhage and white pinpoint foci.

Histopathologic examination revealed severe lesions in brain, lungs, and liver in the examined cats. We observed necrosis and loss of neurons, lymphocytic perivascular cuffing, and gliosis (Figure 1, panel C) in the cerebellum and cerebrum, and especially severe necrosis in the hippocampus. The lungs showed marked congestion, edema, hemorrhage, and severe interstitial pneumonia (Figure 1, panel D), and thrombus in the alveolar capillaries. The liver showed severe necrotic foci and hepatitis. We observed influenza viral antigen in neurons (Figure 1, panel E), glial cells, and alveolar macrophages (Figure 1, panel F). Table 1 describes histopathologic lesions and Table 2 immunohistochemical reactivity.

We recognized H5N6 virus infection in a domestic male cat (cat 1) and juvenile outdoor cats (cats 2 and 3). We observed necrotic lesions and influenza viral antigens in multiple visceral organs, suggesting that the virus caused systemic infection. It seems likely that the neurotropism of H5N6 virus was a key factor contributing to the sudden death of these cats. The results of this study are consistent with those of other studies of HPAI pathogenicity in experimentally infected dogs (6,7).

The histopathologic findings and the localization of H5N6 virus antigen to the lungs and liver, but not to the brain, in cats have been reported (8). In this case, we observed meningoencephalitis. Moreover, the 3 cats showed neurologic symptoms such as salivation and convulsion, which may be related to necrosis and loss of neurons. The severity of the lesions was consistent with the number of cells that reacted with influenza viral antigen. A few studies reported that H9 and H10 influenza viruses were nephrotropic in chickens with low pathogenicity (9,10) and that HPAI H5 virus causes acute renal lesions in mammals and primates, including humans (11). The results of our study suggest that the HPAI H5N6 virus affects cats differently than do other HPAI viruses; therefore, further studies are needed to experimentally infect cats with other HPAI H5 subtypes, including the isolate from this study, for complete clarification.

Previous studies have shown that avian viruses preferentially recognize α -2,3 linkage (SA α 2,3Gal) and bind to type II alveolar cells, which are abundant in the lower

Author affiliation: Animal and Plant Quarantine Agency, Gimcheon, South Korea

DOI: <https://doi.org/10.3201/eid2412.180290>

¹These authors contributed equally to this article.

