

10. McMichael AJ, Woodruff RE, Hales S. Climate change and human health: present and future risks. *Lancet*. 2006;367:859–69. DOI: 10.1016/S0140-6736(06)68079-3

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European Perspective of 2-Person Rule for Biosafety Level 4 Laboratories

To the Editor: Recently, the directors of Biosafety Level 4 (BSL-4) laboratories in the United States published their views of the requirement of having ≥ 2 persons present at all times while biological work is undertaken in a BSL-4 laboratory (*1*). They concluded that safety and security would be better assured in some situations by video monitoring systems rather than by the presence of a fellow scientist. As members of the European Network of Biosafety Level-4 laboratories (Euronet-P4) who have developed guidelines in this area (*2–4*), we discussed the article during a recent network meeting. Biosafety and biosecurity are the major concerns for all involved in BSL-4 activities, and we support the authors' initiative and broadly agree with their position. The consensus among European BSL-4 experts is that, in the interest of safety, standard practice should be for all laboratories to perform a risk assessment before any activity is undertaken. This preliminary assessment is the best way to determine procedures to be used, including whether 2 persons should work together as part of

laboratory procedure. A 2-person rule is inappropriate simply because the best approach is not to have inflexible rules that are not objectively assessed according to laboratory-specific circumstances.

Surveillance video monitoring and data storing have their place in protecting laboratory facilities from unauthorized access and theft of materials, but their effectiveness for ensuring proper handling of pathogens is quite limited. Finally, we agree with the authors that both biosafety and biosecurity must be founded on careful selection and monitoring of staff, without which even the most sophisticated of control systems would fail.

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References

1. Le Duc JW, Anderson K, Bloom ME, Carrion R Jr, Feldmann H, Fitch JP, et al. Potential impact of a 2-person security rule on Biosafety Level 4 laboratory workers. *Emerg Infect Dis* [cited 2009 Jul 28]. Available from <http://www.cdc.gov/EID/content/15/7/e1.htm> DOI: 10.3201/eid1507.081523
2. The European Network of P4 Laboratories 2005–2007 [Euronet–P4]. Brussels: European Commission; 2008 [cited 2009 Jul 28]. Available from http://ec.europa.eu/health/ph_projects/2003/action2/action2_2003_19_en.htm and www.euronetp4.eu
3. Ippolito G, Nisii C, Capobianchi MR. Networking for infectious-disease emergencies in Europe. *Nat Rev Microbiol*. 2008;6:564. DOI: 10.1038/nrmicro1896-c1
4. Nisii C, Castilletti C, Di Caro A, Capobianchi MR, Brown D, Lloyd G, et al. The European Network of P4 Laboratories: enhancing European preparedness for new health threats. *Clin Microbiol Infect*. 2009 May 28 [Epub head of print].

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Multidrug-Resistant *Mycobacterium tuberculosis* Strain from Equatorial Guinea Detected in Spain

To the Editor: Eleven years of molecular epidemiologic data allowed the Spanish Multidrug-resistant Tuberculosis (MDR TB) Surveillance Network to identify a specific MDR *Mycobacterium tuberculosis* strain that had been imported into Spain from Equatorial Guinea (*1*). Our study brings to light the potential dissemination of this strain (named MDR-TBEG) in Equatorial Guinea, a country where little is known about the extent and features of TB or MDR TB. It also highlights that MDR strains can spread across continents, and thus MDR TB's emergence in any country becomes a global problem.

Ten MDR *M. tuberculosis* isolates obtained from 10 patients from Equatorial

torial Guinea were detected in Spain during 2000 through 2008. Evidence of clonality was found within the 10 isolates because all exhibited identical genetic profiles defined by different molecular epidemiology methods (2,3) and mutations involved in drug resistance (Figure). Notably, none of the remaining 504 MDR isolates in the Spanish database matched SIT177, a spoligotype belonging to the Latin American–Mediterranean 9 (LAM9) subfamily (4).

The data routinely collected for all cases of MDR TB have been previously described (1). All 10 patients in the study were from Equatorial Guinea, a small African country on the Gulf of Guinea with a population of $\approx 500,000$, an MDR TB rate $>2.0\%$ (5) of all combined (new and previously treated) TB cases, and an estimated adult HIV prevalence rate of 3.2% (www.who.int/globalatlas/predefinedReports/EFS2008/full/EFS2008_GQ.pdf). The MDR TB isolates were collected within a 9-year period (online Technical Appendix, available from www.cdc.gov/EID/content/15/11/1858-Techapp.pdf): 1 in 2000, 2 in 2001, 3 in 2003, 1 in 2004, 2 in 2007, and 1 in 2008. According to their hospitals of origin, the patients were geo-

graphically dispersed in 6 different Spanish cities. We found that the interval between the patients' arrival in Spain to the initiation of anti-TB treatment was <3 months in 6 patients, 3 of whom were clinically ill at the time of arrival. Seven patients were adult men, 2 were adult women, and 1 was an 8-year-old girl. The patients' mean age was 30 years (range 8–54 years). Three patients were seropositive and 4 were seronegative for HIV infection (the HIV status of 3 patients was unknown). Data on prior anti-TB treatment was available for 7 case-patients, of whom only 1 had a history of antecedent TB chemotherapy. Altogether, 3 patients died before completing treatment, including 2 patients affected by miliary TB, 1 of whom was HIV-coinfected. The third patient who died was a student without a known history of immunosuppression or previous TB who had lived for 2 years in Spain. We could not establish any epidemiologic links between these patients during their stay in Spain.