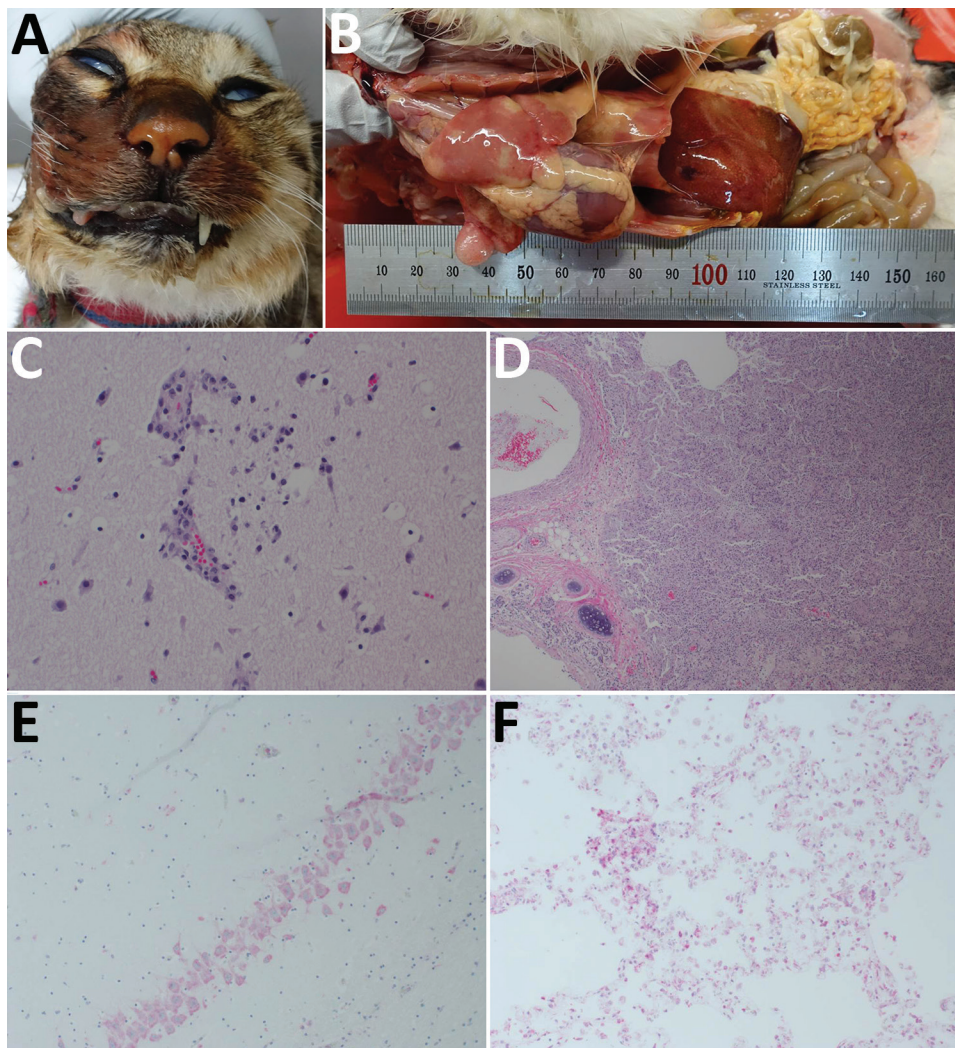


Figure 1. Gross, microscopic, and immunohistochemical (IHC) findings in 3 cats with highly pathogenic avian influenza A(H5N6) virus infection, South Korea. A) Bloody nasal discharge. B) Severe congestion and edema in the lungs and white-colored foci in the liver. C) Gliosis in the brain. Hematoxylin and eosin stain; original magnification $\times 100$. D) Interstitial pneumonia with degenerated pneumocytes. Hematoxylin and eosin; original magnification $\times 40$. E) Influenza viral antigens in neurons. IHC testing; original magnification $\times 100$. F) Influenza viral antigens in alveolar macrophages. IHC testing; original magnification $\times 100$.

respiratory tract of mammals (12,13). These findings are consistent with our observations of severe pneumonia with lung edema in the infected cats.

RNA samples extracted from organs of the cats were positive for influenza H5 and N6 subtypes by reverse transcription PCR. We selected 2 HPAI H5N6 viruses:

Table 1. Pathologic lesions in various tissues of 3 cats diagnosed with highly pathogenic avian influenza, South Korea

Tissue	Lesions	Cat 1	Cat 2	Cat 3
Brain	Neuronal necrosis	Severe	Severe	Mild
	Meningoencephalitis	Moderate	Moderate	Minimal
	Gliosis	Moderate	Moderate	Minimal
Trachea	Lymphocytic tracheitis	Minimal	Minimal	Minimal
Lung	Interstitial pneumonia	Severe	Severe	Severe
	Pneumocytic necrosis	Severe	Moderate	Severe
Heart	Myocytic necrosis	Minimal	Minimal	Minimal
	Lymphocytic myocarditis	Minimal	Minimal	Minimal
Spleen	Lymphocytic necrosis	Minimal	Minimal	Minimal
	Lymphocytic depletion	Mild	Minimal	Minimal
Pancreas	Acinar epithelial necrosis	Minimal	None	None
Intestine	Enterocytic necrosis	Minimal	None	None
	Enteritis	Minimal	None	None
Liver	Hepatic necrosis	Severe	Severe	Severe
Kidney	Tubular necrosis	None	None	None
	Nephritis	None	None	None

Table 2. Reactivity to influenza viral nucleoprotein in various tissues of 3 cats with highly pathogenic avian influenza, South Korea

Tissue	Cells	Reactivity		
		Cat 1	Cat 2	Cat 3
Brain	Neurons	Numerous	Numerous	Moderate
	Glial cells	Moderate	Numerous	Minimal
	Ependymal cells	Numerous	Numerous	Minimal
Trachea	Epithelial cells	Minimal	Minimal	Minimal
Lung	Macrophages	Numerous	Numerous	Numerous
	Vascular endothelial cells	Numerous	Numerous	Numerous
Heart	Myocytes	Minimal	Minimal	Minimal
	Epicardial cells	Minimal	Minimal	Minimal
Spleen	Ellipsoid capillary endothelium	Minimal	Minimal	Minimal
	Macrophages and necrotic debris	Moderate	Minimal	Minimal
Pancreas	Acinar epithelium	Minimal	None	None
Intestine	Crypt epithelium	Minimal	None	None
	Mesenteric ganglial neurons	Minimal	None	None
Liver	Kupffer cells and necrotic debris	Numerous	Numerous	Numerous
Kidney	Tubule epithelium	Minimal	Minimal	Unknown
	Glomeruli	Minimal	None	None

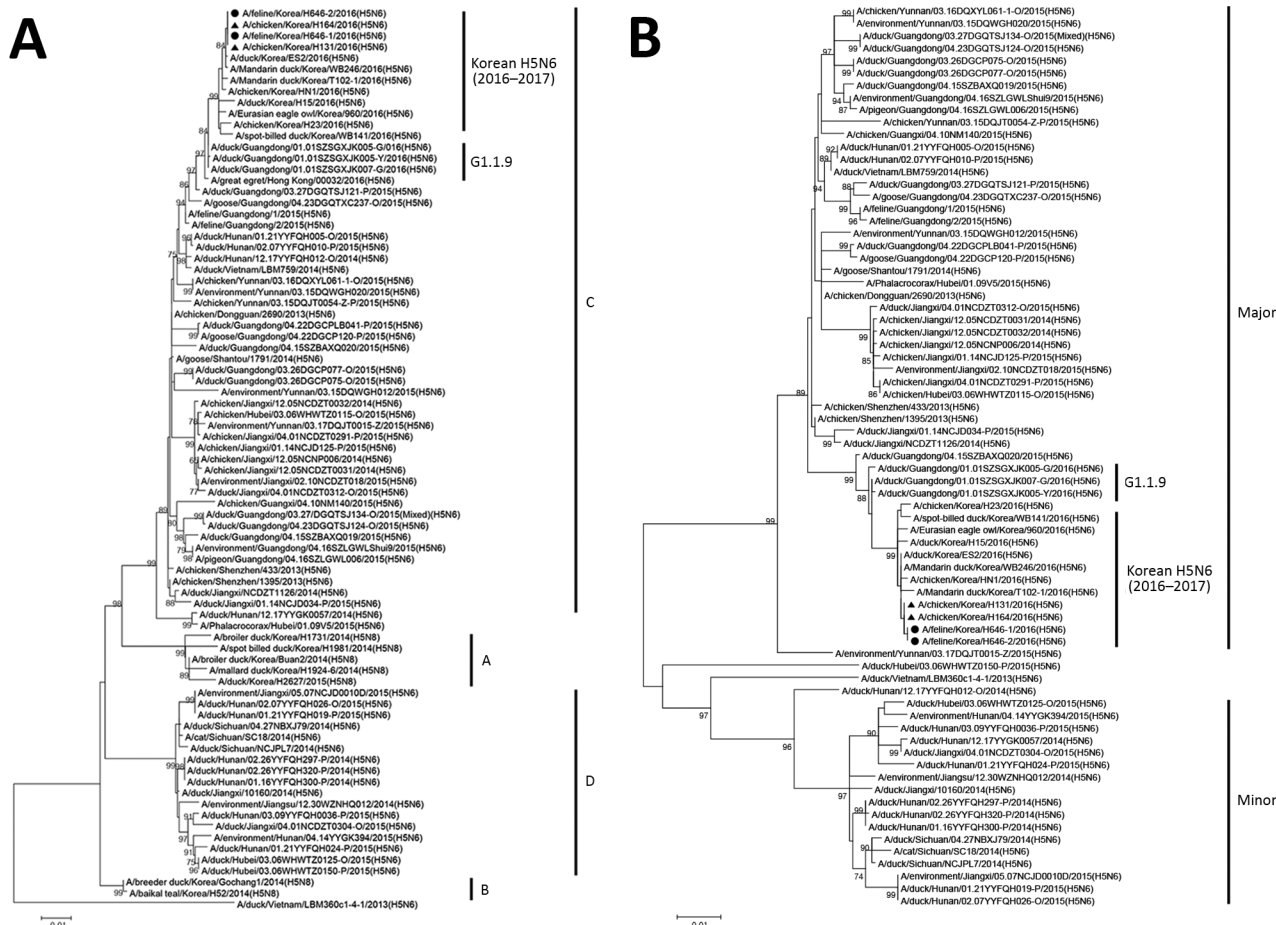


Figure 2. Maximum-likelihood phylogenetic tree of the hemagglutinin (A) and neuraminidase (B) gene segments for highly pathogenic avian influenza A(H5N6) viruses from cats, South Korea, and comparison viruses. Black circles indicate isolates from cats and triangles indicate chicken isolates from this study. Virus sequences from the GISAID EpiFlu database (<http://platform.gisaid.org>) and GenBank were used for each phylogenetic comparison. The genetic subclades are annotated to the right of the tree. The genetic clusters major, minor, and G1.1.9, were designated according to the criteria of Bi et al. (2). The number at each branch indicates a bootstrap value. Scale bars indicate nucleotide substitutions per site.

A/feline/Korea/H646-1/2016(H5N6) from the domestic male cat and A/feline/Korea/H646-2/2016(H5N6) from 1 juvenile outdoor cat. We performed virus isolation, sequencing, and phylogenetic analysis as described (online Technical Appendix). Phylogenetic analyses showed that the H5 genes of the cat isolates belonged to clade 2.3.4.4.C and were very closely related to H5N6 viruses detected in poultry in areas within a radius of 1 km of the cats' location (Figure 2, panel A). In a previous study, clade 2.3.4.4.C H5N6 viruses from the 2016–17 epidemic in South Korea were divided into 5 distinct genogroups (C-1 to C-5) (5). The feline isolates showed high similarity with H5N6 viruses in genogroup C-4, which was detected in domestic poultry nearby during 2016–17 HPAI A(H5N6) outbreaks (Figure 2; online Technical Appendix Figure).

Epidemiologic studies show that the cats might be infected by feeding on or by contact with infected wild birds, although the virus was not isolated from wild birds around this area. The affected domestic cat lived in a house near a small stream where migratory birds were observed and a wide main road. Across the main road, H5N6 virus-affected chicken farms were located within 1 km. In previous reports, cats and tigers were naturally infected by feeding on infected bird carcasses (8,14). In China, H5N6 virus infection in cats has been reported in regions such as Sichuan and Guangdong Provinces (3).

We compared each gene of the feline and chicken H5N6 isolates (online Technical Appendix Table). The hemagglutinin (HA) genes of the viruses contained multiple basic amino acid residues at the HA cleavage site (PLRERRRKR). The amino acid residues on the receptor binding sites of the HA gene of H5N6 viruses were Q226 and G228 (H3 numbering), indicating an avian-like (α 2,3-SA) receptor-binding preference. T160A mutation in the HA gene suggested a possible increased viral affinity for human-like (α 2,6-SA) receptor binding, shown in feline isolates. The neuraminidase genes of feline isolates also had 11 aa deletions at positions 59–69, which were often observed in avian influenza virus lineages adapted to poultry and may increase the virulence to mammals (2). We did not observe amino acid substitution at position E627K of the polybasic 2 gene in the feline isolates.

Conclusions

Our results demonstrate that cats can be directly infected by HPAI H5N6 virus. Cats are companion animals and may act as a vector for influenza transmission to humans. Despite the low probability of H5N6 virus infection in companion animals, avian influenza surveillance will be needed for mammals, including cats, during H5N6 outbreaks.

Acknowledgments

We thank the Animal and Plant Quarantine Agency (APQA); Ministry of Agriculture, Food and Rural Affairs (MAFRA); and regional office for animal disease control for their efforts to control HPAI.