Analysis of drug resistance genes showed that all isolates harbored the *inhA* promoter mutation $-15C \rightarrow T$ (6). Alterations in the *inhA* gene were previously reported in 80% of the isoniazid-resistant isolates from Equatorial

Guinea (5). Notably, a double mutation in the *rpoB* gene affecting codons 531 (Ser531Leu) and 561 (Ile561Val) was detected in the 10 MDR isolates. The presence of this uncommon mutation, Ile561Val, outside the rifampin resistance-determining region supports the hypothesis that the MDR isolates are clonal in origin. Furthermore, we demonstrated the absence of Ile561Val mutation in 3 drug-susceptible *M. tuberculosis* strains with an SIT177-LAM 9 spoligotype pattern, which ruled out a relationship between this spoligotype and the Ile561Val mutation.

Further analysis with phylogenetic markers assigned MDR-TBEG to the principal genetic group 2, the Euro-American lineage of *M. tuberculosis* and its West African sublineage, on the basis of polymorphisms in codons *katG*463 and *gyrA*95, the 7-bp *pks15/1* deletion, and RD174 (7,8), respectively. The analysis of the RD^{Rio} deletion confirmed that the strain belongs to the major RD^{Rio} sublineage of the LAM *M. tuberculosis* spoligotype family (9). This sublineage is a major cause of TB in Rio de Janeiro (Brazil) but has disseminated globally. Additional information on the geographic distribution of SIT177-LAM 9 was obtained from the updated International Spoligotyping Database (SITVIT2) of the Institut Pasteur de Guadeloupe. SITVIT2 (consulted on 23 July 2008) contained 57 isolates belonging to SIT177. Almost 50% ($n = 28$) came from Brazil, and 14% from Africa (Morocco, $n = 6$; Senegal, $n = 2$). The remaining isolates with known countries of origin ($n = 9$) were distributed in other unrelated countries. These data indicate that this particular spoligotype pattern is widely distributed.

We identified 1 MDR strain of *M. tuberculosis* RD^{Rio} sublineage isolated in Spain from Equatorial Guinean patients. Although the transmission of MDR-TBEG in Spain could not be conclusively ruled out, the fact that MDR TB developed in most patients

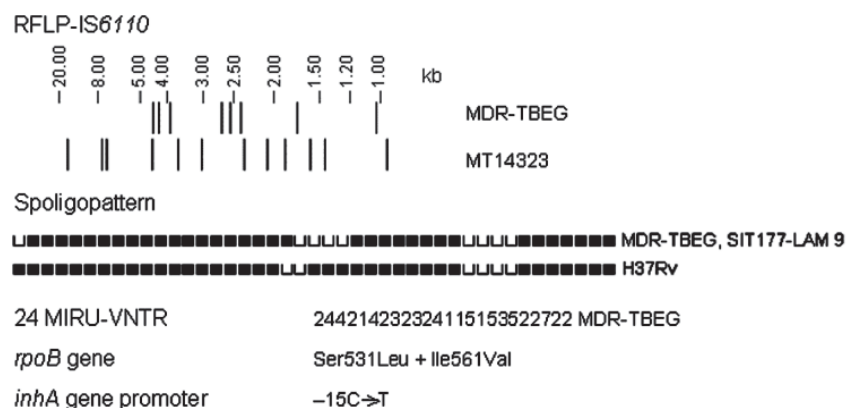


Figure. Genetic profile of the multidrug-resistant tuberculosis Equatorial Guinea strain (MDR-TBEG). RFLP, restriction fragment length polymorphism; SIT, spoligotype international type; LAM, Latin American-Mediterranean; MIRU-VNTR, mycobacterial interspersed repetitive-unit variable-number tandem-repeat. MIRU-VNTR loci order: MIRU 02, VNTR 42, VNTR 43, MIRU 04, MIRU 40, MIRU 10, MIRU 16, 1955, MIRU 20, QUB-11b, ETRA, VNTR 46, VNTR 47, VNTR 48, MIRU 23, MIRU 24, MIRU 26, MIRU 27, VNTR 49, MIRU 31, VNTR 52, QUB-26, VNTR 53, MIRU 39.

within 3 months after their arrival, as well as the spatiotemporal distribution of the MDR TB cases and its clonal origin, strongly suggest that MDR-TBEG was imported into Spain and that active transmission of this particular clone could be occurring in Equatorial Guinea. However, additional molecular and epidemiologic studies should be conducted in this sub-Saharan country to ascertain its role in recent transmission of MDR TB. Greater international efforts should be made to provide appropriate tools to resource-limited areas for fighting against MDR TB and preventing development of extensively drug-resistant TB.