This study was supported by a grant from APQA, MAFRA, Republic of Korea (project code nos. N-1543069-2015-99-01 and B-1543418-2018-19-01).

About the Author

Dr. KyungHyun Lee is a veterinary researcher at the Animal and Plant Quarantine Agency, South Korea. Her research interests include pathology and emerging infectious diseases in animals.

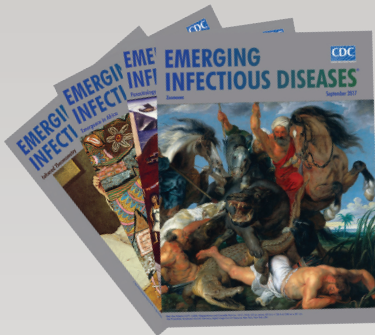
Dr. Eun-Kyoung Lee is a veterinary researcher at the Animal and Plant Quarantine Agency. Her research interests include the molecular epidemiology and surveillance of avian influenza.

References

1. Qi X, Cui L, Yu H, Ge Y, Tang F. Whole-genome sequence of a reassortant H5N6 avian influenza virus isolated from a live poultry market in China, 2013. *Genome Announc*. 2014;2:e00706-14. <http://dx.doi.org/10.1128/genomeA.00706-14>
2. Bi Y, Chen Q, Wang Q, Chen J, Jin T, Wong G, et al. Genesis, evolution and prevalence of H5N6 avian influenza viruses in China. *Cell Host Microbe*. 2016;20:810–21. [10.1016/j.chom.2016.10.022](http://dx.doi.org/10.1016/j.chom.2016.10.022) <http://dx.doi.org/10.1016/j.chom.2016.10.022>
3. Yu Z, Gao X, Wang T, Li Y, Li Y, Xu Y, et al. Fatal H5N6 avian influenza virus infection in a domestic cat and wild birds in China. *Sci Rep*. 2015;5:10704. <http://dx.doi.org/10.1038/srep10704>
4. Li X, Fu Y, Yang J, Guo J, He J, Guo J, et al. Genetic and biological characterization of two novel reassortant H5N6 swine influenza viruses in mice and chickens. *Infect Genet Evol*. 2015;36:462–6. <http://dx.doi.org/10.1016/j.meegid.2015.08.017>
5. Lee EK, Song BM, Lee YN, Heo GB, Bae YC, Joh SJ, et al. Multiple novel H5N6 highly pathogenic avian influenza viruses, South Korea, 2016. *Infect Genet Evol*. 2017;51:21–3. <http://dx.doi.org/10.1016/j.meegid.2017.03.005>
6. Son K, Kim YK, Oem JK, Jheong WH, Sleeman JM, Jeong J. Experimental infection of highly pathogenic avian influenza viruses, clade 2.3.4.4 H5N6 and H5N8, in mandarin ducks from South Korea. *Transbound Emerg Dis*. 2017;65:899–903.
7. Lyoo KS, Na W, Phan LV, Yoon SW, Yeom M, Song D, et al. Experimental infection of clade 1.1.2 (H5N1), clade 2.3.2.1c (H5N1) and clade 2.3.4.4 (H5N6) highly pathogenic avian influenza viruses in dogs. *Transbound Emerg Dis*. 2017;64:1669–75. <http://dx.doi.org/10.1111/tbed.12731>
8. Songserm T, Amonsin A, Jam-on R, Sae-Heng N, Meemak N, Pariyothorn N, et al. Avian influenza H5N1 in naturally infected domestic cat. *Emerg Infect Dis*. 2006;12:681–3. <http://dx.doi.org/10.3201/eid1204.051396>
9. Swayne DE, Alexander DJ. Confirmation of nephrotropism and nephropathogenicity of three low-pathogenic chicken-origin influenza viruses for chickens. *Avian Pathol*. 1994;23:345–52. <http://dx.doi.org/10.1080/03079459408419002>
10. Bonfante F, Fusaro A, Zanardello C, Patrono LV, De Nardi R, Maniero S, et al. Lethal nephrotropism of an H10N1 avian influenza virus stands out as an atypical pathotype. *Vet Microbiol*. 2014;173:189–200. <http://dx.doi.org/10.1016/j.vetmic.2014.07.023>
11. Cardona CJ, Xing Z, Sandrock CE, Davis CE. Avian influenza in birds and mammals. *Comp Immunol Microbiol*

- Infect Dis. 2009;32:255–73. <http://dx.doi.org/10.1016/j.cimid.2008.01.001>
12. van Riel D, Munster VJ, de Wit E, Rimmelzwaan GF, Fouchier RA, Osterhaus AD, et al. H5N1 virus attachment to lower respiratory tract. *Science*. 2006;312:399. <http://dx.doi.org/10.1126/science.1125548>
 13. Shinya K, Ebina M, Yamada S, Ono M, Kasai N, Kawaoka Y. Influenza virus receptors in the human airway. *Nature*. 2006;440:435–6. <http://dx.doi.org/10.1038/440435a>
 14. Keawcharoen J, Oraveerakul K, Kuiken T, Fouchier RA, Amonsin A, Payungporn S, et al. Avian influenza H5N1 in tigers and leopards. *Emerg Infect Dis*. 2004;10:2189–91. <http://dx.doi.org/10.3201/eid1012.040759>

Address for correspondence: Youn-Jeong Lee and Eun-Jin Choi, Animal and Plant Quarantine Agency, hyuksin8ro 177 Gimcheon, Gimcheon 39660, South Korea; email: leeyj700@korea.kr; choiej@korea.kr



**EMERGING
INFECTIOUS DISEASES®**

September 2017

Zoonoses

- Bioinformatic Analyses of Whole-Genome Sequence Data in a Public Health Laboratory
- Convergence of Humans, Bats, Trees, and Culture in Nipah Virus Transmission, Bangladesh
- Processes Underlying Rabies Virus Incursions across US–Canada Border as Revealed by Whole-Genome Phylogeography
- Real-Time Whole-Genome Sequencing for Surveillance of *Listeria monocytogenes*, France
- Role of Food Insecurity in Outbreak of Anthrax Infections among Humans and Hippopotamuses Living in a Game Reserve Area, Rural Zambia
- Molecular Antimicrobial Resistance Surveillance for *Neisseria gonorrhoeae*, Northern Territory, Australia
- Estimated Annual Numbers of Foodborne Pathogen–Associated Illnesses, Hospitalizations, and Deaths, France, 2008–2013
- Epidemiology of *Salmonella enterica* Serotype Dublin Infections among Humans, United States, 1968–2013
- Prevalence of *Yersinia enterocolitica* Bioserotype 3/O:3 among Children with Diarrhea, China, 2010–2015
- Risk for Low Pathogenicity Avian Influenza Virus on Poultry Farms, the Netherlands, 2007–2013
- Patterns of Human Plague in Uganda, 2008–2016
- Serologic Evidence for Influenza C and D Virus among Ruminants and Camelids, Africa, 1991–2015
- Norovirus in Bottled Water Associated with Gastroenteritis Outbreak, Spain, 2016
- Group A Rotavirus Associated with Encephalitis in Red Fox
- Protective Effect of Val₁₂₉-PrP against Bovine Spongiform Encephalopathy but not Variant Creutzfeldt-Jakob Disease
- Imported Infections with *Mansonella perstans* Nematodes, Italy
- Genetic Diversity of Highly Pathogenic Avian Influenza A(H5N8/H5N5) Viruses in Italy, 2016–17
- Microcephaly Caused by Lymphocytic Choriomeningitis Virus
- Influenza A(H3N2) Virus in Swine at Agricultural Fairs and Transmission to Humans, Michigan and Ohio, USA, 2016
- Use of Blood Donor Screening to Monitor Prevalence of HIV and Hepatitis B and C Viruses, South Africa
- Emergence of Plasmid-Mediated Fosfomycin-Resistance Genes among *Escherichia coli* Isolates, France
- Determination of the Ferret Enteric Coronavirus Genome in Laboratory Ferrets
- Myocarditis Caused by Human Parechovirus in Adult
- Cost of Nosocomial Outbreak Caused by NDM-1–Containing *Klebsiella pneumoniae* in the Netherlands, 2015–2016
- Evaluation of 5 Commercially Available Zika Virus Immunoassays
- Epidemiology of *Neisseria gonorrhoeae* Gyrase A Genotype, Los Angeles, California, USA
- Conveyance Contact Investigation for Imported Middle East Respiratory Syndrome Cases, United States, May 2014
- Possible Role of Fish as Transport Hosts for *Dracunculus* spp. Larvae

To revisit the September 2017 issue, go to:

<https://wwwnc.cdc.gov/eid/articles/issue/23/9/table-of-contents>

Highly Pathogenic Avian Influenza A(H5N6) in Domestic Cats, South Korea

Technical Appendix

Materials and Methods

Virus Isolation and Sequencing

Viral RNA extracted from organs of dead cats using a Patho Gene-spin DNA/RNA extraction kit (iNtRON Biotechnology, Sungnam, Korea) according to the manufacturer's instructions were positive for H5 and N6 subtype by RT-PCR (1,2). For virus isolation, the specimens were inoculated into embryonated specific pathogen-free chicken eggs. After incubation at 37°C, allantoic fluid from inoculated eggs was harvested and tested using the hemagglutination assay. All 8 RNA genomic segments were amplified using segment specific primers and directly sequenced (3). The nucleotide sequences of the 8 RNA genomic segments of 2 H5N6 isolates, A/feline/Korea/H646-1/2016(H5N6) and A/feline/Korea/H646-2/2016(H5N6), were deposited to GISAID (accession nos. EPI1123314-21 and EPI1123332-39). In addition, the nucleotide sequences of 2 H5N6 viruses [A/chicken/H131/2016(H5N6) and A/chicken/H164/2016(H5N6)] isolated from chicken farms in Pocheon-gun of Gyeonggi-do province in December 2016, respectively, were deposited to GISAID (EPI1123348-55 and EPI1123340-47).

Phylogenetic Analysis

The nucleotide sequences for phylogenetic analysis were downloaded from GISAID (<http://www.gisaid.org>) or GenBank. All nucleotides optimization and multiple sequence alignment were performed by CLC main workbench software; version 6.8.2 (CLC bio, Aarhus, Denmark). The maximum-likelihood (ML) trees were conducted by MEGA version 6.0 (www.megasoftware.net) using complete coding nucleotide sequences of 8 segments. The nucleotide substitution models (HA, NP, and NA, Hasegawa-Kishino-Yano with Gamma-distribution; PB2, PB1, PA, General Time Reversible with Gamma-distribution; M, Kimura 2-

parameter with Gamma-distribution; NS, Tamura 3-parameter with Gamma-distribution) of each segment for ML trees were computed by MEGA 6 software package (4,5). The test of phylogeny was estimated by bootstrap analysis with 1000 replication. Phylogenetic trees constructed for the PB2, PB1, PA, HA, NP, NA, M and NS segments are shown in Figure 2 and online Technical Appendix Figure.