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References

1. Samper S, Iglesias MJ, Rabanaque MJ, Gómez LI, Lafoz MC, Jiménez MS, et al. Systematic molecular characterization of multidrug-resistant *Mycobacterium tuberculosis* complex isolates from Spain. *J Clin Microbiol*. 2005;43:1220–7. DOI: 10.1128/JCM.43.3.1220-1227.2005
2. Kamerbeek J, Schouls L, Kolk A, van Agterveld M, van Soolingen D, Kuijper S, et al. Simultaneous detection and strain differentiation of *Mycobacterium tuberculosis* for diagnosis and epidemiology. *J Clin Microbiol*. 1997;35:907–14.
3. Supply P, Allix C, Lesjean S, Cardoso-Oelemann M, Rüsch-Gerdes S, Willery E, et al. Proposal for standardization of optimized mycobacterial interspersed repetitive unit variable-number tandem repeat typing of *Mycobacterium tuberculosis*. *J Clin Microbiol*. 2006;44:4498–510. DOI: 10.1128/JCM.01392-06
4. Brudey K, Driscoll JR, Rigouts L, Prodinger WM, Gori A, Al-Hajj SA, et al. *Mycobacterium tuberculosis* complex genetic diversity: mining the fourth international spoligotyping database (Spol-DB4) for classification, population genetics and epidemiology. *BMC Microbiol*. 2006;6:23. DOI: 10.1186/1471-2180-6-23
5. Tudó G, González J, Obama R, Rodríguez JM, Franco JR, Espasa M, et al. Study of resistance to anti-tuberculosis drugs in five districts of Equatorial Guinea: rates, risk factors, genotyping of gene mutations and molecular epidemiology. *Int J Tuberc Lung Dis*. 2004;8:15–22.
6. Herrera-León L, Molina T, Saiz P, Sáez-Nieto JA, Jiménez MS. New multiplex PCR for rapid detection of isoniazid-resistant *Mycobacterium tuberculosis* clinical isolates. *Antimicrob Agents Chemother*. 2005;49:144–7. DOI: 10.1128/AAC.49.1.144-147.2005
7. Sreevatsan S, Pan X, Stockbauer KE, Connell ND, Kreiswirth BN, Whittam TS, et al. Restricted structural gene polymorphism in the *Mycobacterium tuberculosis* complex indicates evolutionarily recent global dissemination. *Proc Natl Acad Sci U S A*. 1997;94:9869–74. DOI: 10.1073/pnas.94.18.9869
8. Gagneux S, DeRiemer K, Van T, Kato-Maeda M, de Jong BC, Narayanan S, et al. Variable host-pathogen compatibility in *Mycobacterium tuberculosis*. *Proc Natl Acad Sci U S A*. 2006;103:2869–73. Medline DOI: 10.1073/pnas.0511240103
9. Lazzarini LC, Huard RC, Boechat NL, Gomes HM, Oelemann MC, Kurepina N, et al. Discovery of a novel *Mycobacterium tuberculosis* lineage that is a major cause of tuberculosis in Rio de Janeiro, Brazil. *J Clin Microbiol*. 2007;45:3891–902. DOI: 10.1128/JCM.01394-07

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Technical Appendix

Table. Demographic and clinical characteristics of 10 patients from Equatorial Guinea infected with multiple drug resistant tuberculosis Equatorial Guinea strain*

Patient no. †	Age, y/sex‡	Date of arrival in Spain	Date of treatment initiation	City of isolation	Resistance pattern§	TB type	Sputum smear¶	History of prior TB treatment	HIV serology
1	23/F	2000 Sep	2000 Nov	Madrid	HR	Pulmonary	NA	NA	NA
2	29/F	2000 May	2000 Jul 20	Madrid	HRES	Pulmonary	Positive	No	NA
3	8/F	EG	NA	Barakaldo	HRZ	Pott's disease		NA	NA
4	54/M	2002 Nov	2003 Jan	Madrid	HRES	Miliary pattern	Positive	No	Negative
5	21/M	2001 Jun	2003 May	Las Palmas de Gran Canaria	HREta	Pulmonary	Positive	No	Negative
6	30/M	2002 Nov	2003 Oct	Alcalá de Henares	HRZEta	Miliary pattern	Positive	NA	Positive
7	27/M	1994	2004 Jun	Madrid	HR	Lymph node		No	Negative
8	49/M	EG	2008 Jan	Zaragoza	HR	Pulmonary	Positive	No	Positive
9	41/M	EG	2007 Sep	Alcalá de Henares	HRECltaEtaRfb	Pulmonary	Negative	Yes	Positive
10	24/M	By the end of 2002	2008 Mar	Galdakao	HREta	Pulmonary	Positive	No	Negative

*NA, not available; EG, resident of Equatorial Guinea referred to a Spanish hospital for diagnosis and treatment.

†All patients were born in Equatorial Guinea.

‡Age in years.

§H, isoniazid; R, rifampin; E, ethambutol; S, streptomycin; Z, pyrazinamide; Eta, ethionamide; Clta, clarithromycin; Rfb, rifabutin.

¶Sputum smear not applicable in Pott's disease or lymph node.