Histopathologic and Immunohistochemical Test

Collected tissues (brain, heart, lung, spleen, kidney, liver, pancreas, intestine) were fixed for 24 hours in 10% buffered neutral formalin and processed for paraffin embedding. Paraffin-embedded sections were cut (4 µm), dewaxed, and stained with hematoxylin and eosin. Duplicate sections were immunohistochemically analyzed to determine the distribution of influenza virus antigens in individual tissues. Briefly, sections were reacted with a monoclonal antibody against influenza A virus nucleoprotein (MCA-400; AbD Serotec, Dusseldorf, Germany), followed by a biotinylated goat anti-mouse IgG as secondary antibody and an avidin-biotin complex system (Ventana Medical Systems, Tucson, AZ, USA). The RedMap Kit (Ventana Medical Systems) served as a chromogen substrate. Slides prepared with serial sections of the same tissues were incubated with PBS instead of antibody as a negative control.

References

1. Munch M, Nielsen LP, Handberg KJ, Jørgensen PH. Detection and subtyping (H5 and H7) of avian type A influenza virus by reverse transcription-PCR and PCR-ELISA. *Arch Virol.* 2001;146:87–97. [PubMed http://dx.doi.org/10.1007/s007050170193](http://dx.doi.org/10.1007/s007050170193)
2. Fereidouni SR, Starick E, Grund C, Globig A, Mettenleiter TC, Beer M, et al. Rapid molecular subtyping by reverse transcription polymerase chain reaction of the neuraminidase gene of avian influenza A viruses. *Vet Microbiol.* 2009;135:253–60. [PubMed http://dx.doi.org/10.1016/j.vetmic.2008.09.077](http://dx.doi.org/10.1016/j.vetmic.2008.09.077)
3. Hoffmann E, Stech J, Guan Y, Webster RG, Perez DR. Universal primer set for the full-length amplification of all influenza A viruses. *Arch Virol.* 2001;146:2275–89. [PubMed http://dx.doi.org/10.1007/s007050170002](http://dx.doi.org/10.1007/s007050170002)
4. Nei M, Kumar S. *Molecular evolution and phylogenetics.* New York: Oxford University Press; 2000.

5. Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Mol Biol Evol.* 2013;30:2725–9. [PubMed](#)
<http://dx.doi.org/10.1093/molbev/mst197>
6. Bi Y, Chen Q, Wang Q, Chen J, Jin T, Wong G, et al. Genesis, evolution and prevalence of H5N6 avian influenza viruses in China. *Cell Host Microbe.* 2016;20:810–21. [PubMed](#)
<http://dx.doi.org/10.1016/j.chom.2016.10.022>
7. Lee EK, Song BM, Lee YN, Heo GB, Bae YC, Joh SJ, et al. Multiple novel H5N6 highly pathogenic avian influenza viruses, South Korea, 2016. *Infect Genet Evol.* 2017;51:21–3. [PubMed](#)
<http://dx.doi.org/10.1016/j.meegid.2017.03.005>

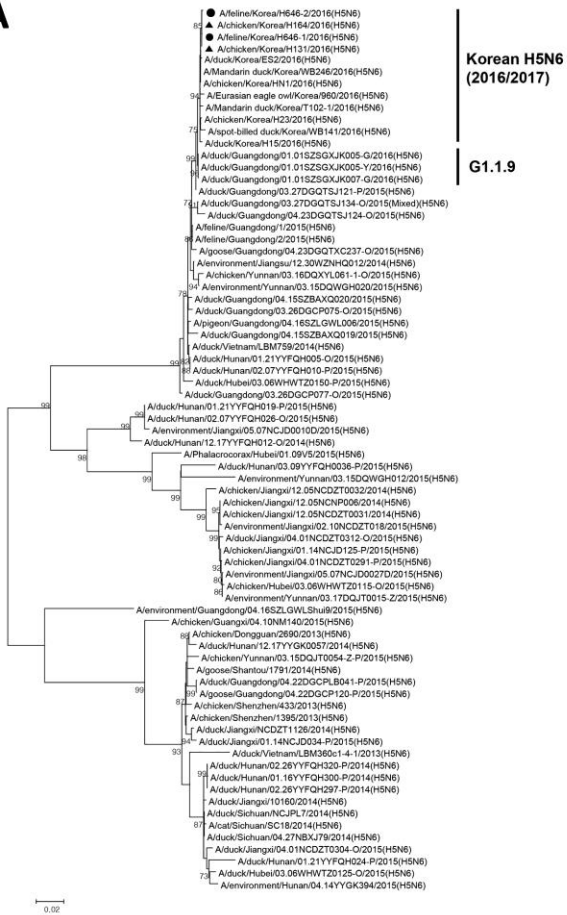
Technical Appendix Table. Comparison of the molecular characteristics of the H5N6 highly pathogenic avian influenza viruses in South Korea*

Strain	Host	Genogroup	Antigenic site of HA			HA cleavage site	NA deletion		M2	NS	NS deletion		PB2		PB1-F2		Length
			156	222	224	321–330 (333–339)†	59–69	26	31	42	80–84	218–230	627	701	66		
			(160)†	(226)†	(228)†												
A/chicken/Korea/H131/2016 (H5N6)	Poultry	C4	A	Q	G	PLRERRRKR	Yes	L	S	S	Yes	No	E	D	N	90	
A/chicken/Korea/H164/2016 (H5N6)	Poultry	C4	A	Q	G	PLRERRRKR	Yes	L	S	S	Yes	No	E	D	N	90	
A/feline/Korea/H646– 1/2016(H5N6)	Feline	C4	A	Q	G	PLRERRRKR	Yes	L	S	S	Yes	No	E	D	N	90	
A/feline/Korea/H646– 1/2016(H5N6)	Feline	C4	A	Q	G	PLRERRRKR	Yes	L	S	S	Yes	No	E	D	N	90	
Avian influenza viruses	–	–	T	Q	G	–	Yes	L	S/N	S/A	Yes	No/ deletion	E	D	N	–	
Human influenza viruses	–	–	A	I/L	S	–	Yes	I	S/N	S	No	No	K	N	S	–	

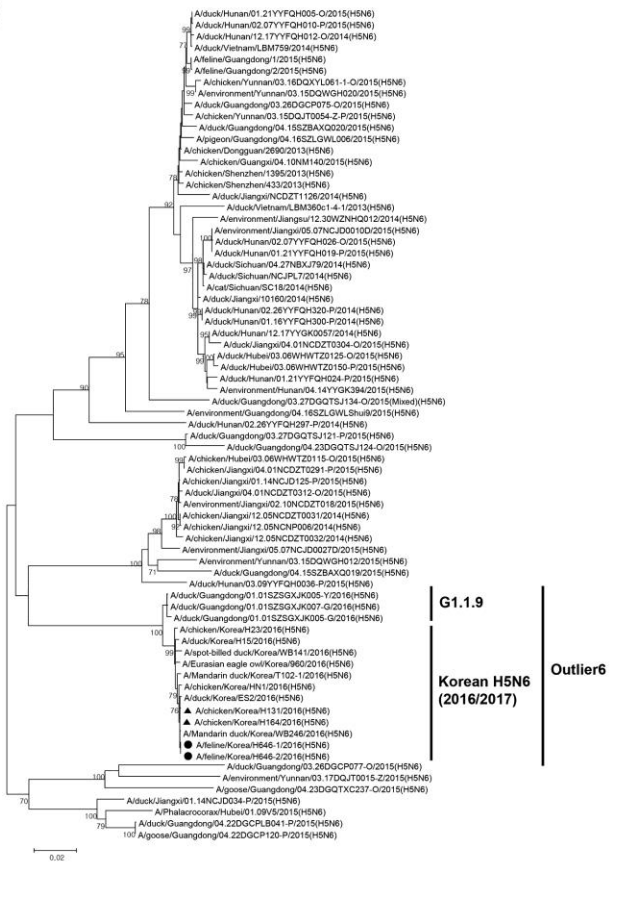
*–, Not determined.

†H3 numbering.

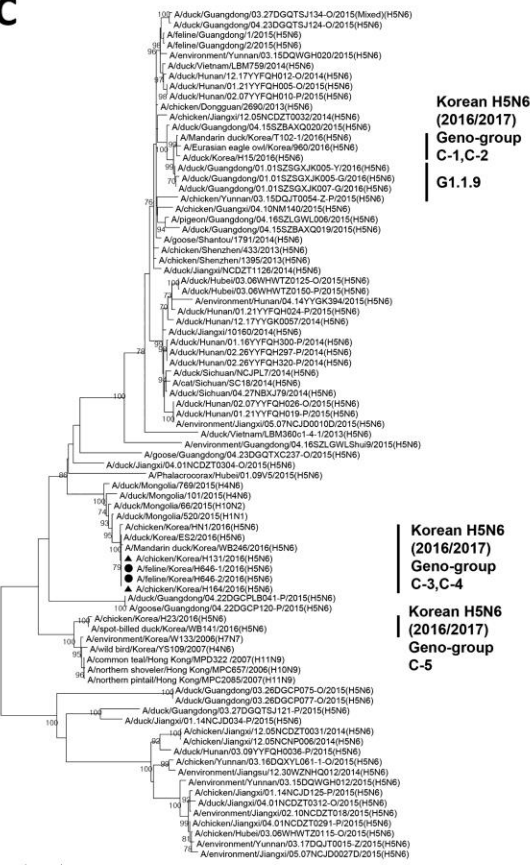
A



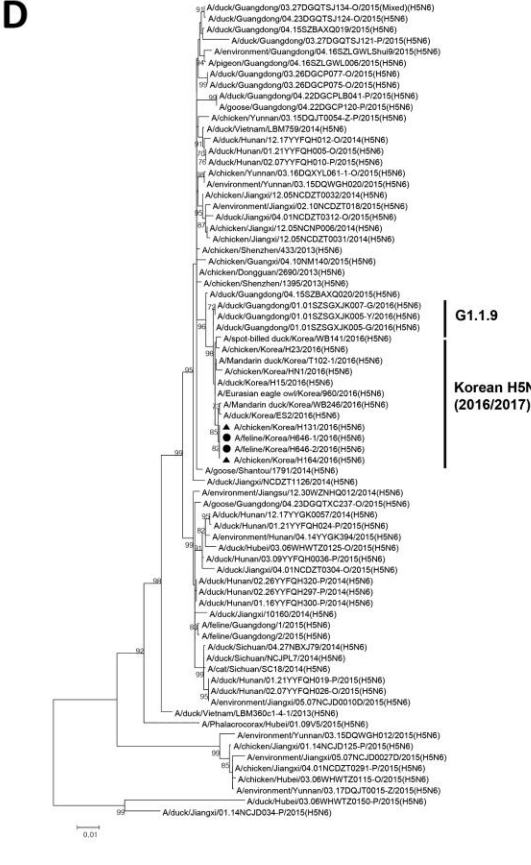
B

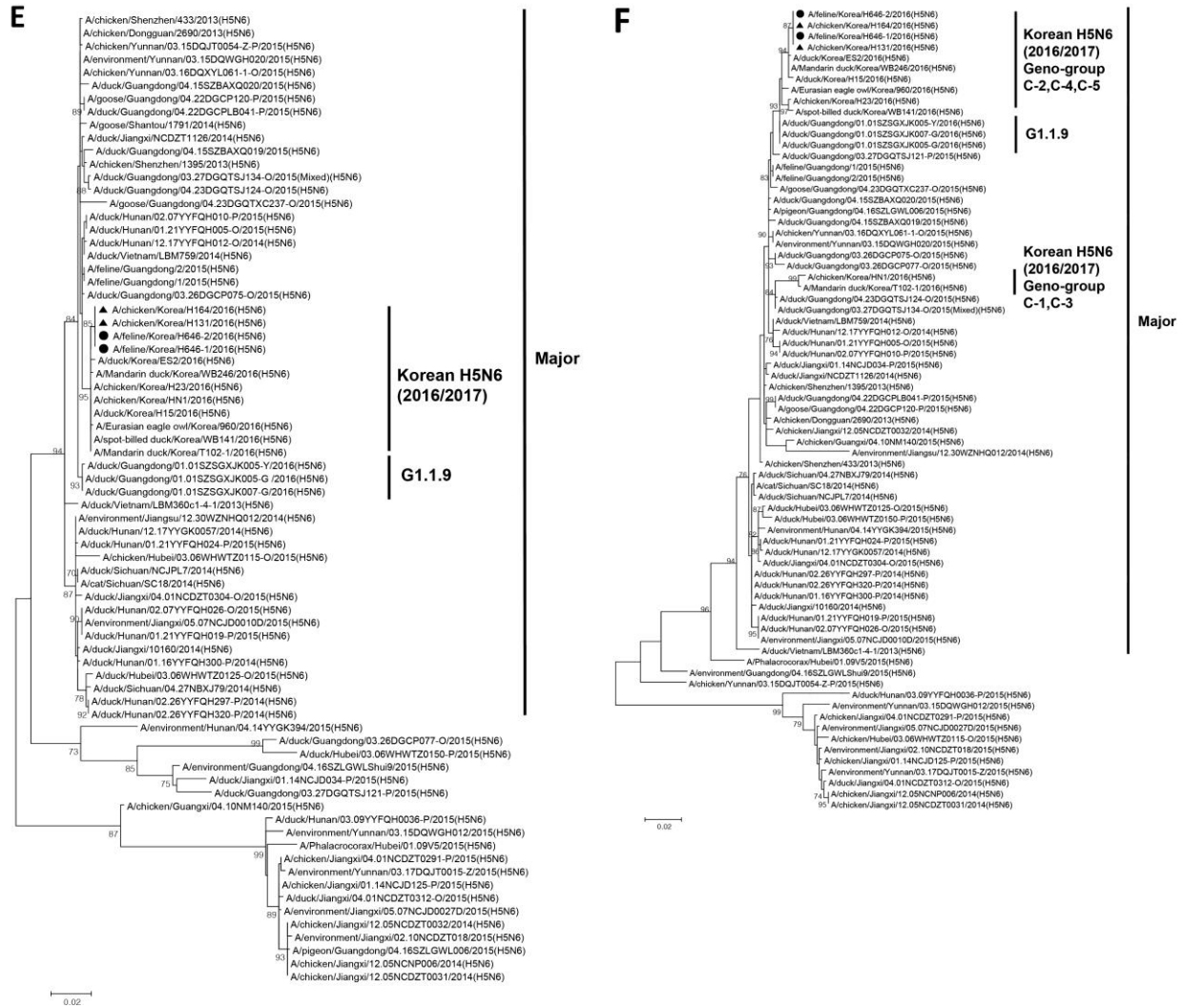


C



D





Technical Appendix Figure (following pages). Maximum-likelihood phylogenetic tree for internal gene segments for the feline H5N6 viruses and references. A) Polymerase basic-2 (PB2); B) polymerase basic-1 (PB1); C) polymerase acidic (PA); D) nucleoprotein (NP); E) matrix (M); F) nonstructural (NS). The highly pathogenic and low pathogenicity influenza virus sequences from the GISAID EpiFlu database (<http://platform.gisaid.org/epi3/frontend#326742>) were used for each phylogenetic comparison. The genetic subclades are annotated to the right of the tree. The genetic clusters; Major, Minor and G1.1.9, were designated according to the criteria of Bi et al. (6) and the genogroups of Korean H5N6 HPAI viruses (C-1 to C-5) are annotated to the right of the tree of PA and NS (7). At each branch, the number indicates a bootstrap value (>70%). Black circles indicate feline isolates in this study and triangles indicate chicken isolates. Scale bar indicates nucleotide substitutions per site.